



Effect of temperature and/or sweetness of beverages on body composition in rats

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Abstract

Sweetened beverages are mainly consumed cold and various processes are activated in response to external temperature variations. However, the effect of internal temperature variations through the ingestion of cold beverages is far from clear. Two experiments were conducted to investigate the effect of beverage temperature on body composition. Sprague–Dawley rats (5–6-week-old males) had free access to food and beverage for 8 weeks. Energy intake, body weight and body composition were monitored. In Expt 1, two groups of rats (*n* 9) consumed water at room temperature (NW about 22°C) or cold (CW about 4°C). In Expt 2, rats were offered room-temperature (N) or cold (C) sweetened water (10% sucrose CSu (*n* 7) and NSu (*n* 8); or 0.05% acesulfame K CAk (*n* 6) and NAK (*n* 8)) for 12 h, followed by plain water. Our results show that in Expt 1, CW had higher lean body mass ($P < 0.001$) and lower body fat gain ($P = 0.004$) as compared with NW. In Expt 2, body weight ($P = 0.013$) and fat ($P \leq 0.001$) gains were higher in the non-energetic sweetened groups, while lean body mass was not affected by the type of sweeteners or temperature. In conclusion, cold water ingestion improved lean body mass gain and decreased fat gain because of increased energy expenditure, while non-energetic sweetener (acesulfame K) increased body fat gain due to improved energy efficiency. Internal cold exposure failed to increase energy intake in contrast to that of external cold exposure.

Key words: Water: Sweeteners: Cold drink intake: Lean body mass: Body fat

Energy metabolism is known to be affected by ambient 'external' temperature. In both humans and rats, external cold exposure was reported to increase energy expenditure^(1,2) and energy intake^(3–5) to cover the cost of energy needed to sustain body temperature at 37°C, in what is known as a form of a protective mechanism against hypothermia. In rats, external cold exposure (6°C) for 3 weeks was reported to stimulate *ad libitum* food intake and thermogenesis (increase in interscapular brown adipose tissue mass), as an adaptive mechanism to low temperature⁽⁶⁾. The activation of brown adipose thermogenesis, through increased heat generation⁽⁷⁾ and cutaneous thermoreceptors⁽⁸⁾, was proposed to counteract the variations in external temperatures. Moreover, in observational studies on humans, the positive association between ambient temperature with body weight and obesity may relate to the energy cost to sustain body temperature at 37°C^(9,10).

Whereas the impact of external cold exposure was extensively studied, the research on the effect of cold beverages ingestion is scarce. In humans, acute ingestion of cold water (about 4°C) was reported to increase energy expenditure (up to 25%)^(11,12) and lower the hypothalamic activity associated with satiety⁽¹³⁾. In broiler chickens, chronic ingestion of cold water (9°C) was reported to be positively associated with higher dietary intake and weight increment, in contrast to the decrease in growth and the metabolic rate usually witnessed under extreme heat stress conditions⁽¹⁴⁾. While, in cows, drinking of chilled water (10°C) induced an increase in feed intake^(15,16). Yet, it is not clear whether body composition is affected by the variations in the temperature of ingested water in the long term.

On the other hand, the consumption of sweetened (both energetic and non-energetic) beverages was reported to

Abbreviations: CAk, cold water (about 4°C) sweetened with 0.05% acesulfame K; CSu, cold water (about 4°C) sweetened with 10% sucrose; CW, cold water; NAK, room-temperature water (about 22°C) sweetened with 0.05% acesulfame K; NSu, room-temperature water (about 22°C) sweetened with 10% sucrose; NW, room-temperature water.

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be associated with the development of obesity and its related diseases^(17–19). Beverages are mainly ingested cold⁽¹³⁾, and the potential involvement of the temperature of these beverages in the aetiology of obesity is far from clear. Furthermore, the impact of non-energetic sweeteners on obesity was surrounded by controversies due to the failure of non-energetic sweeteners to halt the prevalence of obesity in the last couple of decades⁽²⁰⁾. Short-term studies reported favourable outcomes^(21,22), while in contrast, long-term studies reported detrimental effects on weight gain, the incidence of type 2 diabetes and CVD^(23,24). In animals, the impact of non-energetic sweeteners on body weight and food intake was reported to be highly variable, and this may, in part, relate to the mode (diet or fluid), dose and/or type of ingested sweeteners^(25–28).

