

## A comparison of risk factors associated with community-associated methicillin-resistant and -susceptible *Staphylococcus aureus* infections in remote communities

G. R. GOLDING<sup>1</sup>, P. N. LEVETT<sup>2</sup>, R. R. McDONALD<sup>2</sup>, J. IRVINE<sup>3</sup>, M. NSUNGU<sup>4</sup>,  
S. WOODS<sup>4</sup>, A. HORBAL<sup>5</sup>, C. G. SIEMENS<sup>5</sup>, M. KHAN<sup>6</sup>, M. OFNER-AGOSTINI<sup>7</sup>,  
M. R. MULVEY<sup>1,5\*</sup> AND the Northern Antibiotic Resistance Partnership (NARP)

<sup>1</sup> National Microbiology Laboratory, Winnipeg, MB, Canada

<sup>2</sup> Saskatchewan Disease Control Laboratory, Regina, SK, Canada

<sup>3</sup> Population Health Unit, LaRonge, SK, Canada

<sup>4</sup> Northern Intertribal Health Authority, Prince Albert, SK, Canada

<sup>5</sup> University of Manitoba, Winnipeg, MB, Canada

<sup>6</sup> Kelsey Trail Health Region, Melfort, SK, Canada

<sup>7</sup> Public Health Agency Canada, Ottawa, ON, Canada

(Accepted 27 November 2009; first published online 22 January 2010)

### SUMMARY

In this case-control study, cases [community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA),  $n = 79$ ] and controls [community-associated methicillin-susceptible *S. aureus* (CA-MSSA),  $n = 36$ ] were defined as a laboratory-confirmed infection in a patient with no previous hospital-associated factors. Skin and soft tissue were the predominant sites of infection, both for cases (67·1%) and controls (55·6%). Most of the cases (79·7%) and controls (77·8%) were aged <30 years. Investigations did not reveal any significant statistical differences in acquiring a CA-MRSA or CA-MSSA infection. The most common shared risk factors included overcrowding, previous antibiotic usage, existing skin conditions, household exposure to someone with a skin condition, scratches/insect bites, and exposure to healthcare workers. Similar risk factors, identified for both CA-MRSA and CA-MSSA infections, suggest standard hygienic measures and proper treatment guidelines would be beneficial in controlling both CA-MRSA and CA-MSSA in remote communities.

**Key words:** Bacterial infections, bacterial typing, community outbreaks, methicillin-resistant *S. aureus* (MRSA), *Staphylococcus aureus*.

### INTRODUCTION

Methicillin-resistant *Staphylococcus aureus* (MRSA) is a leading nosocomial pathogen that was first reported in Canada in 1981 [1] and has since disseminated

nationwide [2, 3]. Over the last decade, community-associated MRSA (CA-MRSA) infections have rapidly emerged on a global scale in many aboriginal reserves, the military, intravenous drug users, the homeless, penitentiaries, amateur and professional sports teams, day-care centres, and schools [4–11]. Of great concern is that these CA-MRSA strains are: (1) causing infections in often young otherwise healthy individuals with no predisposing nosocomial risk

\* Author for correspondence: Dr M. R. Mulvey, National Microbiology Laboratory, 1015 Arlington St, Winnipeg, MB, R3E 3R2, Canada.  
(Email: Michael\_mulvey@phac-aspc.gc.ca)

factors [10], (2) associated with more severe disease [12–14], and (3) now entering and disseminating within hospitals [15, 16].

In the 1990s, CA-MRSA had been reported only sporadically in Canada [17, 18]. However, since that time two CA-MRSA strains have emerged in Canada, CMRSA7 (USA400) and CMRSA10 (USA300) [19]. CMRSA10 has emerged predominantly in the western provinces of Canada and high incidences have been reported in the homeless, incarcerated, and intravenous drug user populations in the Calgary [7] and Vancouver [5, 20] health regions. In comparison to the incidence of CA-MRSA infections in the large urban centres across Canada, which is represented through the ongoing efforts of the Canadian Nosocomial Infection Surveillance Program [2, 3], very little attention has been directed at the emerging problem of CA-MRSA in northern communities in Canada. CMRSA7 was first reported in Manitoba as an outbreak in the southern portion of that province in the late 1990s, but has since spread to the northern regions of the province between 2000 to 2004 [21, 22]. The emergence of CMRSA7 was thereafter seen in a central eastern Saskatchewan community adjacent to the Manitoba border [10] and has since disseminated into all three northern health regions of Saskatchewan [23] as well as Nunavut [24]. The emergence of CMRSA7 in many of these northern communities is causing primarily skin and soft tissue infections in children with crude rates of infection as high as 196 cases/10 000 in some northern Saskatchewan health regions [23].

