

Effect of sugar fatty acid esters on rumen fermentation in vitro

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1. The effect of sugar fatty acid esters (SFEs; currently used as food additives for human consumption) on rumen volatile fatty acids (VFA) and gas production was studied with sheep rumen contents in vitro.

2. Some SFEs having monoester contents of more than 70% increased the molar proportion of propionate in conjunction with reduction in the acetate:propionate ratio when the individual SFE was added to rumen contents in a final concentration of 4 g/l. Laurate sugar ester was the most potent propionate enhancer and rumen gas depressor, the effective dose being as low as 1 g/l in a final concentration. Fatty acid esters other than SFEs had little, if any, effect on rumen VFA production and their molar proportions.

3. Approximately 50% of laurate sugar ester was hydrolysed by in vitro incubation with rumen fluid for 2 h. The addition of fatty acids and sucrose was also effective in the alterations of rumen VFA and gas production. However, the effect of SFEs on in vitro rumen fermentation was significantly greater than that of their constituent fatty acids or sucrose, or both. Accordingly, the effect appeared to be ascribed to the complex action of SFE itself and to its constituents, free fatty acids and sucrose.

4. SFEs, at the level of 4 g/l, reduced substantially the froth formation (ingesta volume increase) and seemed to be effective for the prevention of bloat.

Sugar fatty acid esters (SFEs), synthesized from sucrose and long-chain fatty acids, usually derived from tallow, are mixtures composed of different amounts of monoesters and polyesters (Table 1). Most of these non-ionic surfactants contain mono-, di- and triesters in varying proportions and have a wide hydrophile—lipophile balance ranging from 1 to 15. Hence, SFEs are currently used as additives for various foods such as ice cream, several cream products and sweets because of their excellent emulsifying capability.

Extensive studies have been conducted to improve the efficiency of rumen fermentation by controlling the number of microbes and their activities with chemical agents (Chalupa, 1977). Of these chemical agents ionophore antibiotics (Bergen & Bates, 1984) and methane inhibitors (Czerkawski & Breckenridge, 1975; Stanier & Davies, 1981; Davies *et al.* 1982; Chalupa *et al.* 1983) can be used to increase energy and protein utilization in ruminants. Effects of fatty acids (Blaxter & Czerkawski, 1966; Demeyer & Henderickx, 1967; Chalupa *et al.* 1983; Jenkins & Palmquist, 1984), oils (Blaxter & Czerkawski, 1966; McAllan *et al.* 1983; Sutton *et al.* 1983) or their calcium soaps (Jenkins & Palmquist, 1984) on rumen fermentation are also of interest. Little is known about the effect of SFEs on rumen fermentation.

The present study was undertaken to examine the effect of SFEs on rumen volatile fatty acids (VFA) and gas production in vitro. A preliminary report of the present study has appeared (Wakita *et al.* 1985).

MATERIALS AND METHODS

SFEs and other fatty acid esters

The compositions of the esters are shown in Table 1. Most of the SFEs (nos. 1–8) consisted of stearate and palmitate esters with different degrees of esterification, and the rest (nos. 9 and 10) contained principally behenate, laurate or oleate instead of stearate and palmitate esters. Other types of fatty acid esters (nos. 12–15) were used for comparison with SFEs.

* For reprints.

Table 1. Sugar fatty acid esters (SFEs; nos. 1-11) and other fatty acid esters (nos. 12-15) used in the experiment

No. of ester	Fatty acid moiety (%)			Ester composition (%)		
	Stearic	Palmitic	Monooester	Di- and triesters		
1	70	30	2		98	
2	70	30	20		80	
3	70	30	30		70	
4	70	30	55		45	
5	70	30	70		30	
6	70	30	75		25	
7	30	70	70		30	
8	30	70	75		25	
9	Behenic 65, stearic 23, arachidonic 5, palmitic 1, other 6*			18		82
10	Lauric 70, myristic 13, palmitic 6, caproic 4, caprylic 4, other 3*			70		30
11	Oleic 73, linoleic 10, palmitoleic 7, palmitic 4, stearic 1, myristoleic 1, other 4*			70		30
12	Propylene glycol ester (C ₁₀₋₁₈)					
12	Glycerin ester (C ₁₄₋₁₈)			80-95		5-20
14	Sorbitan ester (C ₁₂₋₁₈)			90-95		5-10
15	Polyoxyethylene sorbitan monolaurate					Unknown
						Unknown

* Unknown fatty acids.

