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Molecular profiling of foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region for genetic variability and nutritional quality

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Abstract

Agriculture in the Central Himalayan Region depends on the availability of suitable germplasm as well as natural conditions. Due to extreme weather conditions, food and nutrition security is a major issue for communities inhabiting these remote and inaccessible areas. Millets are common crops grown in these areas. Foxtail millet (*Setaria italica* (L.) P. Beauv) is an important crop and forms a considerable part of the diet in this region. The aim of the present study was to explore, collect, conserve and evaluate the untapped genetic diversity of foxtail millet at the molecular level and discover variability in their nutritional traits. A total of 30 accessions having unique traits of agronomic importance were collected and molecular profiling was performed. A total of 63 alleles were generated with an average of 2.52 alleles per locus and average expected heterozygosity of 0.37 ± 0.231 . Significant genetic variability was revealed through the genetic differentiation (Fst) and gene flow (Nm) values. Structure-based analysis divided whole germplasm into three sub-groups. Rich variability was found in nutritional traits such as dietary fibre in husked grains, carbohydrate, protein, lysine and thiamine content. The collected germplasm may be useful for developing nutritionally rich and agronomically beneficial varieties of foxtail millet and also designing strategies for utilization of unexploited genetic diversity for food and nutrition security in this and other similar agro-ecological regions.

Introduction

Since pre-historic times, food plants have been domesticated, selected, exchanged and improved by farmers in traditional ways within traditional production systems. This process has also been utilized for scientific crop improvement, which led to the Green Revolution and a significant rise in crop yields. Globally, approximately half of the increase in food production can be attributed to genetic improvement. Millions of lives depend upon the extent to which crop genetic improvement can keep pace with the growing global population, changing the climate and shrinking environmental resources (Ronald, 2011). Plant geneticists, as well as plant breeders, consider molecular marker-assisted selection (MAS) a useful additional tool in crop improvement/breeding programmes to optimize selection efficiency (Dwivedi *et al.*, 2007; Xu and Crouch, 2008). Recently, the amount of molecular genetic markers for relevant plant breeding traits has increased (Lammerts van Bueren *et al.*, 2010). Without the knowledge of linkage to genes for specific traits of interest (for the plant breeder), molecular markers can still be used to determine the genetic relatedness between two different individual plants or the genetic diversity within a gene pool (Lammerts van Bueren *et al.*, 2010). Although increasing, use of molecular markers is still modest in plant breeding. A major reason for this is the lack of appropriate markers with high selective value for many traits of interest to breeders (Tuvešson *et al.*, 2009). Particularly, those quantitative traits which depend largely on environmental factors, GEI (genotype–environment interactions), are integrated into QTL (quantitative trait loci) analysis, resulting in markers reflecting the amount and direction of the reaction of the plant to an environmental input. Such markers are valuable over a larger range of environments (Backes and Østergard, 2008). Several genes contributing to one trait can be pyramided in one genotype using linked markers. Hence, molecular profiling would be useful particularly in the utilization of untapped genetic diversity. Crop diversity is gradually shrinking in most parts of the world, with most of mankind living off a few plant species and the human diet composed mainly of a few major crops such as wheat, rice, maize and

potatoes. However, crop diversity with known traits can provide the basis for offering higher yielding and more reliable plants, which can support low-income farmers and consumers. It can also deliver security to the poor by making harvests more resilient to environmental changes.

Millet, which is one of the oldest foods known to man and possibly the first cereal grain used for domestic purposes, is grown in the Central Himalayan Region (CHR). Foxtail millet (*Setaria italica* (L.) P. Beauv) is an important annual crop of the genus *Setaria*, family Gramineae. It is a self-pollinated crop (Leonard and Martin, 1963) where cross-pollination averages about 0.04 (Li *et al.*, 1935). Foxtail millet is not just a cereal of the Old World, it is also used widely in Africa, the Americas, Australia and Eurasia (Lin *et al.*, 2012). It is a good source of dietary fibre and certain amino acids such as lysine and thiamine, which are otherwise low/deficient in cereal food. Dietary fibres found in carbohydrates aid digestion by moving food quickly through the intestines. A fibre-rich diet helps to decrease constipation and other digestive problems, lower blood cholesterol and reduce the risk of heart disease (Jones, 2001; Jones *et al.*, 2006). In human nutrition, lysine is an essential amino acid involved in the creation of collagen and absorption of calcium. It prevents cold sores (caused by the virus called herpes simplex labialis). A deficiency of lysine can lead to anaemia, bloodshot eyes and fatigue (Sahley and Birkner, 2000). Another amino acid analysed in seeds, thiamine, is an essential nutrient for normal body function (Lonsdale, 1990). Its deficiency may result in damage to the nervous system as well as to the heart and other muscles, which are the symptoms of beriberi disease (Lee, 1994). Thiamine is important due to its role as a coenzyme for reactions catalysed by enzymes; thiamine is required for mitochondrial oxidative decarboxylation (Sica, 2007), the pentose phosphate pathway and the citric acid cycle (Wooley, 2008). Due to a lack of stable and improved varieties, foxtail millet production is unstable and the market is chaotic. Therefore, there is an urgent need to develop varieties with stable and higher yield to stabilize the fluctuation of the production (Li *et al.*, 2014). In this context, evaluation of untapped genetic diversity of foxtail millet for nutritional traits is very important because this will enable the development of nutritionally rich varieties, particularly for resource-poor and malnourished regions, using the existing genetic diversity of crops.

With a relatively small genome (515 Mb), foxtail millet is a suitable plant for molecular and genetic research (Wang *et al.*, 1998). Foxtail millet (*S. italica*) exhibits numerous properties (for instance, C₄ photosynthesis) that make it an ideal model for functional genomic studies in the Panicoid grasses (Diao *et al.*, 2014). With a high-quality reference genome sequence (Bennetzen *et al.*, 2012) and a high-density haplotype map of genome variation (Jia *et al.*, 2013) and other genomic data (Kumari *et al.*, 2013; Pandey *et al.*, 2013; Suresh *et al.*, 2013; Muthamilarasan *et al.*, 2014; Zhang *et al.*, 2014; Yadav *et al.*, 2015), this species can now truly be considered as a novel model system for genetic and genomic studies in other cereal and millet crops (Doust *et al.*, 2009; Li and Brutnell, 2011; Lata *et al.*, 2013; Muthamilarasan and Prasad, 2015) to develop new varieties suitable for different regions.

Without irrigation and application of fertilizers, foxtail millet can survive and produce economic yield comparable with other crops of similar lifespan cultivated with optimal inputs. Thus, collection and nutritional evaluation of untapped genetic diversity might open some new avenues of research and breeding. Therefore, in the present study, an effort has been made to assess the extent of variability in 30 accessions of foxtail millet collected from the CHR using molecular markers and nutritional quality

traits in order to identify specific germplasm for crop improvement.