The factors influencing the emergence and spread of CA-MRSA may be different in these northern communities compared to factors identified in studies involving large urban centres where cultural, socio-economic, medical, and environmental conditions are different. To better understand the dissemination of CA-MRSA in remote northern communities a case-control study was undertaken to identify potential risk factors associated with the acquisition and dissemination of CA-MRSA in two selected northern Saskatchewan health regions.

## METHODS

### Setting

The study was a prospective cohort of CA-MRSA and community-associated methicillin-susceptible *S. aureus* (CA-MSSA) infections identified from September 2004 to March 2007 within selected

communities of the Kelsey Trail Health Region (KTHR) and the Mamawetan Churchill River Health Region (MCRHR). The KTHR encompasses a large geographic area of east-central Saskatchewan, ~44 369 km<sup>2</sup>, and has an estimated population of 42 098 (0·9 per km). The MCRHR is located in the north-eastern area of Saskatchewan and encompasses the largest health region in the province, ~93 427 km<sup>2</sup>, and has an estimated population of 22 427 (0·2 per km) ([www.health.gov.sk.ca/covered-population2008](http://www.health.gov.sk.ca/covered-population2008)).

### Identification of MRSA and MSSA isolates

Isolates were identified as *S. aureus* using standard microbiological methodologies at the Nipawin Hospital Laboratory (NHL), Sandy Bay Health Center (SBHC), and the Saskatchewan Disease Control Laboratory (SDCL). MRSA was defined as isolates of *S. aureus* showing growth on oxacillin agar screen plates (Mueller–Hinton agar supplemented with 4% NaCl and oxacillin, 6 µg/ml) incubated at 35 °C for 24 h. All isolates identified as phenotypically positive for MRSA from the NHL or SBHC were sent to SDCL for confirmation by detecting for the presence of *mecA* and *nuc* genes using PCR [25].

### Case definitions

In this study, cases (CA-MRSA) and controls (CA-MSSA) were defined as a laboratory-confirmed community-acquired infection according to the Center for Disease Control and Prevention (CDC) definitions (non-hospitalization >48 h before the positive MRSA culture; no dialysis, surgery, residence in a long-term care facility, or hospitalization in the past year, no prior MRSA infection in the past year, and no percutaneous device or indwelling catheter) [26]. Individuals identified as potential study participants were contacted by community healthcare workers directly to determine their interest in participating in the study. Patients consenting to participate were interviewed using a standard investigation form (available at [www.narp.ca](http://www.narp.ca)), which captured demographic and exposure to possible risk factors. The study was approved by the research ethics boards of the University of Manitoba, the University of Saskatchewan, and Health Canada.

### Isolate characterization

All CA-MRSA and CA-MSSA isolates were submitted to the National Microbiology Laboratory (NML)

for additional molecular characterization, which included pulsed-field gel electrophoresis (PFGE) and *spa*-typing performed as previously described [27, 28]. PCR for detection of the *mupA* gene was as previously described [29]. Antimicrobial susceptibility of isolates was determined using standard broth microdilution panels according to CLSI guidelines [30]. Breakpoints used for fusidic acid and mupirocin resistance, which were not provided in the CLSI guideline, were as previously described [31, 32]. Inducible resistance to clindamycin in macrolide-resistant strains of CA-MRSA and CA-MSSA was detected by a standardized disk approximation test [30].

### Statistical analysis

Data was entered into Epi-Info version 3.4.3 (CDC, USA). Univariate analysis was performed to compare the demographic variables, site of infection, and medical history between the CA-MRSA and CA-MSSA cases. Fisher's exact test or  $\chi^2$  tests where appropriate were used to compare categorical variables. Unpaired Student's *t* test was used for continuous data. Multivariate logistic regression analysis was used to determine characteristics associated with acquisition of CA-MRSA or CA-MSSA. Stepwise logistic regression models for  $P < 0.25$  were used in the multivariate analysis. All tests were two-tailed, and a *P* value of  $< 0.05$  was considered statistically significant. Multivariate analysis was performed using SPSS version 11.0 (SPSS Inc., USA).