All the ester compounds were obtained from Mitsubishi Kasei Industry Co. (Yokkaichi, Japan).

Rumen fluid, incubation, and analysis of products

Samples of rumen contents were taken before the morning feed from sheep (fitted with rumen fistulas) which were given feed in two equal meals (at 09.00 and 19.00 hours) of 350 g lucerne (*Medicago sativa*) hay-cubes and 200 g concentrate feed. Rumen samples were strained through two layers of surgical gauze and used within 30 min of sampling.

To determine the effect of esters on VFA production by rumen contents *in vitro*, the strained rumen fluid (10 ml) was pipetted into a 25 ml test-tube containing 0.22 g finely-ground feed (a mixture of 7 parts lucerne hay-cubes and 4 parts concentrate). Each ester compound listed in Table 1 was dissolved or suspended in distilled water to give a concentration of 44 mg/ml and in most experiments 1 ml of this solution was added to the test-tube so that the final concentration of ester was 44 mg/11 ml. After replacing the air in the test-tube with mixed gas (nitrogen-carbon dioxide; 95.5, v/v) the tube was closed with a rubber stopper and incubated in a water-bath at 39° for 2 h with occasional mixing to prevent the layering of feed particles. Except for the omission of ester compounds, the control tube was prepared and incubated in the same manner. After 2 h incubation the reaction mixture was mixed with 1 ml 2.0 M-sulphuric acid, followed by centrifugation. A portion (5 ml) of the supernatant fraction was mixed with 1 ml 2.5 M-sulphuric acid containing *m*-phosphoric acid (200 g/l), allowed to stand overnight, and then centrifuged. The resulting supernatant fraction was used for VFA determination by gas-liquid chromatography as described by Suto (1973). The VFA production during 2 h incubation was calculated by subtracting VFA concentration in the blank tube, in which the reaction was stopped just before incubation, from that in the experimental tube.

In the determination of gas production 10 ml of the strained rumen fluid was incubated for 1 h at 39° in reaction vessels consisting of glass tubes (15 mm in diameter and 80 mm in length) fitted with rubber balloons at one end. The tube was charged with the same feed (0.22 g) and an ester (4 g/l) as for the VFA incubations. After expelling the air in the reaction vessel by squeezing the balloon and thus pushing the reaction mixture up, the top was closed tightly with a rubber stopper and the whole immersed in a water-bath at 39°. After 1 h incubation, 1 ml 2.0 M-sulphuric acid was injected through the stopper and the resulting gas phase was separated from the reaction mixture by injecting 1 ml liquid paraffin. Samples of gas (0.5 ml) at 39° were taken out with a 2.5 ml gas-syringe and the composition was analysed by a gas-liquid chromatographic procedure (Ushida *et al.* 1982). The remaining gas in the vessel was withdrawn into an air-tight 10 ml syringe to measure total gas volume at 39°. CO₂ and methane volumes were calculated by multiplying a total gas volume by the component percentage of individual gas. The gas production during 1 h incubation was calculated by a method similar to the calculation of VFA production.

The ability of rumen contents to froth was assessed by measurement of the ingesta volume increase (IVI) carried out as described by Jacobson *et al.* (1957), except that in the present experiment 50 ml rumen fluid were used, instead of 200 ml in the original method with cattle. The rumen fluid (50 ml in a 100 ml graduated cylinder) was allowed to stand at 39° in a water-bath for 45 min and the increase in volume was recorded and presented as percentage increase.

Degradation of ester. The concentration of SFE no. 10 was estimated by the colorimetric method of Hodge & Hofreiter (1962) which gave a linear calibration curve between optical density and sucrose contents in SFE molecules. Assuming that sucrose liberated from SFE is fermented rapidly by rumen microbes, the method could give an approximate amount of undegraded SFE. The procedure was as follows. The strained rumen fluid was incubated with feed and with or without SFE as for VFA incubations. Every 30 min 1 ml 1.0 M-

sulphuric acid was added to a tube followed by centrifugation; the supernatant fraction was kept. A portion of the supernatant fraction (1 ml) was mixed with 1 ml phenol solution (50 g/l) and 5 ml 18 M-sulphuric acid, mixed well and allowed to stand at room temperature for 30 min. The optical density of the developed yellow colour was determined at 490 nm with a spectrophotometer. The degradability of the ester was calculated by making corrections for blank values.