Materials and methods

Plant materials and seed quality analysis

Plant exploration and germplasm collection expeditions were conducted in the CHR of India, i.e. Uttarakhand State, which is a large geographical area known for vagaries of weather. A total of 30 accessions (including four controls used for comparison) having unique traits of agronomic importance were collected from areas with different climatic conditions as well as different altitudes in this region and were evaluated for nutritional quality and genetic variability for three consecutive years (2011–2013) at one site located at 29°24'N, 79°30'E, 1480 m a.s.l. Four accessions, IC337300, IC355776, IC418394 and IC469880, which are preferred by the farming community across the region and occupy the largest area, were used as controls for comparison. Total dietary fibre content was determined using the AOAC method 2001.03 (AOAC 2005). Total carbohydrate content was estimated by the anthrone reagent method (Morris, 1948), while protein content was determined by the folin ciocalteau reagent method (Lowry *et al.*, 1951) and fat content by AOAC method 996.01 (AOAC 1998). Estimation of lysine and thiamine amino acid content was achieved following the methods of Hurrell and Carpenter (1981) and Chen *et al.* (1999), respectively. All data are given on a dry weight basis.

Simple sequence repeat markers for molecular evaluation

The DNA was extracted and purified from seeds using mini CTAB (cetyl trimethylammonium bromide) method (Saghai-Marouf *et al.*, 1984) and a working solution of 20 ng/μl was prepared for simple sequence repeat (SSR) amplification. These accessions were profiled with genome-wide SSR markers (Zhang *et al.*, 2014). Genomic SSR markers were selected based on their high polymorphism information content (PIC) value (0.7 or more) and represented different chromosomes. All of the 25 SSR loci (Table 1) were run at one touchdown cycle, whereas earlier different annealing temperatures were used for polymerase chain reaction (PCR) amplification (Zhang *et al.*, 2014). The PCR was carried out using 15 μl reaction mixture that included 5.68 μl H₂O, 1.5 μl of 10× buffer, 1.2 μl of 25 mM MgCl₂, 0.3 μl of 10 mM dNTPs mix, 0.6 μl of 10 mM forward and reverse primer each and 5 μl of 20 ng/μl DNA. All reagents were from MBI fermentas (St. Leon-Rot, Germany). The PCR was performed using touchdown cycle: initial denaturation at 94 °C for 3 min, then ten cycles of denaturation at 94 °C for 30 s, touchdown annealing starting at 62 °C for 30 s and decreasing 0.7 °C per cycle and extension at 72 °C for 1 min followed by 35 cycles of denaturation at 94 °C for 30 s, primer annealing at temperature 55 °C for 30 s and primer extension at 72 °C for 1 min with a final extension step at 72 °C for 4 min, 3% metaphor agarose gel was used for separating the SSR amplification products and photographed using a SYNGENEG-Box Gel Documentation unit (Syngenta, Cambridge, UK).

Statistical analysis

Experiments were conducted for three consecutive years (2011–2013) in a randomized block design (RBD) with three replications. Data for each parameter were analysed for statistical

Table 1. Sequence of the simple sequence repeat (SSR) primer pairs used for molecular profiling of foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region

SSR loci	Forward sequence	Reverse sequence
SICAAS2017	F-GGCGCCATGGGTGGATGCGGAATTGTA	R-TCAGTTGTCTCCCTCGCCGCTCAGGA
SICAAS3010	F-TGCGTGTGGCGATGTGAGATGGA	R-ACCATCGGTTTCTGGCTTGCC
SICAAS3052	F-TGGAGACTTGGACACCTCTCTCTC	R-GCAAACCTGCTTTCAGCTATCGACCCA
SICAAS8028	F-GTTGCAGTTTCCATGCAGTTAG	R-TCGAGAACATCTTGTGAGCATC
SICAAS9030	F-GCAAGCAACAGTCCATGGCCGA	R-GCTGCGAGCTTGGGTGACAGA
SICAAS7038	F-GCGTTAATGGGCTGGAATTATG	R-TCATGTACGTCGCTAATCGTGA
SICAAS5049	F-GGTTTCTGCTTCTCTCCGCTCCA	R-AGGACACGGAGACCCTGCTGAAC
SICAAS5020	F-CCAACAAAACATGCCCTGGGTAAGCA	R-CGTGAGCGCCCTTTAGAGGAGGGTA
SICAAS9054	F-GGGATTGGGTGGGACGGC	R-CGCGCCACGCTCTGTAAC
SICAAS9067	F-GCTAGGCGGCTTCCCAAGGC	R-GCCGCCATCTCTCCCGTA
SICAAS7024	F-TGGTCGACGTTCTCCACTCTTA	R-CCTTGTGCGATTTATCCCTTC
SICAAS4010	F-ACTCTCCAGCCAACCAACCAACCAAG	R-TCCCTCCCCTGTTCTTCTTTAGCG
SICAAS5013	F-GCTGGTGCAGCAGATGCTACGTTT	R-GGATTTCTTTGTACCCTGCCCGC
SICAAS4024	F-TATGGGCTCGTTATGGGCCAACCA	R-ATACTGGGCTCCGGCAGCATCGAA
SICAAS4037	F-TGATGGGCTTGATTGGGACGGCTCG	R-TCCCTTTTGTGCGACGCGCTGA
SICAAS5081	F-ACCACAAATGCTGGCCTTAC	R-GTTGTACCGATCCGAGAGA
SICAAS5034	F-GTGTCTCGCTCTCCACCAAGACTC	R-ACGAGACGTCAAATCTGCAAATGGC
SICAAS6101	F-CATGGTGCCTTGCAATTAGA	R-TGCAGTTCAGTGAGACATACAAAAC
SICAAS6035	F-AATACCACACAAGCATCAGGAG	R-GGCGATGGAGTGCATTTTATTA
SICAAS9095	F-GGCCCTGGTAGTAACCTTTTA	R-AAGATGAACGATCCCAAGTACG
SICAAS3056	F-TGCGCAGGCAGCTTGTGTGTGT	R-AATCGGATTTCCCTCCGCCCT
SICAAS4061	F-CTGTTATGTGATGCTGCCAAT	R-AATGACTCCAAATGCACAGC
SICAAS4027	F-ACAACGCCACAGCCAAGCTCCCTCT	R-TGTGCGAGCCAGGCCAACACAAGGCT
SICAAS5005	F-TCTGCCATCAGGCCAAGTACAGT	R-CGGCACCCCATTTTCTTGAGCCA
SICAAS6055	F-GGCCACCCAATATAACAAAC	R-ACAAAGTGATGTGCTCCATGA

significance using two-way analysis of variance (ANOVA) to compare the means considering accession and trait as independent variables.