A study was conducted to address some of the potential causes of these controversies by investigating the impact of the temperature of ingested beverages on body weight measures. The long-term effect of temperature manipulation of plain, energetic and non-energetic sweetened water on body measures, using the most common concentrations, was studied. Two experiments were conducted. In the first, we investigated whether presenting rats with cold water (about 4°C) can affect body weight measures and energy metabolism and can generate an increase in food intake in the same ways as with changes in external temperature. In the second experiment, we examined if beverage temperature manipulation can affect the impact of sweeteners – energetic (sucrose) and non-energetic (acesulfame K) – on body weight measures.

Materials and methods

Animal housing

The Institutional Animal Care and Use Committee of the American University of Beirut, Lebanon, approved the experimental protocol (no. 18-02-453). The study was performed by following the criteria outlined in the Guide for the Care and Use of Laboratory Animals. Male Sprague–Dawley rats (5–6 weeks old, Animal Care Facility, American University of Beirut) were housed individually in wire-bottom cages in a temperature 22 (SD 1)°C and light (reverse light cycle 12 h dark–12 h light, lights off at 10.00 hours) controlled room. Food was offered in regular food containers, easily accessible for rats, and water was given in water bottles specific for the wire-bottom cages.

Experimental design

Two separate experiments were executed. In each, all rats were placed on a 1-week adaptation period to familiarise them with the environment and the semi-synthetic diet (online Supplementary Table S1) and, then afterwards, they were randomly divided into their respective experimental groups for 8 weeks as follows:

Expt 1: effect of cold and room-temperature drinking water on body weight measures. Two groups of rats (*n* 9 per group) were used, group 1 (CW) was given cold water (about 4°C) (online Supplementary Fig. S1) and group 2 (NW) was given water at room temperature (about 22°C).

Expt 2: effect of cold and room-temperature sweetened (energetic and non-energetic) drinking water on body weight measures. Four groups were used in which sweetened (sucrose or acesulfame K) water was offered for 12 h (in the dark phase), followed by 12 h of plain water. The beverages of two of these groups were provided at a cold temperature (online Supplementary Fig. S1), as follows:

- Group 1 (CSu, *n* 7): Cold (about 4°C) sweetened water (10% sucrose energetic sweetener) was offered for 12 h, followed by 12 h of cold (about 4°C) plain water.
- Group 2 (NSu, *n* 8): Room-temperature (about 22°C) sweetened water (10% sucrose energetic sweetener) was offered for 12 h, followed by 12 h of room-temperature plain water.
- Group 3 (CAk, *n* 6): Cold (about 4°C) sweetened water (0.05% acesulfame K non-energetic sweetener; HYET Sweet) was offered for 12 h followed by 12 h of cold (about 4°C) plain water.
- Group 4 (NAk, *n* 8): Room-temperature (about 22°C) sweetened water (0.05% acesulfame K non-energetic sweetener) was offered for 12 h, followed by 12 h of plain room-temperature water.

Rats were fed *ad libitum* for 8 weeks their respective beverage and a semi-synthetic powder control diet (online Supplementary Table S1) based on the AIN-93G⁽²⁹⁾ for optimal growth.

On the day of sacrifice, overnight fasted rats were anaesthetised with isoflurane (Forane®) and blood was collected from the superior vena cava. After that, rats were euthanised by severing their hearts and their livers were immediately excised, weighed, frozen in liquid N₂ and stored at –80°C. Blood samples were centrifuged at 2200 *g* (3°C) for 15 min, and aliquots of plasma were collected and stored at –80°C until analysed.

Cold beverages

The bottles of the varied beverages (plain and/or sweetened water) were frozen; 10 ml of the corresponding cold beverage was added to the bottle before being weighed and offered to the rats. Bottles were wrapped in thermal insulators and changed every 6 h in order to maintain their coldness at a mean of 4°C throughout the experimental period (online Supplementary Fig. S1).

Food intake, body weight and composition

Food and water intake (difference in the weight of the water bottles), body weight and body composition using NMR minispec (LF110 BCA analyzer) were measured twice per week. Body weight, lean body mass and body fat mass were expressed as gross weight as well as gain changes from baseline to minimise the impact of the variations in initial body weight within the groups. Energy expenditure was estimated from the total energy intake and changes in body mass and composition⁽³⁰⁾. Energy efficiency was calculated as the energy retained in the body per 100 kJ of energy consumed (kJ/100 kJ).

Livers were freeze-dried (FreeZone 6 Freeze Dryer, LABCONCO), and lipid was extracted with light petroleum



(40–60°C) using an ANKOMXT10 extractor (ANKOM Technology). All determinations were carried out in duplicate.

Plasma analyses

Fasting plasma glucose, total cholesterol, HDL-cholesterol, TAG, albumin, plasma urea nitrogen and creatinine were determined with the Vitros 350 Chemistry System (Ortho-Clinical Diagnostics). The plasma insulin concentration was measured with an enzyme immunoassay using the Rat/Mouse Insulin ELISA Kit (EZRMI-13 K) (EMD Millipore Corporation).