The results were analysed statistically by Dunnett's *t* statistic (Dunnett, 1955).

RESULTS

Effects of SFEs on rumen VFA production in vitro

Tables 2 and 3 show total VFA production and their composition in rumen fluid incubated with SFE (4 g/l) or without SFE (control) *in vitro*. Of six SFEs containing 70% stearate and 30% palmitate moieties (Table 1, nos. 1–6), some SFEs (nos. 5 and 6) increased total VFA concentration and propionate proportion in conjunction with a decrease in acetate proportion, but the rest of these SFEs (nos. 1–4) had little effect on propionate proportion (Table 2). SFEs containing 70% palmitate and 30% stearate moieties (nos. 7 and 8) altered total VFA production and their molar proportion in the same pattern as the above SFEs (nos. 5 and 6) did, the differences in VFA molar proportion and acetate:propionate (A:P) ratio from control values being significant (Table 2).

SFEs having 70% monoester of different fatty acid moieties (nos. 10 and 11) also increased the molar proportion of propionate and decreased the A:P ratio (Table 3), while the SFE having 18% monoester (no. 9) had no effect on propionate proportion. This effect of SFEs appeared to be closely associated with the monoester content in the products, since SFEs containing more than 70% monoester increased VFA, in particular propionate proportion, but those containing less than 55% monoester had little effect on VFA molar proportion (Tables 2 and 3). Among the effective SFEs having the same ester composition (70% monoester and 30% di- and triester, nos. 5, 7, 10 and 11), laurate ester (no. 10) was the most potent propionate enhancer, followed by nos 7, 5 and 11 (Tables 2 and 3). Of two SFE pairs with the same ester composition (nos. 6 and 8, and nos. 5 and 7), palmitate ester tended to be more effective than stearate ester (Table 2).

Because of the large effect of laurate ester (no. 10), the effect of increasing doses of the ester on total VFA and propionate production was examined but not reported here in detail. The additions of the ester ranging from 0.1 to 0.8% in a final concentration significantly increased propionate molar proportion with a concurrent decrease of acetate proportion in a dose-response manner, but the additions had little effect on total VFA production.

The addition of fatty acid esters at 4 g/l other than SFEs (nos. 12–15) caused no significant effect on total VFA production and their relative proportions, except for valerate.

Degradation rate of SFEs and effects of the structural fatty acids or sucrose, or both, on VFA production in vitro

The apparent degradability of SFE no. 10 is shown in Table 4. The ester was degraded gradually during incubation and 56% of the added ester seemed to be degraded after 2 h. If the SFE or products inhibit the degradation of endogenous sugar, then the true degradation of SFE is greater than the apparent degradation as given in Table 4.

An attempt was made to determine whether the effect of SFEs is due to their structural fatty acids or sucrose, or both (see Table 5). The amounts of sucrose and fatty acids added to the reaction mixture was similar to those in the corresponding SFE (4 g/l) as described

Table 2. Effect of sugar fatty acid esters (SFEs; nos. 1-8) on rumen total volatile fatty acids (VFA) production, their percentage composition and acetate: propionate (A:P) ratio when sheep rumen fluid was incubated *in vitro* with or without SFE in a final concentration of 4 g/l (Values are means with their standard errors for seven observations)

SFE no. †	Total VFA (mmol/2 h)		Acetic		Propionic		Butyric		iso-Valeric		n-Valeric		A:P	
	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
Control	66.5	1.2	65.1	0.5	23.3	0.4	8.4	0.3	0.9	0.1	2.3	0.3	2.79	0.06
1	68.4	2.0	64.0	0.6	23.5	0.4	8.7	0.3	1.0	0.1	2.8	0.3	2.72	0.07
2	65.0	3.6	64.6	0.8	23.5	0.3	8.2	0.2	1.0	0.1	2.7	0.4	2.75	0.06
3	72.8	2.3	65.1	1.0	23.5	1.4	7.5**	0.2	0.8	0.1	3.1	0.3	2.77	0.03
4	75.4	2.3	64.8	0.3	23.5	0.2	7.4***	0.2	0.8	0.0	3.5*	0.1	2.76	0.04
5	75.6	2.1	61.7***	0.3	26.2***	0.3	7.4***	0.2	0.9	0.1	3.8***	0.2	2.35***	0.03
6	76.6*	1.7	61.6***	0.5	26.7***	0.2	7.1***	0.2	0.8	0.1	3.8***	0.2	2.31***	0.03
7	69.7	3.0	59.4***	0.5	29.3***	0.6	7.8	0.2	1.0	0.1	2.5	0.3	2.03***	0.05
8	70.0	2.8	58.4***	0.6	30.4***	0.5	7.4***	0.2	1.1	0.1	2.7	0.4	1.92***	0.05