## RESULTS

A case-control study was undertaken comparing patients who had acquired either a CA-MRSA (case) or CA-MSSA (control) infection in order to identify potential risk factors for acquiring CA-MRSA infections. A total of 115 patients, representing 79 CA-MRSA and 36 CA-MSSA infections, fulfilled the screening criteria and provided consent to participate in this study. No significant differences were observed when comparing gender or age of patients who had acquired either a CA-MRSA or CA-MSSA infection (Table 1). The majority of CA-MRSA (79.7%) and CA-MSSA (77.8%) infections were in individuals aged  $< 30$  years (Table 1). Skin and soft tissue was the predominant site of infection for both CA-MRSA (67.1%) and CA-MSSA (55.6%), but no significant differences were observed when comparing specific anatomical sites of infection (data not shown). Of

Table 1. *Gender and age distribution of CA-MRSA and CA-MSSA infections*

	CA-MRSA		CA-MSSA	
	Total	%	Total	%
Gender				
Male	43	54.4	14	38.9
Female	36	45.6	22	61.1
Age (yr)				
0–10	38	48.1	21	58.3
> 10–20	14	17.7	5	13.9
> 20–30	11	13.9	2	5.6
> 30–40	6	7.6	3	8.3
> 40–50	7	8.9	1	2.8
> 50–60	2	2.5	3	8.3
> 60–70	0	0	1	2.8
> 70–80	0	0	0	0
> 80	1	1.3	0	0

note, only CA-MRSA isolates (7.7%,  $P = 0.10$ ) were obtained from the buttock region.

The investigation surveys used to identify potential risk factors for the acquisition of CA-MRSA did not reveal any statistical differences between a CA-MRSA and CA-MSSA infection (Table 2). For individuals with CA-MRSA infections, slightly higher proportions who were smokers (OR 2.12, 95% CI 0.75–6.14,  $P = 0.12$ ) and lower proportions with access to indoor plumbing (OR 0.3, 95% CI 0.06–1.23,  $P = 0.06$ ) were noted. Shared common risk factors included existing skin conditions, exposure to someone with a skin condition or MRSA infection, scratches and insect bites, exposure to healthcare workers, previous antibiotic usage, and overcrowding (Table 2). In regard to antimicrobial usage, the average number of antimicrobials prescribed in the previous year was 2.5 (median 2, range 0–10) and 1.7 (median 2, range 0–4) for patients with either a CA-MRSA or CA-MSSA infection, respectively. Concerning overcrowding, the average number of people per household was 6.3 (median 7, range 1–14) and 5.4 (median 5.5, range 1–9) for CA-MRSA and CA-MSSA infections, respectively.

A high proportion of the patients, 88.7% for CA-MRSA and 79.3% for CA-MSSA, were prescribed antimicrobials for their infection. The descending order of the top three classes of antimicrobials empirically prescribed alone or in combination with another drug for the treatment of CA-MSSA infections was a folate pathway inhibitor [trimethoprim-sulfamethoxazole (TMP-SXT)] (50%) >  $\beta$ -lactam

Table 2. Investigative forms to identify potential risk factors for CA-MRSA infections (percentage in parentheses calculated based on total Yes/No respondents)