Alleles were scored and the presence or absence of alleles was converted to '1' and '0', respectively, for data analysis. The software program NTSYS-PC ver 2.1 (Rohlf, 2000) was used to calculate Jaccard's similarity coefficient and Unweighted Pair Group Method with Arithmetic Mean (UPGMA) cluster analysis. Nei's gene diversity statistics were calculated using POPGENE version 1.32 (Yeh *et al.*, 1999). The PIC was calculated using the formula $1 - \sum p_{ij}^2$ (Anderson *et al.*, 1993), where p_{ij} is the frequency of j th allele for i th SSR locus. Software STRUCTURE 2.3.4 (Pritchard *et al.*, 2000) was run using a burn-in of 1 00 000, a run length of 1 00 000 (admixture model) and number of populations was inferred using Structure Harvester (Earl and von Holdt, 2012) keeping K values (1–10) with five iterations at each K value.

Results

Seed quality traits are the main criteria for selection of a food crop to be grown at large scale and commercialized in any region. From the present study, it is evident that substantial variability

is available in the dietary fibre, carbohydrate and fat content of the seeds. The fibre content of seeds varied from 4.88% (in IC357343) to 5.95% (in IC338633) whereas carbohydrate content varied from 58.50% in IC337300 to 61.18% in IC406534 (Table 2). In addition, fat content ranged from 3.97% in IC337335 to 5.08% in IC337327. Protein content in seeds was found to vary from 10.03% in IC337300 to 12.29% in IC355800. Amino acid lysine in seeds ranged from 2.31 mg/g (in IC337300 and IC355794) to 2.78 mg/g (in IC383568), while thiamine content was found to vary from 5.46 µg/g (in IC337307) to 5.95 µg/g (in IC338633). Thousand grain weight were found to vary from 1.17 g (in IC338639) to 2.24 g (in IC337318) (Table 2). There is a statistically significant ($P \leq 0.05$) correlation between dietary fibre and thiamine content, indicating that accessions rich in dietary fibre will also be rich in thiamine content (Table 3), as well as carbohydrate and lysine content, indicating that accessions rich in carbohydrate will also be rich in lysine content.

A total of 25 genome-wide SSR loci were used for genotyping to investigate genetic variation (Fig. 1). The number of alleles ranged from two to four, with an average of 2.5 ± 0.77 alleles/SSR locus. Furthermore, statistical analysis using POPGENE version

Table 2. Variability in the nutritional traits and seed weight of the foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region

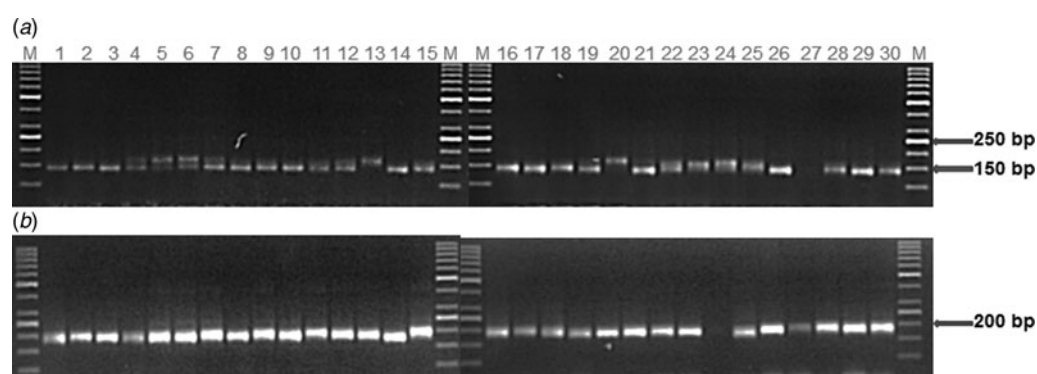
Sl. No.	IC No.	Dietary fibre (%)	Carbohydrates (%)	Fat (%)	Protein (%)	Lysine (mg/g)	Thiamine ($\mu\text{g/g}$)	1000 grain weight (g)
1	IC337300	5.51	58.5	4.21	10.03	2.31	5.51	2.12
2	IC355776	5.87	60.93	4.89	11.47	2.72	5.87	2.11
3	IC418394	5.93	60.06	4.61	10.12	2.36	5.93	2.06
4	IC469880	5.91	59.38	4.15	11.98	2.32	5.91	2.23
5	IC337303	5.33	58.69	4.79	11.03	2.56	5.53	2.21
6	IC337307	5.46	59.16	4.37	10.51	2.42	5.46	1.87
7	IC337311	5.61	60.2	4.81	11.12	2.36	5.61	2
8	IC337313	5.53	60.31	4.66	11.66	2.53	5.53	2.02
9	IC337318	5.55	60.45	4.08	11.89	2.38	5.55	2.24
10	IC337327	5.71	60.85	5.08	10.12	2.47	5.71	1.83
11	IC337335	5.89	61.11	3.97	10.72	2.62	5.89	1.89
12	IC337338	5.22	59.12	4.24	10.45	2.42	5.78	2.09
13	IC337340	5.81	59.65	4.52	10.95	2.35	5.81	1.45
14	IC338633	5.95	59.93	4.36	10.22	2.52	5.95	2.04
15	IC338639	5.88	59.32	4.68	10.8	2.48	5.88	1.17
16	IC338650	5.72	60.66	4.37	11.55	2.55	5.72	1.79
17	IC340855	5.66	58.77	4.05	12.13	2.39	5.66	1.39
18	IC340963	5.68	59.86	5.06	10.48	2.66	5.68	1.23
19	IC341376	5.82	60.72	4.77	11.14	2.65	5.82	1.96
20	IC341382	5.92	60.88	4.28	10.85	2.59	5.92	1.6
21	IC355794	5.74	60.65	4.73	12.21	2.31	5.74	1.97
22	IC355800	5.76	60.34	4.8	12.29	2.46	5.76	1.32
23	IC357343	4.88	60.08	4.46	10.83	2.56	5.69	1.78
24	IC383467	5.62	60.18	4.82	11.62	2.62	5.86	1.88
25	IC383568	5.94	60.68	4.16	11.78	2.78	5.94	1.69
26	IC383633	5.77	60.42	4.39	11.25	2.71	5.77	2.16
27	IC393056	5.72	60.12	4.57	10.43	2.69	5.72	1.73
28	IC406534	5.88	61.18	4.23	10.65	2.52	5.88	1.72
29	IC436955	5.85	59.23	4.52	10.4	2.61	5.85	1.57
30	IC469863	5.91	58.89	4.07	11.22	2.56	5.90	2.01
	Minimum	4.88	58.50	3.97	10.03	2.31	5.46	1.17
	Maximum	5.95	61.18	5.08	12.29	2.78	5.95	2.24
	Average	5.70	60.01	4.49	11.06	2.51	5.76	1.84
	STDEV	0.24	0.77	0.31	0.67	0.13	0.14	0.30
	CV%	4.24	1.29	6.92	6.06	5.32	2.50	16.32

1.32 software revealed an average effective number of alleles, Shannon's Information Index, expected heterozygosity, Nei's expected heterozygosity, average heterozygosity of 1.8 ± 0.61 , 0.6 ± 0.32 , 0.4 ± 0.23 , 0.4 ± 0.22 and 0.2 ± 0.11 , respectively (Table 4). The observed heterozygosity of the germplasm ranged from 0.000 to 0.414 with an average of 0.06 ± 0.104 , while PIC values varied from 0.064 to 0.66 with an average of 0.366. Genetic differentiation (F_{st}) among germplasm populations was