Mean values were significantly different from control value: * $P < 0.05$, ** $P < 0.02$, *** $P < 0.01$.
 † For details, see Table 1.

Table 3. Effect of sugar fatty acid esters (SFEs; nos. 8-11) and other fatty acid ester (nos. 12-15) on rumen total volatile fatty acids (VFA) production, their percentage composition and acetate: propionate (A:P) ratio when sheep rumen fluid was incubated *in vitro* with or without fatty acid ester in a final concentration of 4 g/l

(Values are means with their standard errors for eight observations)

Fatty acid ester no. †	Total VFA (mmol/2 h)		Acetic		Propionic		Butyric		iso-Valeric		n-Valeric		A:P	
	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE	Mean	SE
Control	75.1	2.4	63.5	0.8	22.7	0.7	10.3	0.4	1.1	0.1	2.4	0.1	2.85	0.07
8	76.6	1.4	59.1***	0.4	30.4***	0.7	8.5	0.3	0.9	0.1	1.1***	0.1	1.93***	0.06
9	73.2	2.1	63.4	0.8	22.5	0.7	10.3	0.4	1.3	0.0	2.5	0.1	2.81	0.05
10	75.8	1.7	57.6***	1.1	29.7***	0.5	7.2***	0.3	1.4***	0.1	3.8***	0.1	1.93***	0.05
11	74.7	1.5	60.8	1.1	23.7	0.6	9.4	0.4	1.9***	0.1	4.3***	0.2	2.57***	0.06
12	73.7	1.5	63.3	0.8	22.6	0.7	10.3	0.4	1.0	0.1	2.9***	0.1	2.80	0.05
13	73.3	2.1	64.0	1.1	22.5	0.6	10.2	0.4	1.0	0.1	2.3	0.0	2.85	0.06
14	72.1	1.7	63.6	0.9	22.9	0.7	10.0	0.5	1.1	0.0	2.3	0.1	2.74	0.05
15	73.6	1.5	63.2	0.9	22.4	0.6	10.1	0.4	1.8***	0.1	2.4	0.1	2.80	0.07

Means values were significantly different from control value; *** $P < 0.01$.

† For details, see Table 1.

Table 4. *Degradability of sugar fatty acid ester (SFE; no. 10*)*

(Values are means with their standard errors for three observations)

Incubation period (min)	Phenol-sulphuric acid active substances (mg/ml)				Difference (D)	Degradability of SFE (4.05 - D)/4.05
	SFE (4 mg/ml)		Blank			
	Mean	SE	Mean	SE		
0	13.16	0.13	9.11	0.22	4.05	0
30	7.20	0.09	4.17	0.30	3.03	0.25
60	6.66	0.18	4.14	0.08	2.53	0.38
90	6.15	0.10	4.03	0.05	2.12	0.48
120	5.70	0.08	3.93	0.06	1.77	0.56

* For details, see Table 1.

in Table 5. All additives slightly increased total VFA production in comparison with the control. Some additives (except fatty acids) increased propionate and decreased acetate and A:P ratio, a maximum effect being observed with SFE additives (Table 5). The result suggests that sucrose may serve as a precursor for VFA production and that fatty acids have some effect on VFA production, but the effect of SFEs is not due completely to their hydrolysates *per se*, because there were significant differences between the effects of the SFEs and their constituents (Table 5).

Effects of SFEs on rumen gas production and IVI

Effects of two SFEs (nos. 7 and 10) and their constituents on gas production are shown in Table 6. Both SFEs depressed gas production, particularly methane production, with no accumulation of hydrogen (Table 6). The inhibitory effect of SFE no. 7 on rumen gas production was more potent than that of its structural fatty acids or fatty acids plus sucrose, but the effect of SFE no. 10 was less potent than that of its fatty acids or combinations of fatty acids and sucrose (Table 6).