Statistical analysis

The required number of rats was calculated using previously determined weight gain data (6.0 (SD 0.95) g/d) and assuming a 25 % difference in the mean, with a statistical power of 90 % and a 5 % significance level. Data were expressed as mean values and standard deviations. SPSS Statistics 25.0 software (IBM Corp.) was used for statistical analysis.

Expt 1: Unpaired sample *t* test was performed to compare the data between the two groups, and two-way ANOVA with time and temperature as fixed variables was used to analyse the results throughout the 8-week experimental period.

Expt 2: Two-way ANOVA with beverage temperature and sweetener type as factors was performed. Multiple-way ANOVA (general linear model) using time, sweetener and temperature as fixed factors was used to analyse the results throughout the 8-week experimental period.

Results

Expt 1: effect of cold and room-temperature drinking water on body weight measures

Baseline body weight, lean body mass and body fat were similar between the groups, and these parameters failed to reach significance by the end of the experimental period (online Supplementary Table S2). When changes from baseline (gain) were monitored over the experimental period, no differences in body weight gain were found between the groups ($P=0.220$) (Fig. 1(a)), while changes in body composition were detected, in which lean body mass of the cold water group (CW) was significantly higher than that of the room-temperature group (NW) ($P<0.001$) (Fig. 1(b)). However, the changes in body fat mass of the room-temperature group were significantly higher ($P=0.004$) than that of the cold water group (Fig. 1(c)).

No differences in water intake were seen between the groups, and both energy intake and efficiency were not found to be significantly affected by the temperature of ingested water. Nevertheless, the computed total energy expenditure of the cold water group (CW) was significantly higher than that of the normal temperature group (NW) ($P=0.003$) (Table 1).

Moreover, the temperature of the ingested water was found not to affect liver weight and composition, as well as epididymal fat tissue weight (Table 1).

The measured fasting plasma metabolites (glucose, insulin, cholesterol, albumin, plasma urea nitrogen and creatinine) were not affected by the temperature of the drinking water except for

the TAG levels that were found to be lower in the cold water group ($P=0.031$) (Table 2).

Expt 2: effect of cold and room-temperature sweetened (energetic and non-energetic) drinking water on body weight measures

Baseline body weight, lean body mass and body fat were similar between the different groups, and no significant differences in these parameters were observed at the end of the experimental period (online Supplementary Table S3). However, when changes from baseline (gain) were monitored, a higher body weight gain was observed in the non-energetic sweetened groups (CAK and NAK) as compared with the energetic/sucrose sweetened groups (CSu and NSu) ($P=0.013$). In contrast, weight gain was not affected by the temperature of the beverage (Fig. 2(a)). Lean body mass gain of the cold temperature groups (CSu and CAK) was found to be slightly, but not significantly, higher than that of the normal temperature groups ($P=0.056$) (Fig. 2(b)). Body fat mass gain of the non-energetic sweetened groups was found to be significantly higher than that of the energetic sweetened groups ($P<0.001$), and a significant interaction ($P=0.041$) between the type of sweetener and temperature was detected (Fig. 2(c)).

The total energy intake level was similar between the four groups (Table 3). The energetic sweetened groups (CSu and NSu) had a lower energy intake from the diet since part of the energy was derived from the sucrose content of the beverage. In the energetic sweetened groups, sweetened water contributed to about 10 % and 25 % of the energy intake of the cold (CSu) and room-temperature (NSu) water, respectively (Table 3). The volume of ingested room-temperature sweetened water was higher than that ingested by the cold temperature groups. Nevertheless, the ingestion of plain water was significantly lower among the room-temperature groups (NSu and NAK) as compared with the cold temperature groups (CSu and CAK) (Table 3).

In contrast, the energy intake of sweetened drinking water was the highest among the normal temperature groups (NSu and NAK) irrespective of their energetic content. Moreover, the ingestion of cold sweetened water seems to decrease the energy expenditure of the energetic sweetened water group and increase the energy expenditure of the non-energetic sweetened water one ($P=0.026$ for cold and sweetness interaction). However, the energy efficiency of the non-energetic sweetener groups was significantly higher than that of the energetic sweetened groups ($P=0.049$), and this was not affected by the temperature of the ingested sweetened water (Table 3).