0.42 and the gene flow (N_m) was 0.34 (Table 4). Allelic data were converted into a 0/1 binary matrix based on the absence or presence of alleles, respectively, and was subjected to UPGMA cluster analysis (based on Jaccard's similarity coefficient matrix (Supplementary Table 1)) to further elucidate the genetic relatedness among different accessions (Fig. 2). The most distant accessions were IC469880 and IC469863 and most similar were IC337313 and IC337318. The Bayesian approach was followed

Table 3. Correlation matrix of foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region based on nutritional traits

	Dietary fibre (%)	Carbohydrates (%)	Fat (%)	Protein (%)	Lysine (mg/g)	Thiamine (µg/g)	1000 grain weight (g)
Dietary fibre (%)	1						
Carbohydrates (%)	0.315	1					
Fat (%)	-0.076	0.181	1				
Protein (%)	0.073	0.17	-0.089	1			
Lysine (mg/g)	0.171	0.381	0.135	-0.043	1		
Thiamine (µg/g)	0.664	0.327	-0.145	-0.023	0.315	1	
1000 grain weight (g)	-0.18	0.008	-0.194	0.003	-0.13	-0.161	1

**Fig. 1.** Gel of foxtail millet accessions with SICAAS5049 (a) and SICAAS5020 (b) SSR loci, 1: IC337300, 2: IC337303, 3: IC337307, 4: IC337311, 5: IC337313, 6: IC337318, 7: IC337327, 8: IC337335, 9: IC337338, 10: IC337340, 11: IC338633, 12: IC338639, 13: IC338650, 14: IC340855, 15: IC340963, 16: IC341376, 17: IC341382, 18: IC355776, 19: IC355794, 20: IC355800, 21: IC357343, 22: IC383467, 23: IC383568, 24: IC383633, 25: IC393056, 26: IC406534, 27: IC418394, 28: IC436955, 29: IC469863, 30: IC469880.

to infer population structure using STRUCTURE 2.3.4 software and delta K value was calculated using Structure Harvester. The highest delta K value was reported at $K = 3$, indicating that all 30 accessions could be grouped into three well-defined sub-groups (Fig. 3).

Discussion

Foxtail millet germplasm from the CHR was found to have ample variability in nutritional traits. After achieving food security in different parts of the world, nutritional security is the next focus of global agricultural research. The rich variability in fibre content of millet germplasm found in the present study could be due to the difference in their genetic makeup. Accumulating evidence favours the view that increased intake of dietary fibre has beneficial effects including prevention or alleviation of maladies such as cardiovascular disease, diabetes, diverticulosis and colon cancer (Abdul-Hamid and Luan, 2000). Some of the collected accessions have higher fibre content than all four controls, i.e. widely cultivated accessions, used in the current study. Information on fibre is now the third most sought-after health information in supermarkets in countries such as India, Australia, Western Europe and North America (Mehta, 2005). Hence, accessions high in fibre content might be preferable for breeders and important for a foxtail improvement programme.

Millet is considered as carbohydrates and dietary carbohydrate is essential for gastrointestinal integrity and functioning (Flight, 2006). They supply the body with the energy required for its various activities (Eastwood, 2003) and help in transporting

crucial micronutrients. In seeds of any species, carbohydrate content usually remains stable; however, considerable variability in this nutritional constituent makes the germplasm useful for breeding programmes.

Millets, particularly foxtail millet, are low in saturated fat; however, they are a good source of polyunsaturated fats (Akoh and Min, 2007), which help to lower low-density lipoprotein (LDL) cholesterol. In turn, low-LDL cholesterol reduces the risk of heart disease (Chow, 2008), so low-LDL foods such as foxtail millet become an important component of nutritional security (Diniz *et al.*, 2004).

In addition, considerable variability was found in the protein content of seeds. Although cereal grains contain relatively little protein compared with legume seeds, cereals are the most important food crops. They provide over 200 million tonnes of protein for the nutrition of humans and livestock, which is about three times the amount derived from the more protein-rich legume seeds (Shewry and Halford, 2002). In addition to their nutritional importance, cereal seed proteins also influence the utilization of the seed in food processing. The storage proteins of cereals are of immense importance in determining the quality and end-use properties of the grain (Shewry and Halford, 2002). Essential amino acids are crucial for nutritional security; hence, two amino acids (lysine and thiamine) were analysed in the germplasm and showed remarkable variability in their content. The food supply of developed countries is rich in lysine; however, in poor countries where cereals dominate the food supply, lysine is the most limiting amino acid in the food supply (Baker, 2007). It is a strictly indispensable amino acid in humans and animals

Table 4. Characteristics of simple sequence repeat (SSR) loci used for diversity analysis in foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region

SSR loci	Chromosome (location, Kb)	na	ne	I	Exp_Het	Nei	Ave_Het	PIC	Fst	Nm
SICAAS2017	Chr.2 (41 333.1)	2	1.4	0.46	0.29	0.29	0.11	0.285	0.68	0.12
SICAAS3010	Chr.3 (4137.9)	3	2.59	1.02	0.63	0.61	0.22	0.615	0.65	0.13
SICAAS3052	Chr.3 (50 461.4)	3	2.92	1.08	0.67	0.66	0.48	0.657	0.24	0.8
SICAAS8028	Chr.8 (34 290.9)	4	1.38	0.56	0.28	0.27	0.25	0.302	0.26	0.72
SICAAS9030	Chr.9 (6681.4)	3	2.94	1.09	0.67	0.66	0.34	0.66	0.46	0.3
SICAAS7038	Chr.7 (29 176.6)	4	2.58	1.15	0.62	0.61	0.31	0.612	0.53	0.22
SICAAS5049	Chr.5 (23 544.2)	3	1.78	0.78	0.45	0.44	0.34	0.438	0.23	0.85
SICAAS5020	Chr.5 (28 017.3)	2	1.15	0.25	0.13	0.13	0.13	0.128	0.43	0.33
SICAAS9054	Chr.9 (23 440.3)	2	1.07	0.15	0.07	0.07	0.02	0.067	0.07	3.43
SICAAS9067	Chr.9 (58 046.7)	2	1.56	0.54	0.36	0.36	0.25	0.358	0.28	0.66
SICAAS7024	Chr.7 (20 228.2)	2	1.92	0.67	0.49	0.48	0.27	0.48	0.46	0.29
SICAAS4010	Chr.4 (40 053.8)	2	1.62	0.57	0.39	0.38	0.31	0.384	0.26	0.7
SICAAS5013	Chr.5 (12 297.9)	4	1.62	0.78	0.39	0.38	0.26	0.381	0.51	0.24
SICAAS4024	Chr.4 (1918.4)	4	2.78	1.14	0.65	0.64	0.34	0.64	0.43	0.33
SICAAS4037	Chr.4 (33 729.8)	2	1.15	0.26	0.14	0.13	0.06	0.133	0.8	0.06
SICAAS5081	Chr.5 (46 767.8)	2	1.23	0.33	0.19	0.19	0.14	0.186	0.33	0.52
SICAAS5034	Chr.5 (37 301.8)	2	2	0.69	0.51	0.5	0.23	0.499	0.55	0.21
SICAAS6101	Chr.6 (3493.8)	2	1.6	0.56	0.38	0.38	0.26	0.375	0.43	0.33
SICAAS6035	Chr.6 (35 880.9)	2	1.07	0.15	0.07	0.06	0.06	0.064	0.3	0.57
SICAAS9095	Chr.9 (2878.5)	2	1.07	0.15	0.07	0.07	0.06	0.066	0.47	0.29
SICAAS3056	Chr.3 (518.2)	2	1.43	0.48	0.31	0.3	0.21	0.302	0.3	0.59
SICAAS4061	Chr.4 (36 164.2)	2	1.47	0.5	0.33	0.32	0.25	0.32	0.36	0.44
SICAAS4027	Chr.4 (38 133.1)	2	2	0.69	0.51	0.5	0.2	0.499	0.5	0.25
SICAAS5005	Chr.5 (3492.1)	2	1.17	0.28	0.15	0.15	0.12	0.147	0.37	0.43
SICAAS6055	Chr.6 (7657.6)	3	2.28	0.9	0.57	0.56	0.36	0.561	0.37	0.43
Mean		2.52	1.75	0.61	0.37	0.37	0.22	0.366	0.42	0.34
St. Dev		0.77	0.61	0.32	0.2	0.2	0.11	0.198		