The effect of SFEs on IVI is shown in Table 7. The addition of SFE (nos. 7 and 10) inhibited froth formation in rumen contents during incubation in comparison with the control.

DISCUSSION

Our results demonstrate that some SFEs stimulated rumen VFA production, in particular propionate production, and inhibit gas production which could lead to the reduction of the IVI value of rumen contents. Based on the potency of SFEs inducing these changes, it seems likely that monoester contents over 70% are required for these changes to take place (Tables 2 and 3), and that the SFE effects are associated with the intact SFE molecule as a surfactant and its constituents, fatty acids and sucrose. The latter view appears to be supported by the differential effects between SFEs and their constituents (Tables 5 and 6) and the degradation rate of SFE (Table 4).

It is well documented that free fatty acids, particularly higher unsaturated fatty acids, alter rumen fermentation towards more propionate and less methane (Blaxter & Czerkawski, 1966; Demeyer & Henderickx, 1967; Chalupa *et al.* 1984). Several mechanisms by which the acids inhibit methanogenesis have been suggested, e.g. a direct toxic effect towards methanogenic bacteria (Prins *et al.* 1972) and a physico-chemical inhibition, involving adhesion of the acids to the cell, possibly uncoupling energy systems (Galbraith

Table 5. Effect of sugar fatty acid esters (SFEs; nos. 7 and 10†) and their constituents on rumen total volatile fatty acids (VFA) production, their percentage composition and acetate: propionate (A:P) ratio when sheep rumen fluid was incubated *in vitro* with or without SFE or its constituents

(Values are means with their standard errors for eight observations)

Additive	Total VFA (mmol/2 h)		Acetic acid (%)		Propionic acid (%)		A:P	
	Mean	SE	Mean	SE	Mean	SE	Mean	SE
Control (no additive)	61.1	2.8	64.1	2.2	23.1	1.6	2.77	0.04
SFE no. 7								
SFE § (4.00 mg/ml)	65.8	1.3	58.9***	1.7	29.1***	0.5	2.02***	0.03
Sucrose (1.99 mg/ml) + fatty acids	69.3**	0.6	59.0***	0.3	25.3***†††	0.2	2.33***†††	0.02
Sucrose (1.99 mg/ml)	72.3***	3.1	59.4***	0.7	25.1***†††	0.3	2.37***†††	0.03
Fatty acids	62.3	1.5	60.3†††	1.0	23.8†††	0.7	2.53***†††	0.12
SFE no. 10								
SFE ¶ (4.00 mg/ml)	77.7***	1.2	58.8***	0.3	29.7***	0.5	1.98***	0.07
Sucrose (2.30 mg/ml) + fatty acids ††	83.7***	1.2	58.8***	0.4	23.8†††	0.1	2.47***†††	0.01
Sucrose (2.30 mg/ml)	72.0***	1.1	59.6***	0.5	29.7***†††	0.5	2.36***†††	0.01
Fatty acids ††	71.5***	0.8	62.5†††	0.2	22.3†††	0.1	2.80†††	0.02

In each column differences from control values were significant: * $P < 0.05$, ** $P < 0.02$, *** $P < 0.01$.

In each column within no. 7 or no. 10 SFE group, differences from SFE additives were significant: ††† $P < 0.01$.

† For details, see Table 1.

§ No. 7 SFE (4.00 mg) contained (mg) sucrose 1.99, stearic acid 0.65, palmitic acid 1.36.

|| The same amounts of sodium stearate and palmitate as those in 4.00 mg of no. 7 SFE were added to 1 ml of the reaction mixture.

¶ No. 10 SFE (4.00 mg) contained (mg) sucrose 2.30, lauric acid 1.23, myristic acid 0.26, palmitic acid 0.13, caproic acid 0.04, caprylic acid 0.04.

†† The same amounts of sodium laurate, myristate and palmitate, and of caproic and caprylic acids as those in 4.00 mg of no. 10 SFE were added to 1 ml of the reaction mixture.