Risk factors	CA-MRSA			CA-MSSA		
	Yes	No	Unknown/ Declined	Yes	No	Unknown/ Declined
Alcohol abuse	7 (9.0)	71	1	3 (8.8)	31	2
Asthma	9 (12.0)	66	4	3 (8.6)	32	1
Diabetes mellitus	6 (7.9)	70	3	3 (8.6)	32	1
Emphysema/COPD	2 (2.6)	74	3	0 (0)	33	3
Heart failure/CHF	2 (2.6)	75	2	1 (2.9)	34	1
HIV/AIDS	0 (0)	78	1	0 (0)	35	1
Immunosuppressive therapy	1 (1.4)	72	6	0 (0)	34	2
Liver disease	1 (1.3)	77	1	0 (0)	35	1
Malignancy	0 (0)	76	3	0 (0)	32	4
Chronic renal insufficiency	2 (2.6)	74	3	0 (0)	34	2
Eczema or psoriasis	9 (12.5)	63	7	6 (17.6)	28	2
Other chronic skin condition	8 (10.1)	71	0	4 (11.1)	32	0
Insect bite (site of infection)	13 (17.3)	62	4	4 (11.8)	30	2
Scratch/cut (site of infection)	17 (23.0)	57	5	8 (24.2)	25	3
Intravenous drug use	1 (1.3)	75	3	0 (0)	35	1
Current smoker	27 (34.6)	51	1	7 (20)	28	1
Antibiotic Rx in past year	70 (88.6)	9	0	35 (97.2)	1	0
Additional people in home	75 (94.9)	4	0	35 (97.2)	1	0
Exposure to healthcare workers in the past year	19 (24.7)	58	2	7 (20.6)	27	2
Exposure to nursing-home residents or persons in hospital in the past year	4 (5.2)	73	2	3 (8.8)	31	2
Household contact to person with MRSA	11 (21.2)	41	27	5 (22.7)	17	14
Visit to local emergency department in the past year	22 (28.9)	54	3	8 (22.9)	27	1
Stay in a correctional facility/prison in the past year	3 (3.9)	73	3	2 (5.9)	32	2
Day-care attendance	7 (9.0)	71	1	3 (8.6)	32	1
Exposure to person with skin condition/infection	22 (33.3)	44	13	12 (40)	18	6
Travel out of province	11 (17.5)	52	16	4 (11.8)	30	2
Indoor plumbing	49 (73.1)	18	12	27 (90)	3	6

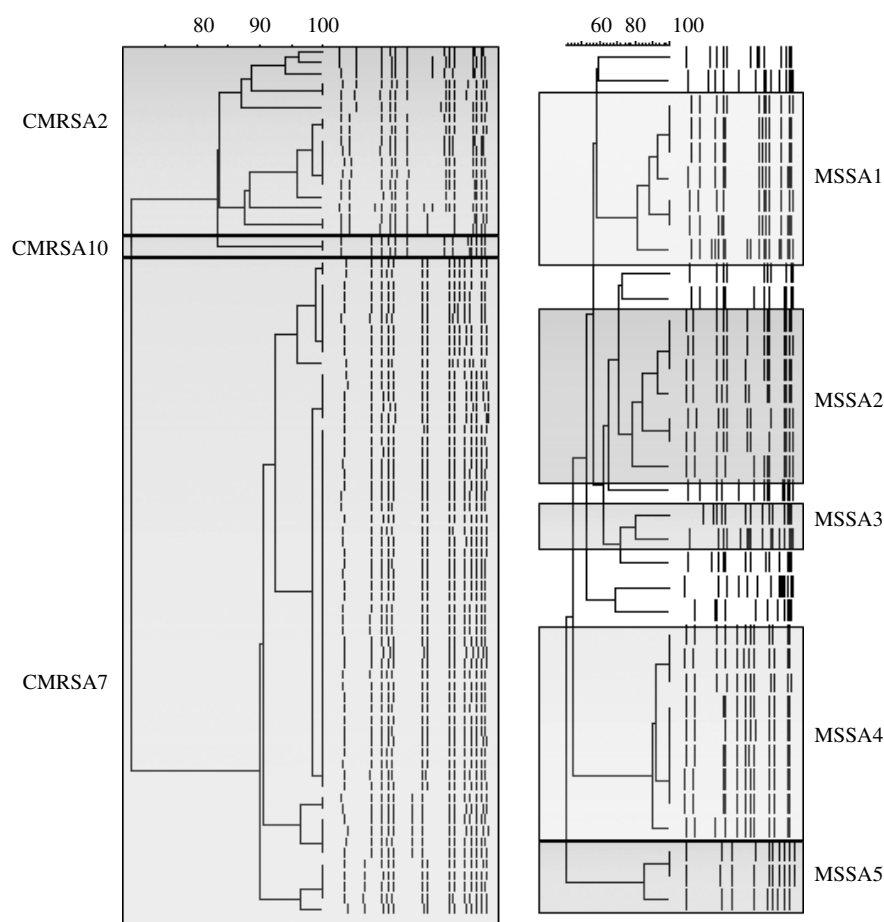
COPD, Chronic obstructive pulmonary disease; CHF, congestive heart failure.