na, observed number of alleles; ne, effective number of allele; I, Shannon's information index; Exp_Het, expected heterozygosity; Nei, Nei's gene diversity, Ave_Het, average heterozygosity; PIC, polymorphism information content; Fst, genetic differentiation; Nm, gene flow.

(Tome and Bos, 2007); hence, untapped genetic diversity of foxtail millet having considerable variability in lysine content might be important for human nutrition as well as for feed.

Thiamine, also known as vitamin B₁, is essential for energy metabolism (Bettendorff *et al.*, 2014). Thiamine deficiency is uncommon in economically developed regions due to diversified diets and thiamine fortification of grains (Nathoo *et al.*, 2005; Dwyer *et al.*, 2015); however, severe thiamine deficiency leading to beriberi does occur in areas where dietary sources of thiamine are limited, such as Southeast Asia (Khounnorath *et al.*, 2011; Coats *et al.*, 2012). Ample variability in the thiamine content of foxtail millet germplasm signifies its potential relevance for breeding.

Foxtail millet is considered as a food suitable for diabetics, so its popularization and cultivation may help to minimize this global problem (Kam *et al.*, 2016). Unlike other common cereal foods such as rice and wheat, foxtail millet releases glucose steadily without affecting the body's metabolism (Jali *et al.*, 2012).

The available foxtail millet diversity has vast scope for supporting commercially grown crops by reducing pressure on their availability; it is also a cheap source of nutrients and can be raised at low management cost (Sankhala *et al.*, 2004).

Diversity based on phenotypic and morphological characters usually varies with environment. Molecular markers have been proven to be powerful tools in the assessment of genetic variation and in the elucidation of genetic relationships within and among species (Chakravarthi and Naravaneni, 2006). Unlike morphological traits, molecular markers are not affected by environment (Staub *et al.*, 1997). Collecting DNA marker data to determine whether phenotypically similar cultivars are genetically similar would, therefore, be of great interest in breeding for economically important traits (Duzyaman, 2005). In the present study, the number of alleles ranged from two to four, with an average of 2.5 ± 0.77 alleles/SSR locus, which is comparable with an earlier study by Jia *et al.* (2009), i.e. 2.5 alleles/SSR locus, and higher than other previous reports: 2.15 alleles/SSR locus (Pandey *et al.*, 2013), 2.2 alleles/SSR locus (Gupta *et al.*, 2012; Kumari

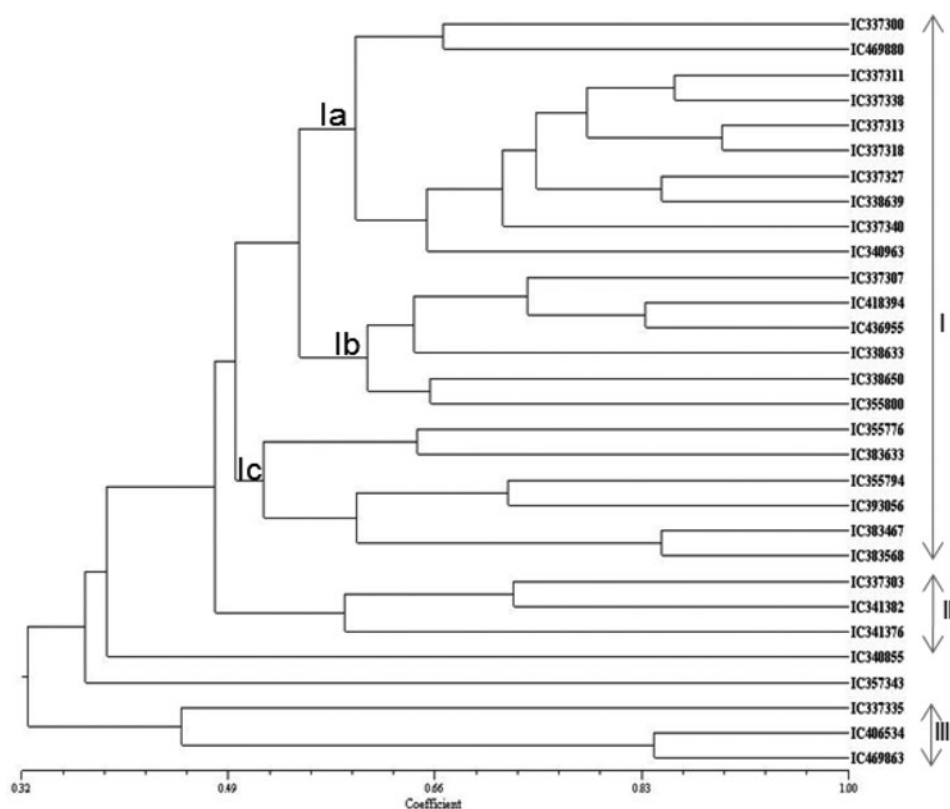


Fig. 2. UPGMA cluster of 30 foxtail millet accessions based on simple sequence repeat (SSR) markers.