Table 6. Effect of sugar fatty acid esters (SFEs; nos. 7 and 10†) and their constituents on rumen gas production (ml/h per 10 ml rumen fluid) when sheep rumen fluid was incubated with or without SFE or its constituents

(Values are means with their standard errors for three observations)

Additive	Total gas		CO ₂		CH ₄		H ₂
	Mean	SE	Mean	SE	Mean	SE	
Control (no additive)	3.5	0.1	2.9	0.1	0.6	0.0	Tr
SFE no. 7							
SFE§ (4.00 mg/ml)	2.6	0.6	2.3	0.5	0.3***	0.0	Tr
Sucrose (1.99 mg/ml) + fatty acids	3.4	0.2	2.8	0.2	0.6†††	0.0	Tr
Fatty acids	3.3	0.3	2.7	0.4	0.6†††	0.0	Tr
SFE no. 10							
SFE¶ (4.00 mg/ml)	2.3*	0.1	2.0	0.0	0.3***	0.0	Tr
Sucrose (2.30 mg/ml) + fatty acids ††	1.7***	0.1	1.5**	0.1	0.2***†††	0.0	Tr
Fatty acids ††	1.4***	0.2	1.1***	0.1	0.3***	0.0	Tr

Tr, trace.

In each column differences from control values were significant: **P* < 0.05, ***P* < 0.02, ****P* < 0.01.

In each column within no. 7 or no. 10 SFE group, differences from SFE additives were significant: †††*P* < 0.01.

† For details, see Table 1.

§ No. 7 SFE (4.00 mg) contained (mg) sucrose 1.99, stearic acid 0.65, palmitic acid 1.36.

|| The same amounts of sodium stearate and palmitate as those in 4.00 mg of no. 7 SFE were added to 1 ml of the reaction mixture.

¶ No. 10 SFE (4.00 mg) contained (mg) sucrose 2.30, lauric acid 1.23, myristic acid 0.26, palmitic acid 0.13, caproic acid 0.04, caprylic acid 0.04.

†† The same amounts of sodium laurate, myristate and palmitate, and of caproic and caprylic acids as those in 4.00 mg of no. 10 SFE were added to 1 ml of the reaction mixture.

Table 7. Effect of sugar fatty acid esters (SFEs; nos. 7, 10 and 11) on rumen ingesta volume increase (IVI; %) when sheep rumen fluid was incubated in vitro with or without SFE in a final concentration of 4 g/l

(Values are means with their standard errors for three observations)

SFE no. †	IVI during:					
	15 min		30 min		45 min	
	Mean	SE	Mean	SE	Mean	SE
Control	10.6	0.7	19.7	0.9	27.3	1.3
7	4.7***	0.3	7.9***	0.6	11.3***	0.7
10	4.7***	0.7	7.0***	0.6	11.3***	0.7
11	4.2***	0.1	6.7***	0.3	9.3***	0.3

In each column differences from control values were significant: ****P* < 0.01.

† For details, see Table 1.

& Miller, 1973). Because of active esterases in rumen fluid (Table 4; Czerkawski & Breckenridge, 1975), fatty acids and sucrose released from SFEs can partly contribute to the alteration of rumen fermentation towards more propionate and less methane.

With higher fatty acids and oils, the increase in molar proportion of propionate is associated with depression of fibre digestion (Blaxter & Czerkawski, 1966; McAllan *et al.*

1983; Sutton *et al.* 1983; Jenkins & Palmquist, 1984). Long-chain fatty acids inhibit the growth of certain rumen bacteria (Henderson, 1973), the growth of cellulolytic species being markedly inhibited by the addition of oleic acid to the culture medium (Maczulak *et al.* 1981). In contrast, tallow calcium soaps do not lower digestibilities of fibre in the rumen (Jenkins & Palmquist, 1984). Assuming that 40% SFEs remain intact in the rumen for 2 h (Table 4), it is possible that SFEs as well as tallow soaps may have a far less inhibitory effect on fibre digestion in the rumen than free fatty acids alone.

It has been shown that the addition of non-ionic surfactants (sucrose monostearate and sucrose monopalmitate) to fungal cultures results in marked increases in yields of amylase (Takahashi *et al.* 1960), cellulase, sucrase, xylanase and glucanase (Reese & Maguire, 1969), the action appearing to be an effect of the surfactant on cell permeability. The undegraded SFEs could act as a surfactant on rumen microbes in the same way, and stimulate microbial uptake of nutrients by emulsifying fermentation medium and by a possible alteration of permeability of cell membranes, which in turn could enhance VFA production. However, the detailed mechanisms of SFE action, including selection for rumen microbes, are uncertain and must await elucidation.