At the organ level, no significant differences were found between the groups in terms of total liver weight (g), percentage liver weight of body weight (g/100 g body weight) and epididymal adipose tissue weight. Nonetheless, liver fat (%) of the non-energetic sweetened groups was more than 50 % higher than that of the energetic sweetened groups ($P=0.004$), and this was associated with lower liver water (%) ($P=0.039$) (Table 3). All fasting plasma parameters were found not to be affected neither by energetic content nor by the temperature of the water except for that of plasma urea nitrogen, which was found to be higher in the non-energetic sweetened groups (Table 4).



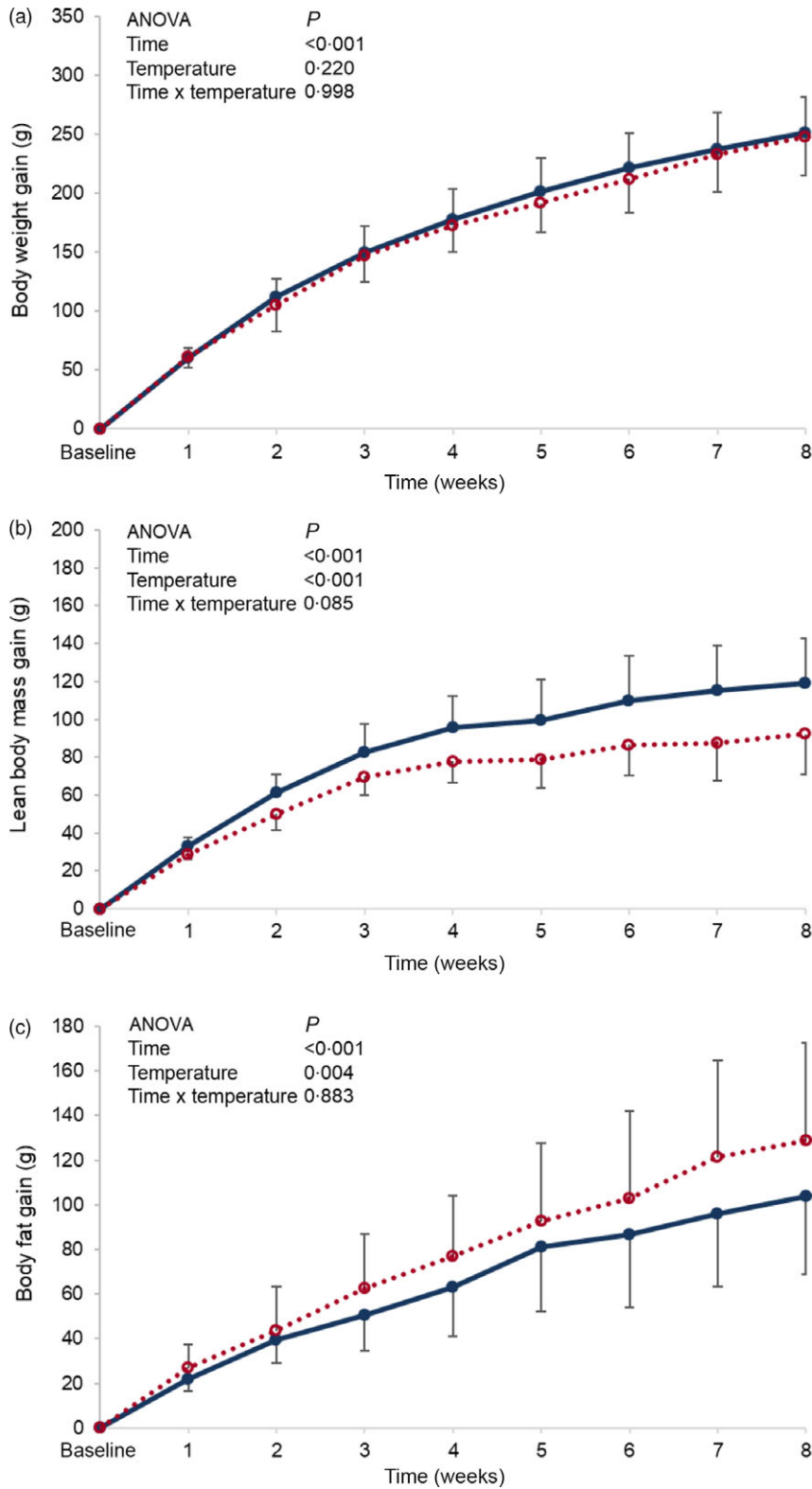


Fig. 1 Expt 1 – Effect of cold and room-temperature drinking water on body weight measures. Weekly body weight (a), lean body mass (b) and body fat (c) gain from baseline in grams of the two groups of rats over the 8-week experimental period. Group CW: cold water (about 4°C); Group NW: room-temperature water (about 22°C). Data are expressed as mean values and standard deviations of all values. A two-way ANOVA was performed with time and temperature of the water as factors, and time set as random. Significance was set at $P < 0.05$. —●—, CW; ····, NW.