(including  $\beta$ -lactamase-sensitive and -resistant penicillins and/or cephalosporins) (33.3%) > macrolide (25%). The descending order of the top three empirically prescribed antimicrobial classes alone or in combination with another drug for the treatment of CA-MRSA infections in the region were  $\beta$ -lactam (including  $\beta$ -lactamase-sensitive and -resistant penicillins and/or cephalosporins) (57.1%) > TMP-SXT (28.6%) > mupirocin (14.3%)  $\geq$  macrolide (14.3%).

CA-MRSA and CA-MSSA isolates were characterized by PFGE and *spa*-typing. For CA-MRSA, Canadian MRSA PFGE epidemic type CMRSA7 (USA400/MW2) was the predominant clone in these regions, accounting for 74.7% of all tested CA-MRSA isolates. Of the 59 CMRSA7 isolates, three *spa* types were identified (t128,  $n=48$ ; t127,  $n=7$ ; and t1788,  $n=4$ ). The remaining two CA-MRSA PFGE

clusters, CMRSA2 (USA100/800) and CMRSA10 (USA300), represented 22.8% and 2.5% of the remaining isolates, respectively (Fig. 1). The *spa* types for the CMRSA2 and CMRSA10 isolates were t311 and t008, respectively. Of the CA-MRSA isolates, 46.8% were found to harbour the genes encoding for PVL, which were only identified amongst the CMRSA7 (35/59) and CMRSA10 (2/2) isolates. The 24 CMRSA7 PVL-negative isolates were identified amongst various PFGE fingerprint patterns, but were indistinguishable from the PVL-positive isolates of the same PFGE fingerprint pattern.

In comparison to CA-MRSA, CA-MSSA were genetically more diverse with a number of different PFGE strain types identified. Eight CA-MSSA isolates displayed unique PFGE fingerprints, whereas the fingerprints for the remaining 28 isolates were



**Fig. 1.** Pulsed-field gel electrophoresis (PFGE) dendrogram of the CA-MRSA ( $n=79$ ) and CA-MSSA ( $n=36$ ) isolates. MRSA and MSSA PFGE clusters are indicated by the shaded blocks defined by  $>80\%$  similarity index and *spa* type. Canadian epidemic MRSA PFGE types are indicated in text. PFGE clusters of MSSA isolates were arbitrarily labelled 1–5 for this study.

distributed in five PFGE clusters (MSSA1–5) (Fig. 1). MSSA4 was the predominant CA-MSSA strain in the region, accounting for 25% of the typed CA-MSSA isolates, and were all found to be t159 by *spa*-typing. A second cluster, MSSA1, accounted for 19.4% of the CA-MSSA isolates and the predominant *spa* type amongst these isolates was t084. Two of the remaining CA-MSSA PFGE fingerprint clusters, MSSA2 (*spa* types t311, t002) and MSSA5 (*spa* types t012, t018) were related to Canadian PFGE MRSA epidemic types CMRSA2 and CMRSA4, respectively, by both PFGE and *spa*-typing [33]. All CA-MSSA isolates were negative for the genes encoding PVL using PCR.

Of the 11 antimicrobials tested, no resistance to vancomycin, linezolid, TMP–SXT, and tetracycline was observed for the CA-MRSA and CA-MSSA isolates (Table 3). CA-MRSA were statistically more likely to be resistant to ciprofloxacin (OR 4.32, 95%

CI 0.87–29.0,  $P=0.04$ ) and mupirocin (OR 13.92, 95% CI 4.5–45.0,  $P<0.001$ ). All mupirocin-resistant CA-MRSA isolates displayed high level resistance ( $\geq 256 \mu\text{g/ml}$ ) in comparison to only 36% (5/14) of the CA-MSSA isolates. Select high level mupirocin-resistant isolates were all found to harbour the *mupA* gene by PCR. The minimum inhibitory concentration for low-level mupirocin-resistant CA-MSSA isolates ranged from 4–16  $\mu\text{g/ml}$  and all were negative by PCR for the *mupA* gene. In addition to mupirocin, resistance to another topical antimicrobial, fusidic acid, was relatively high in both CA-MRSA (21.5%) and CA-MSSA (19.4%) isolates from these regions. High levels of resistance to erythromycin and clindamycin was also noted for both CA-MRSA (30.4%/22.8%) and CA-MSSA (41.7%/27.8%) isolates. Inducible resistance to clindamycin accounted for 12/18 (66.7%) and 7/11 (63.6%) of the clindamycin-resistant CA-MRSA and CA-MSSA isolates, respectively.