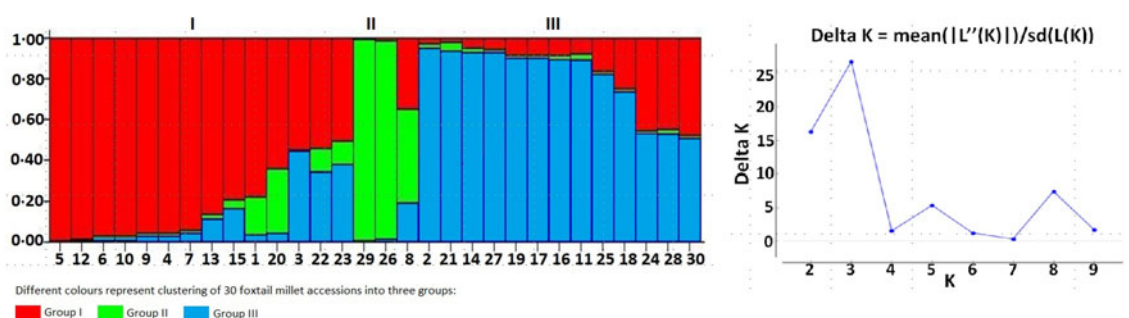


Fig. 3. Structure and ΔK statistics-based graphical representation of estimated number of clusters for K values 1 to 10. 1: IC337300, 2: IC337303, 3: IC337307, 4: IC337311, 5: IC337313, 6: IC337318, 7: IC337327, 8: IC337338, 9: IC337335, 10: IC337340, 11: IC338633, 12: IC338639, 13: IC338650, 14: IC340855, 15: IC340963, 16: IC341376, 17: IC341382, 18: IC355776, 19: IC355794, 20: IC355800, 21: IC357343, 22: IC383467, 23: IC383568, 24: IC383633, 25: IC393056, 26: IC406534, 27: IC418394, 28: IC436955, 29: IC469863, 30: IC469880. Colour online.

et al., 2013) and 2.4 alleles/SSR locus (Lin *et al.*, 2012). The observed heterozygosity of the germplasm is comparable with that obtained earlier by Gupta *et al.* (2012). The PIC values are also comparable with the earlier report by Gupta *et al.* (2012), i.e. 0.45, and lower than values of 0.69 and 0.45 reported by Jia *et al.* (2009) and Liu *et al.* (2011), respectively. Jia *et al.* (2009) and Liu *et al.* (2011) found higher diversity in Chinese germplasm as compared with the Indian germplasm used in the present study. This may be because different SSR markers were used in these studies; secondly, China is the centre of origin of foxtail millet whereas region-specific (i.e. CHR of India) germplasm was used in the present study for evaluation of nutritional quality. The Shannon's Information Index, Nei's expected heterozygosity and PIC values obtained in the present study also suggest considerable variability in the *S. italica* germplasm analysed. Based on the present analysis, the most informative SSR markers identified

for *S. italica* are SICAAS3052, SICAAS9030, SICAAS4024, SICAAS3010 and SICAAS7038. According to Wright's (1931) rule, if the value of N_m is <1 , populations will diverge. A low N_m value is generally believed to occur in self-pollinating systems such as foxtail millet. In the present study, the obtained N_m value was 0.34, indicating significant genetic differentiation. It has been observed that the accessions IC338633, IC406534, IC337327, IC355800, IC383568, IC338633 and IC337318 showed the highest values of dietary fibre, carbohydrate, fat, protein, lysine, thiamine and 1000 seed weight, respectively, which are higher than all four controls included in the present study, hence may be utilized in foxtail millet improvement for the traits studied. Furthermore, pairs of accessions showing the minimum and maximum values for these traits were examined for their Jaccard's similarity coefficient values (Supplementary Table 1) based on SSR markers used in the present study. The following pairs of accessions showed 27–

73% variation between them based on Jaccard's similarity coefficient values (IC338633 and IC357343, 0.53); (IC406534 and IC337300, 0.27); (IC337327 and IC337335, 0.432); (IC355800 and IC337300, 0.567); (IC383568 and IC355794, 0.567); (IC338633 and IC337300, 0.40) and (IC337318 and IC338639, 0.73) and may be useful in breeding programmes for improving dietary fibre, carbohydrate, fat, protein, lysine, thiamine and thousand grain weight, respectively. Hence, molecular profiling may be useful to select genotypes for nutritional breeding. Selection and utilization of available genetic variability for nutritional quality based on morpho-physiological traits and molecular markers may be an easy and useful approach for food and nutritional security in resource-poor regions as well as crop improvement (Trivedi et al., 2015).

Conclusion

A wide gene pool with known nutritional traits having diversity at the molecular level might be decisive for continued improvement of a crop through breeding. Results reveal considerable diversity and correspondence of molecular clustering and nutritional traits that can be utilized for designing further strategies, tailoring nutritionally rich varieties.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S0021859618000382>

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Ethical standards. Not applicable.