Table 1. Expt 1 – Effect of cold and room-temperature drinking water on energy intake, expenditure, efficiency and some organ weights (Wt) (Mean values and standard deviations)*

	CW		NW		<i>P</i>
	Mean	SD	Mean	SD	
Mean energy intake (kJ/d)	390.51	26.20	376.84	31.94	0.329
Mean water intake (g/d)	24.70	3.37	29.49	8.95	0.163
Mean energy efficiency (kJ/100 kJ)	20.74	5.04	25.51	6.40	0.099
Mean energy expenditure (kJ/d)	308.72	16.41	279.33	18.79	0.003
Liver wet wt (g)	14.50	1.65	14.60	2.34	0.918
Liver % (g/100 g body wt)	2.91	0.24	2.94	0.30	0.803
Liver water (%)	69.69	1.83	69.97	0.91	0.689
Liver fat in wet (%)	5.07	1.07	4.93	1.23	0.817
Epididymal adipose tissue wt (g)	12.08	3.91	13.33	3.85	0.505

Group CW, cold water (about 4°C); Group NW, room-temperature water (about 22°C). * Rats were maintained for 8 weeks on cold or room-temperature water. An unpaired sample *t* test was performed. Significance was set at a *P* < 0.05.

Table 2. Expt 1 – Effect of cold and room-temperature drinking water on plasma metabolites (Mean values and standard deviations)*

	CW		NW		<i>P</i>
	Mean	SD	Mean	SD	
Glucose (mmol/l)	14.65	2.95	13.80	2.89	0.552
Insulin (ng/ml)	5.15	3.57	6.52	7.90	0.645
Total-cholesterol (mmol/l)	1.73	0.38	1.91	0.50	0.420
HDL-cholesterol (mmol/l)	1.50	0.18	1.60	0.50	0.358
TAG (mmol/l)	0.42	0.08	0.67	0.28	0.031
Albumin (g/l)	39.30	4.20	40.20	2.90	0.610
Plasma urea nitrogen (mmol/l)	4.56	0.89	4.52	0.40	0.905
Creatinine (μmol/l)	30.06	6.19	26.52	7.96	0.257

Group CW, cold water (about 4°C); Group NW, room-temperature water (about 22°C). * An unpaired sample *t* test was performed. Significance was set at a *P* < 0.05.

Discussion

Thermoregulation studies have focused mainly on the effect of external cold exposure on body composition, energy balance and others, while little is known about the influence of internal cold exposure. The present study was designed to address the impact of internal cold exposure that was induced by the ingestion of cold beverages, including plain water, energetic and non-energetic sweetened water. The study adopted several pragmatic approaches to mimic real-life scenarios. First, the studied temperature was similar to the commonly used refrigerated beverages. Second, the concentration of sweeteners (energetic and non-energetic) was comparable to that used in commercially available beverages. Third, the sweetened water was offered for 12 h/d, for it not to be the only source of fluid, and to avoid the continuous ingestion of sweeteners and any potential problem in osmolarity.

Our results show that the ingestion of cold water was able to alter body composition by increasing lean body mass gain and reducing fat mass accumulation. However, changes in body composition were not evident in the cold sweetened (energetic or non-energetic) groups. The alterations in body composition

were not associated with any significant changes in energy intake, unlike what was reported following external cold exposure^(4,6). Though, in other animals, chicks and cows, drinking of chilled water was shown to increase feed consumption^(14,16). Adaptation to external cold exposure was reported to increase both food intake and expenditure to sustain body thermoneutrality^(4,5,31). These adaptive measures are, in part, mediated through cutaneous cold thermoreceptors (TRPM18), mainly since their deletion induced weight gain that was associated with a reduction in fat oxidation⁽³²⁾. This points towards a minor role for digestive tract thermoreceptors in the control of feeding, although they were reported to affect gut motility^(33,34). Moreover, it is reasonable to postulate that the observed increase in lean body mass following internal cold exposure might have been an adaptive process to sustain body thermoneutrality, primarily since lean mass induces heat production^(35,36). In brief, these findings imply that the adaptive mechanisms for internal cold exposure are not identical to that of external cold exposure^(1,6).