In view of the marked IVI reducing activity of SFEs, they could be useful in the treatment of bloat in ruminants. In fact, several surfactants have been widely used for the prevention or treatment of both legume and grain bloats (Clarke & Reid, 1974; Laby, 1975).

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REFERENCES

- Bergen, D. G. & Bates, D. D. (1984). *Journal of Animal Science* **58**, 1465–1483.
- Blaxter, K. L. & Czerkawski, J. (1966). *Journal of the Science of Food and Agriculture* **17**, 417–421.
- Chalupa, W. (1977). *Journal of Animal Science* **46**, 585–599.
- Chalupa, W., Patterson, J. A., Parish, R. C. & Chow, A. W. (1983). *Journal of Animal Science* **57**, 186–194.
- Chalupa, W., Rickabaugh, B., Kronfeld, D. S. & Sklan, D. (1984). *Journal of Dairy Science* **67**, 1439–1444.
- Clarke, R. T. J. & Reid, C. S. W. (1974). *Journal of Dairy Science* **57**, 753–785.
- Czerkawski, J. W. & Breckenridge, G. (1975). *British Journal of Nutrition* **34**, 429–446.
- Davies, A., Nwaonu, H. N., Stanier, G. & Boyle F. T. (1982). *British Journal of Nutrition* **47**, 565–576.
- Demeyer, D. I. & Henderickx, H. K. (1967). *Biochimica et Biophysica Acta* **137**, 484–497.
- Dunnnett, C. W. (1955). *American Statistical Association Journal* **50**, 1096–1121.
- Galbraith, H. & Miller, T. B. (1973). *Journal of Applied Bacteriology* **36**, 659–675.
- Henderson, C. (1973). *Journal of Agricultural Science, Cambridge* **81**, 107–112.
- Hodge, J. E. & Hofreiter, B. T. (1962). In *Methods in Carbohydrate Chemistry*, vol. 1, pp. 380–394. [R. L. Whistler and M. L. Wolfrom, editors]. New York: Academic Press.
- Jacobson, D. R., Lindahl, I. L., McNeill, J. J., Shaw, J. C., Doetsch, R. N. & Davis, R. E. (1957). *Journal of Animal Science* **16**, 515–524.
- Jenkins, T. C. & Palmquist, D. L. (1984). *Journal of Dairy Science* **67**, 978–986.
- Laby, R. H. (1975). In *Digestion and Metabolism in the Ruminant*, pp. 537–550. [I. W. McDonald and A. C. I. Warner, editors]. Armidale NSW 2351. Australia: The University of New England Publishing Unit.
- McAllan, A. B., Knight, R. & Sutton, J. D. (1983). *British Journal of Nutrition* **49**, 433–440.
- Maczulak, A. E., Dehority, B. A. & Palmquist, D. L. (1981). *Applied and Environmental Microbiology* **42**, 856–862.
- Prins, R. A., Van Nevel, C. J. & Demeyer, D. I. (1972). *Antonie van Leeuwenhoek Journal of Microbiology and Serology* **38**, 281–287.
- Reese, E. T. & Maguire, A. (1969). *Applied Microbiology* **17**, 242–245.
- Stanier, G. & Davies, A. (1981). *British Journal of Nutrition* **45**, 567–578.
- Suto, T. (1973). In *Methods in the Clinical Examination of the Bovine*, pp. 39–42 [R. Nakamura, T. Yonemura and T. Suto, editors]. Tokyo: Nosan Gyoson Bunka Kyokai. (In Japanese).
- Sutton, J. D., Knight, R., McAllan, A. B. & Smith, R. H. (1983). *British Journal of Nutrition* **49**, 419–432.
- Takahashi, J., Abekawa, Y. & Yamada, K. (1960). *Nippon Nogei Kagaku Kaishi* **34**, 1043–1045.
- Ushida, K., Miyazaki, A. & Kawashima, R. (1982). *Japanese Journal of Zootechnical Science* **53**, 412–416.
- Wakita, M., Yamada, Y. & Hoshino, S. (1985). *Proceedings of the 3rd AAAP Animal Science Congress* **2**, 866–868.