References

- Abdul-Hamid A and Luan YS (2000) Functional properties of dietary fibre prepared from defatted rice bran. *Food Chemistry* **68**, 15–19.
- Akoh CC and Min DB (2007) *Food Lipids: Chemistry, Nutrition and Biotechnology*, 3rd Edn. New York: Marcel Dekker.
- Anderson JA, Churchill GA, Autrique JE, Tanksley SD and Sorrells ME (1993) Optimizing parental selection for genetic linkage maps. *Genome* **36**, 181–186.
- AOAC (1998) AOAC official method 996.01 fats (total, saturated, unsaturated and monounsaturated) in cereal products. In Horwitz W (ed.), *AOAC Official Methods of Analysis*, 17th Edn. Arlington, VA: Association of Official Agricultural Chemists.
- AOAC (2005) AOAC official methods 2001.03 dietary fiber in foods containing resistant maltodextrin. In Horwitz W (ed.), *AOAC Official Methods of Analysis*, 18th Edn. Gaithersburg, MD: Association of Official Agricultural Chemists.
- Backes G and Østergaard H (2008) Molecular markers to exploit genotype-environment interactions of relevance in organic growing systems. *Euphytica* **163**, 523–531.
- Baker DH (2007) Lysine, arginine, and related amino acids: an introduction to the 6th amino acid assessment workshop. *The Journal of Nutrition* **137** (suppl. 2), 1599S–1601S.
- Bennetzen JL, Schmutz J, Wang H, Percifield R, Hawkins J, Pontaroli AC, Estep M, Feng L, Vaughn JN, Grimwood J, Jenkins J, Barry K, Lindquist E, Hellsten U, Deshpande S, Wang X, W, X, Mitros T, Triplett J, Yang X, Ye CY, Mauro-Herrera M, Wang L, Li P, Sharma M, Sharma R, Ronald PC, Panaud O, Kellogg EA, Brutnell TP, Doust AN, Tuskan GA, Rokhsar D and Devos KM (2012) Reference genome sequence of the model plant *Setaria*. *Nature Biotechnology* **30**, 555–561.
- Bettendorff L, Lakaye B, Kohn G and Wins P (2014) Thiamine triphosphate: a ubiquitous molecule in search of a physiological role. *Metabolic Brain Disease* **29**, 1069–1082.
- Chakravarthi BK and Naravaneni R (2006) SSR marker based DNA fingerprinting and diversity study in rice (*Oryza sativa* L.). *African Journal of Biotechnology* **5**, 684–688.
- Chen Q, Li D, Yang H, Zhu Q, Zheng H and Xu J (1999) Novel spectrofluorimetric method for the determination of thiamine with iron(III) tetrasulfonatophthalocyanine as a catalyst. *The Analyst* **124**, 771–775.
- Chow CK (ed.) (2008) *Fatty Acids in Foods and Their Health Implications*, 3rd Edn. Boca Raton, FL: CRC Press.
- Coats D, Shelton-Dodge K, Ou K, Khun V, Seab S, Sok K, Prou C, Tortorelli S, Moyer TP, Cooper LE, Begley TP, Enders F, Fischer PR and Topazian M (2012) Thiamine deficiency in Cambodian infants with and without beriberi. *Journal of Pediatrics* **161**, 843–847.
- Diao X, Schnable J, Bennetzen JL and Li J (2014) Initiation of *Setaria* as a model plant. *Frontiers of Agricultural Science and Engineering* **1**, 16–20.
- Diniz YS, Cicogna ACC, Padovani CR, Santana LS, Faine LA and Novelli ELB (2004) Diets rich in saturated and poly unsaturated fatty acids: metabolic shifting and cardiac health. *Nutrition* **20**, 230–234.
- Doust AN, Kellogg EA, Devos KM and Bennetzen JL (2009) Foxtail millet: a sequence-driven grass model system. *Plant Physiology* **149**, 137–141.
- Duzyaman E (2005) Phenotypic diversity within a collection of distinct okra (*Abelmoschus esculentus*) cultivars derived from Turkish land races. *Genetic Resources and Crop Evolution* **52**, 1019–1030.
- Dwivedi SL, Crouch JH, Mackill DJ, Xu Y, Blair MW, Ragot M, Upadhyaya HD and Ortiz R (2007) The molecularization of public sector crop breeding: progress, problems and prospects. *Advances in Agronomy* **95**, 163–318.
- Dwyer JT, Wiemer KL, Dary O, Keen CL, King JC, Miller KB, Philbert MA, Tarasuk V, Taylor CL, Gaine PC, Jarvis AB and Bailey RL (2015) Fortification and health: challenges and opportunities. *Advances in Nutrition* **6**, 124–131.
- Earl DA and von Holdt BM (2012) Structure harvester: a website and program for visualizing structure output and implementing the Evanno method. *Conservation Genetics Resources* **4**, 359–361.
- Eastwood M (2003) *Principles of Human Nutrition*, 2nd Edn. Malden, MA: Blackwell Science.
- Flight I (2006) Cereal grains and legumes in the prevention of coronary heart disease and stroke: a review of the literature. *European Journal of Clinical Nutrition* **60**, 1145–1159.
- Gupta S, Kumari K, Sahu PP, Vidapu S and Prasad M (2012) Sequence-based novel genomic microsatellite markers for robust genotyping purposes in foxtail millet (*Setaria italica* (L.) P. Beauv.). *Plant Cell Reports* **31**, 323–337.
- Hurrell RF and Carpenter KJ (1981) The estimation of available lysine in foodstuffs after Maillard reactions. *Progress in Food and Nutrition Science* **5**, 159–176. (Accessed 13 March 2018).
- Jali MV, Kamatar MY, Jali SM, Hiremath MB and Naik RK (2012) Efficacy of value added foxtail millet therapeutic food in the management of diabetes and dyslipidemia in type 2 diabetic patients. *Recent Research in Science and Technology* **4**, 3–4.
- Jia G, Huang X, Zhi H, Zhao Y, Zhao Q, Li W, Chai Y, Yang L, Liu K, Lu H, Zhu C, Lu Y, Zhou C, Fan D, Weng Q, Guo Y, Huang T, Zhang L, Lu T, Feng Q, Hao H, Liu H, Lu P, Zhang N, Li Y, Guo E, Wang S, Wang S, Liu J, Zhang W, Chen G, Zhang B, Li W, Wang Y, Li H, Zhao B, Li J, Diao X and Han B (2013) A haplotype map of genomic variations and genome-wide association studies of agronomic traits in foxtail millet (*Setaria italica*). *Nature Genetics* **45**, 957–961.
- Jia X, Zhang Z, Liu Y, Zhang C, Shi Y, Song Y, Wang T and Li Y (2009) Development and genetic mapping of SSR markers in foxtail millet (*Setaria italica* (L.) P. Beauv.). *Theoretical and Applied Genetics* **118**, 821–829.
- Jones JM (2001) The benefits of eating breakfast cereals. *Cereal Foods World* **46**, 461–464, 466–467.