In the first experiment, the changes in body composition were highly attributed to increased energy expenditure (about 10%) that seems to equate to about 8 h of external cold exposure at 4°C⁽⁷⁾. In line, energy expenditure was reported to increase under conditions of short-term ingestion of cold water and/or exposure to cooler external temperature^(1,2,11,12,37). In humans, the ingestion of cold water at 22°C was reported to increase post-prandial thermogenesis by about 30%, in which the cost of water warming to 37°C contributed to about 40% of the increase⁽³⁸⁾. In the present study, the cost of warming is expected to be higher as the water was offered at 4°C. The ability of systemic β-adrenoreceptor blockade to reduce thermogenesis implicates the sympathetic nervous system in water thermogenesis⁽³⁸⁾, which is believed to be mediated by osmoreceptors since the ingestion of saline solution failed to substantially affect thermogenesis^(12,39,40). Furthermore, the cost of the observed increase in energy expenditure with cold water ingestion seems to be mainly derived from fat oxidation, as indicated by the reduction in body fat and plasma TAG. In agreement, external cold exposure was reported to increase lipoprotein lipase of white adipose tissue and heart, as well as fatty acid oxidation activities in heart and white and brown adipose tissues^(41–43). Moreover, the cost of the thermogenic effect of water ingestion in male subjects was mainly derived from lipid oxidation⁽³⁸⁾.

On the other hand, in the energetic/non-energetic sweetener study, the impact of the ingestion of cold temperature water on lean body mass was of low magnitude. Surprisingly, despite the similarities between the groups in terms of total energy intake, the ingestion of non-energetic sweetened water was associated with higher body weight and body fat mass and this was paralleled by higher hepatic fat content. Increased fat deposition in the non-energetic sweetened groups seems to be related to an increase in energy efficiency. Non-energetic sweeteners were proposed to disrupt the energetic signal associated with sweetness, and thus its failure to provide energy content may consequently enhance energy deposition through triggering the starvation mode, which is known to reduce energy expenditure and increase in energy efficiency⁽⁴⁴⁾. Indeed, it was widely suggested that eating sweet non-energetic substances brings the