- Jones JR, Lineback DM and Levine MJ (2006) Dietary reference intakes: implications for fiber labeling and consumption: a summary of the international life sciences institute North America fiber workshop, June 1–2, 2004, Washington, DC. *Nutrition Reviews* **64**, 31–38.
- Kam J, Puranik S, Yadav R, Manwaring HR, Pierre S, Srivastava RK and Yadav RS (2016) Dietary interventions for type 2 diabetes: how millet comes to help. *Frontiers in Plant Science* **7**, 1454.
- Khounnorath S, Chamberlain K, Taylor AM, Soukaloun D, Mayxay M, Lee SJ, Phengdy B, Luangxay K, Sisouk K, Soumphonphakdy B, Latsavong K, Akkhavong K, White NJ and Newton PN (2011) Clinically unapparent infantile thiamin deficiency in Vientiane, Laos. *PLoS Neglected Tropical Diseases* **5**, e969. <https://doi.org/10.1371/journal.pntd.0000969>
- Kumari K, Muthamilarasan M, Misra G, Gupta S, Subramanian A, Parida SK, Chattopadhyay D and Prasad M (2013) Development of eSSR-markers in *Setaria italica* and their applicability in studying genetic diversity, cross-transferability and comparative mapping in millet and non-millet species. *PLoS ONE* **8**, e67742.
- Lammerts van Bueren ET, Backes G, de Vriend H and Østergård H (2010) The role of molecular markers and marker assisted selection in breeding for organic agriculture. *Euphytica* **175**, 51–64.
- Lata C, Gupta S and Prasad M (2013) Foxtail millet: a model crop for genetic and genomic studies in bioenergy grasses. *Critical Reviews in Biotechnology* **33**, 328–343.
- Lee YK (1994) The Beri-beri Hospital, Singapore (1907–1925). *Singapore Medical Journal* **35**, 306–311.
- Leonard WH and Martin JH (1963) *Cereal Crops*. New York: Macmillan.
- Li P and Brutnell TP (2011) *Setaria viridis* and *Setaria italica*, model genetic systems for the Panicoid grasses. *Journal of Experimental Botany* **62**, 3031–3037.
- Li HW, Meng CJ and Liu TM (1935) Problems in the breeding of millet (*Setaria italica* (L.) Beauv.). *Agronomy Journal* **27**, 963–970.
- Li S, An S, Liu Z, Cheng R and Wang Z (2014) Innovation of the new superior quality foxtail millet (*Setaria italica* (L.) P. Beauv.) variety-Jigu32 with characteristics of stress resistance, stable and high yield and its physiological mechanism. *Agricultural Sciences* **5**, 304–316.
- Lin HS, Chiang CY, Chang SB, Liao GI and Kuoh CS (2012) Genetic diversity in the foxtail millet (*Setaria italica*) germplasm as determined by agronomic traits and microsatellite markers. *Australian Journal of Crop Science* **6**, 342–349.
- Liu Z, Bai G, Zhang D, Zhu C, Xia X, Cheng R and Shi Z (2011) Genetic diversity and population structure of elite foxtail millet (*Setaria italica* (L.) P. Beauv.) germplasm in China. *Crop Science* **51**, 1655–1663.
- Lonsdale D (1990) Thiamine deficiency and sudden deaths (letter; comment). *The Lancet* **336**, 376.
- Lowry OH, Rosenbrough NJ, Farr AL and Randall RJ (1951) Protein measurement with the Folin-phenol reagent. *Journal of Biological Chemistry* **193**, 265–275.
- Mehta RS (2005) Dietary fiber benefits. *Cereal Foods World* **50**, 66–71.
- Morris DL (1948) Quantitative determination of carbohydrates with Dreywood's anthrone reagent. *Science* **107**, 254–255.
- Muthamilarasan M and Prasad M (2015) Advances in *Setaria* genomics for genetic improvement of cereals and bioenergy grasses. *Theoretical and Applied Genetics* **128**, 1–14.
- Muthamilarasan M, Venkata Suresh B, Pandey G, Kumari K, Parida SK and Prasad M (2014) Development of 5123 intron-length polymorphic markers for large-scale genotyping applications in foxtail millet. *DNA Research* **21**, 41–52.
- Nathoo T, Holmes CP and Ostry A (2005) An analysis of the development of Canadian food fortification policies: the case of vitamin B. *Health Promotion International* **20**, 375–382.
- Pandey G, Misra G, Kumari K, Gupta S, Parida SK, Chattopadhyay D and Prasad M (2013) Genome-wide development and use of microsatellite markers for large-scale genotyping applications in foxtail millet (*Setaria italica* (L.)). *DNA Research* **20**, 197–207.
- Pritchard JK, Stephens M and Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* **155**, 945–959.
- Rohlf FJ (2000) *NTSYS-PC, Numerical Taxonomy System for the PC Exeter Software*, Version 2.1. Setauket: Applied Biostatistics Inc.
- Ronald P (2011) Plant genetics, sustainable agriculture and global food security. *Genetics* **188**, 11–20.
- Saghai-Marouf MA, Soliman KM, Jorgensen RA and Allard RW (1984) Ribosomal DNA spacer-length polymorphisms in barley: Mendelian inheritance, chromosomal location and population dynamics. *Proceedings of the National Academy of Sciences of the United States of America* **81**, 8014–8018.
- Sahley BJ and Birkner KM (2000) *Healing with Amino Acids and Nutrients*. San Antonio, TX: Pain and Stress Publications, Inc.
- Sankhala A, Chopra S and Sankhala AK (2004) Effect of processing on tannin, phytate and in vitro iron in underutilized millets – bajra (*Pennisetum typhoideum*) and Kangni (*Setaria italica*). *Indian Journal of Nutrition and Dietetics* **41**, 55–62.
- Shewry PR and Halford NG (2002) Cereal seed storage proteins: structures, properties and role in grain utilization. *Journal of Experimental Botany* **53**, 947–958.
- Sica DA (2007) Loop diuretic therapy, thiamine balance, and heart failure. *Congestive Heart Failure* **13**, 244–247.
- Staub JC, Serquen FC and McCreight JD (1997) Genetic diversity in cucumber (*Cucumis sativus* L.). III. An evaluation of Indian germplasm. *Genetic Resources and Crop Evolution* **44**, 315–326.
- Suresh VB, Muthamilarasan M, Misra G and Prasad M (2013) FmMdb: a versatile database of foxtail millet markers for millets and bioenergy grasses research. *PLoS ONE* **8**, e71418.
- Tome D and Bos C (2007) Lysine requirement through the human life cycle. *Journal of Nutrition* **137**(suppl. 2), 1642S–1645S.
- Trivedi AK, Arya L, Verma M, Verma SK, Tyagi RK and Hemantaranjan A (2015) Genetic variability in proso millet (*Panicum miliaceum*) germplasm of Central Himalayan region based on morpho-physiological traits and molecular markers. *Acta Physiologica Plantarum* **37**, 23. <https://doi.org/10.1007/s11738-014-1770-y>
- Turesson S, Svensson E, Happstadius I, Henriksson T and Kazman E (2009) Application of markers when breeding for baking quality. In Østergård H, Lammerts van Bueren ET and Bouwman-Smits L (eds), *Proceedings of the BioExploit/Eucarpia Workshop on the Role of Marker Assisted Selection in Breeding Varieties for Organic Agriculture*. Wageningen, The Netherlands: BioExploit, pp. 38–39. Available at <https://www.eucarpia.org/organic-and-low-input-agriculture/190-workshop-on-the-role-of-marker-assisted-selection-in-breeding-varieties-for-organic-agriculture.html> (Accessed 13 March 2018).
- Wang ZM, Devos KM, Liu CJ, Wang RQ and Gale MD (1998) Construction of RFLP-based maps of foxtail millet, *Setaria italica* (L.) P. Beauv. *Theoretical and Applied Genetics* **96**, 31–36.
- Wooley JA (2008) Characteristics of thiamin and its relevance to the management of heart failure. *Nutrition in Clinical Practice* **23**, 487–493.
- Wright S (1931) Evolution in Mendelian populations. *Genetics* **16**, 97–159.
- Xu Y and Crouch JH (2008) Marker-assisted selection in plant breeding: from publications to practice. *Crop Science* **48**, 391–407.
- Yadav CB, Bonthala VS, Muthamilarasan M, Pandey G, Khan Y and Prasad M (2015) Genome-wide development of transposable element-based markers in foxtail millet and construction of an integrated database. *DNA Research* **22**, 79–90.
- Yeh FC, Yang RC and Boyle T (1999) *Popgene Version 1.32. Microsoft Window-Based Freeware for Population Genetic Analysis*. Edmonton, Canada: Quick User Guide, University of Alberta and Center for International Forestry Research. Available at https://sites.ualberta.ca/~fyeh/popgene_download.html (Accessed 13 March 2018).
- Zhang S, Tang C, Zhao Q, Li J, Yang L, Qie L, Fan X, Li L, Zhang N, Zhao M, Liu X, Chai Y, Zhang X, Wang H, Li Y, Li W, Zhi H, Jia G and Diao X (2014) Development of highly polymorphic simple sequence repeat markers using genome-wide microsatellite variant analysis in foxtail millet (*Setaria italica* (L.) P. Beauv.). *BMC Genomics* **15**, 78.