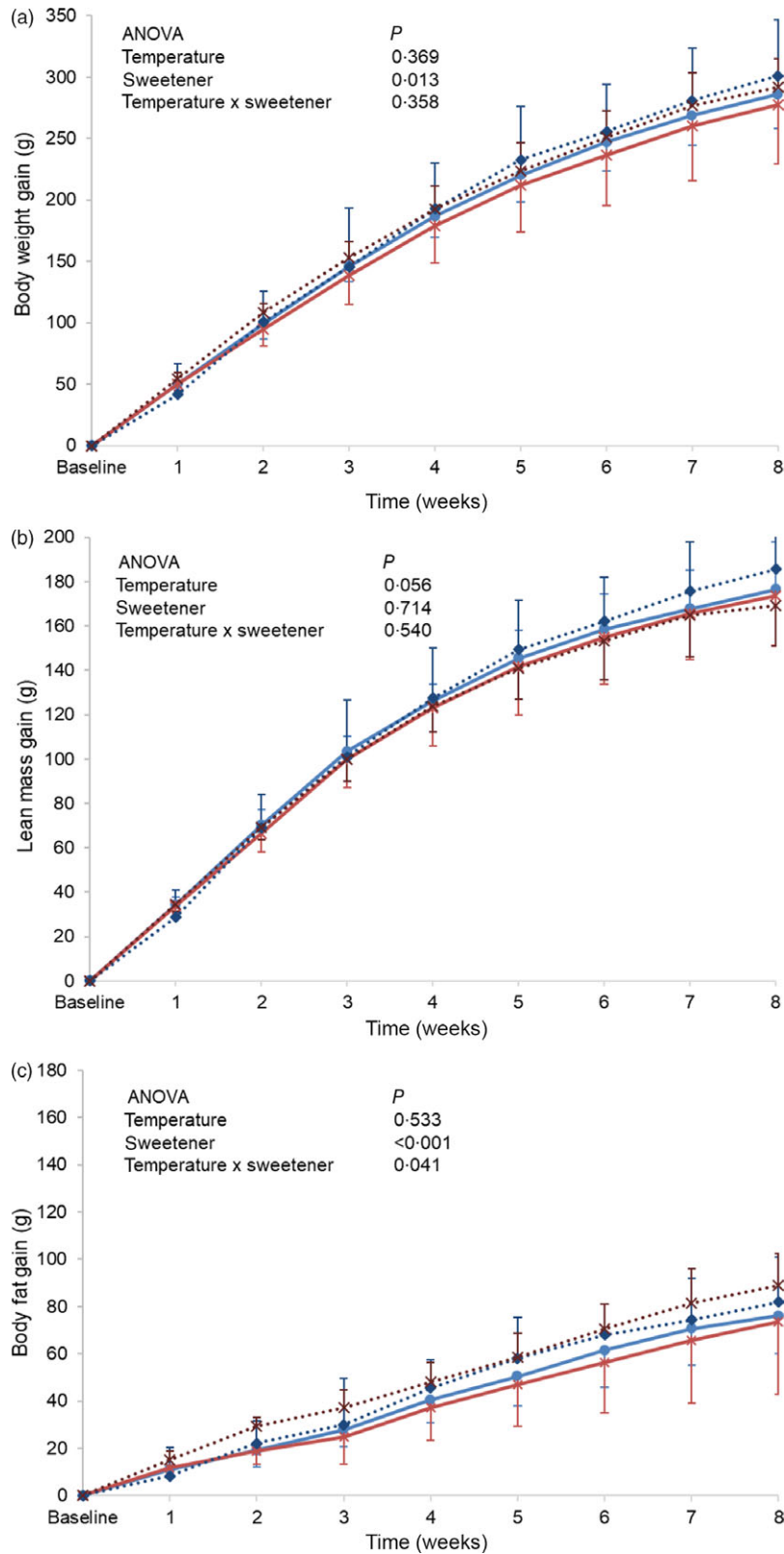


Fig. 2. Expt 2 – Effect of cold and room-temperature drinking sweetened (energetic and non-energetic) water on body weight measures. Weekly body weight (a), lean body mass (b) and body fat (c) gain from baseline in grams of the four groups of rats over the 8-week experimental period. Group CSu: cold water (about 4°C) sweetened with 10 % sucrose; Group NSu: room-temperature water (about 22°C) sweetened with 10 % sucrose; Group CAK: cold water (about 4°C) sweetened with 0.05 % acesulfame K; Group NAK: room-temperature water (about 22°C) sweetened with 0.05 % acesulfame K. Data are expressed as mean values and standard deviations of all values. A multiple-way ANOVA was performed with time, set as random, temperature of beverages and type of sweeteners as factors. Significance was set at $P < 0.05$. —●—, CSu; —●—, NSu; ...●..., CAK; ...●..., NAK.

Table 3. Expt 2 – Effect of cold and room-temperature drinking sweetened (energetic and non-energetic) water on energy intake, expenditure and efficiency and some organ weights (Wt) (Mean values and standard deviations)*

	CSu		NSu		CAk		NAk		P		
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Cold	Sweet	Cold × Sweet
Mean energy intake (kJ/d)	353.43	26.18	371.98	43.11	366.53	29.54	354.30	13.86	0.782	0.841	0.185
Energy from the diet (kJ/d)	318.18	22.35	281.24	34.14	366.53	29.54	354.30	13.86	0.018	<0.001	0.215
Energy from the fluid (kJ/d)	35.25	11.00	90.75	29.50	00.00	00.00	00.00	00.00	<0.001	<0.001	<0.001
Mean fluid intake (g/d)	37.91	7.38	62.43	18.43	44.53	10.56	48.46	7.13	0.004	0.421	0.031
Sweetened water intake (g/d)	21.05	6.57	54.20	17.62	29.24	8.93	40.00	7.84	<0.001	0.488	0.015
Plain water intake (g/d)	16.86	2.61	8.23	1.83	15.29	2.67	8.46	2.26	<0.001	0.447	0.309
Mean energy efficiency (kJ/100 kJ)	18.81	2.29	17.11	4.20	19.36	2.66	21.18	2.19	0.956	0.049	0.128
Mean energy expenditure (kJ/d)	286.62	19.36	307.30	29.88	294.92	17.51	278.99	10.08	0.763	0.209	0.026
Liver wet wt (g)	14.01	1.96	13.84	2.98	14.42	2.74	13.26	1.28	0.450	0.921	0.578
Liver % (g/100 g body wt)	3.12	0.28	3.13	0.40	3.12	0.37	2.93	0.28	0.468	0.448	0.432
Liver water (%)	72.41	1.98	71.64	1.71	69.33	5.70	69.73	1.99	0.870	0.039	0.613
Liver fat in wet (%)	3.57	1.36	3.62	0.82	5.65	2.63	5.64	1.84	0.972	0.004	0.965
Epididymal adipose tissue wt (g)	7.81	2.15	7.48	2.90	8.10	2.48	8.29	0.95	0.928	0.515	0.754

Group CSu, cold water (about 4°C) sweetened with 10% sucrose; Group NSu, room-temperature water (about 22°C) sweetened with 10% sucrose; Group CAk, cold water (about 4°C) sweetened with 0.05% acesulfame K; Group NAk, room-temperature water (about 22°C) sweetened with 0.05% acesulfame K.

* A two-way ANOVA was performed with time and temperature of the beverages as factors. Significance was set at $P < 0.05$.

Table 4. Expt 2 – Effect of cold and room-temperature drinking sweetened (energetic and non-energetic) water on plasma metabolites (Mean values and standard deviations)

	CSu		NSu		CAk		NAk		P		
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Cold	Sweet	Cold × Sweet
Glucose (mmol/l)	14.26	4.40	16.32	6.39	11.81	1.49	13.98	3.67	0.222	0.168	0.973
Insulin (ng/ml)	1.76	1.43	1.26	0.73	1.20	0.94	2.09	1.72	0.421	0.932	0.067
Total-cholesterol (mmol/l)	1.53	0.13	1.60	0.35	1.66	0.45	1.65	0.27	0.809	0.468	0.747
HDL-cholesterol (mmol/l)	1.39	0.07	1.41	0.20	1.43	0.19	1.46	0.12	0.751	0.420	0.928
TAG (mmol/l)	0.52	0.08	0.58	0.15	0.45	0.08	0.54	0.10	0.062	0.195	0.715
Albumin (g/l)	42.86	3.53	46.25	2.25	42.67	3.20	42.50	3.78	0.195	0.116	0.154
Plasma urea nitrogen (mmol/l)	5.05	0.60	4.78	0.60	5.42	0.47	5.54	0.60	0.723	0.015	0.372
Creatinine (μmol/l)	30.32	4.73	28.74	4.21	30.95	4.84	29.84	6.58	0.495	0.658	0.904

Group CSu, cold water (about 4°C) sweetened with 10% sucrose; Group NSu, room-temperature water (about 22°C) sweetened with 10% sucrose; Group CAk, cold water (about 4°C) sweetened with 0.05% acesulfame K; Group NAk, room-temperature water (about 22°C) sweetened with 0.05% acesulfame K.

* A two-way ANOVA was performed with time and temperature of the beverages as factors. Significance was set at $P < 0.05$.

sweet taste without the energy content, and animals use sweet taste to predict the energetic contents of food which will create a positive energy balance through increased food intake and/or diminished energy expenditure⁽⁴⁴⁾. Our findings come in opposition to popular beliefs about non-energetic sweeteners. Artificial sweeteners have been widely used as a substitution to sugar, providing the sweet taste without any energy load; this results in short-term weight and fat reduction^(21,22,45). However, long-term studies have associated them with weight gain^(24,26,44,46). Indeed, in rats, the consumption of foods or fluids containing a non-nutritive sweetener, saccharin, as compared with glucose, has led to an increase in food intake, body weight and body fat accumulation⁽⁴⁷⁾. Additionally, the weight gain of rats maintained on non-energetic sweetened (acesulfame K or saccharin) yogurt was higher than that kept on glucose sweetened yogurt⁽⁴⁸⁾. In line, we found higher body fat content among the non-energetic (acesulfame K) sweetened groups.

It is worth noting that the volume of ingested cold sweetened (energetic and non-energetic) water was smaller than that of the normal temperature water. Despite the high contribution of

sucrose-sweetened water (about 10% cold group and 24% room-temperature group) to energy intake, no significant differences were observed in total daily energy intake, as well as in plasma insulin, glucose and lipid profile. These similarities may imply that the observed differences in body composition were not related to changes in insulin sensitivity. The capacity of sucrose ingestion to increase postprandial energy expenditure⁽¹²⁾ may have partially explained the observed difference in energy expenditure between the sucrose sweetened groups, and a strong association was found between the amount of sucrose consumed from fluid and energy expenditure ($r = 0.761$, $P = 0.001$). On the other hand, increased plasma urea nitrogen of the non-energetic sweeteners groups may be related to their higher dietary intake of protein as compared with that of sucrose sweetened groups.

Conclusion

In conclusion, internal variation in body temperature by the manipulation of the temperature of drinking water was found

to affect body composition. Ingestion of cold water had a favourable effect on lean body mass accompanied by a decrease in fat mass gain. These changes in body composition were associated with an increase in energy expenditure, while both energy intake and efficiency were not affected. On the other hand, non-energetic sweetened (0.05 % acesulfame K) water was able to increase body fat gain and energy efficiency as compared with the energetically sweetened (10 % sucrose) water, while energy intake and expenditure were not affected. Our findings show that the impact of internal cold exposure is different from that of external cold exposure, especially in relation to energy intake. Further studies are required to explore the mechanisms for the adaptation to the ingestion of cold water.

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The authors declare that there are no conflicts of interest.

Supplementary material

For supplementary material referred to in this article, please visit <https://doi.org/10.1017/S0007114520003359>

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