Table 3. Antimicrobial susceptibilities for CA-MRSA and CA-MSSA infections

Antibiotic	CA-MRSA				CA-MSSA			
	Resistant (%)	Range	MIC <sub>50</sub>	MIC <sub>90</sub>	Resistant (%)	Range	MIC <sub>50</sub>	MIC <sub>90</sub>
Erythromycin	30.4	0.25–>8	1	>8	41.7	0.25 to >8	1	>8
Clindamycin	22.8	≤0.25 to >8	≤0.25	≤0.25	27.8	≤0.25 to >8	≤0.25	≤0.25
Ciprofloxacin	20.3	0.12 to >8	0.25	>8	5.6	0.12 to >8	0.25	0.5
Vancomycin	0	0.5 to 1	1	1	0	0.5 to 1	1	1
Rifampin	1.3	≤0.25 to 4	≤0.25	≤0.25	0	≤0.25	≤0.25	≤0.25
Linezolid	0	1 to 4	2	4	0	1 to 4	2	4
TMP–SXT	0	≤0.25	≤0.25	≤0.25	0	≤0.25	≤0.25	≤0.25
Tetracycline	0	≤2	≤2	≤2	0	≤2	≤2	≤2
Fusidic acid	21.5	≤0.06 to >8	0.25	>8	19.4	0.12 to >8	0.25	>8
Mupirocin	78.5	0.12 to >128	>128	>128	38.9	≤0.12 to >128	0.25	>128
Gentamicin	7.6	≤0.5 to >16	≤0.5	1	13.9	≤0.5 to >16	≤0.5	>16

TMP–SXT, Trimethoprim–sulfamethoxazole; MIC, minimum inhibitory concentration.

## DISCUSSION

Anecdotal evidence from northern Saskatchewan and other Canadian communities suggests that physicians and prescribing nurses frequently opt to empirically prescribe antimicrobials without obtaining laboratory identification of the infecting organism or drug resistance. It has therefore been speculated that liberal antimicrobial prescribing practices in northern communities may contribute to community acquisition of CA-MRSA. In this study, exposure to antibiotics in the year prior to acquiring the infection was very high in people with either a CA-MRSA (88.6%) or CA-MSSA (97.2%) infection (Table 2). Data obtained for the treatment of CA-MRSA or CA-MSSA infections revealed that a high proportion of individuals were prescribed antimicrobials. Due to the high usage of  $\beta$ -lactams, appropriate antibiotics were more likely to be given to patients with a MSSA infection, which could be contributing to treatment failures, recurrent infections, and/or further dissemination of CA-MRSA within these communities.

It should be noted that this study had some limitations, including the small number of CA-MSSA infections ( $n=36$ ) that reduced the study's statistical power and also prevented the use of matching criteria. In addition, the average follow-up time was 6.1 months from time of positive culture to completed questionnaires, which could present some recall bias into the study. Unique risk factors for the acquisition of a CA-MRSA infection *vs.* a CA-MSSA infection were not identified, which coincides with a recently reported community-onset MRSA infection Danish

study [34]. In the Danish study, the only two significant risk factors identified included non-Danish origin and prior hospitalization for more than 7 days within 6 months before infection, the latter of which would have been screened out in our study. Common risk factors for CA-MRSA and CA-MSSA identified here, including younger age groups, exposure to healthcare workers, previous antibiotic usage, chronic skin conditions, and household contact to person with MRSA and exposure to person with a skin condition (Table 3), were similar to a recent investigation of CA-MRSA in a remote northern community in Nunavut, Canada [24]. As with this study, the predominant CA-MRSA clone identified in the Nunavut community was CMRSA7 (USA400) [24].

Community-associated outbreaks in Canada's larger urban centres have predominantly involved CMRSA10 (USA300). The most common risk factors reported for CMRSA10 infections have included history of illicit drug use, homelessness, and incarceration [5, 7], which was not pertinent in this study nor in the Nunavut study [24] as risk factors for CMRSA7 infection. These different risk factors might be contributing to the separate dissemination of CMRSA7 throughout the northern prairies and Nunavut, whereas CMRSA10 is predominating in western Canada's larger urban centres. A second possibility, which remains to be examined, is that differences in host genetics, immunity, and/or microbiomes could result in enhanced susceptibility of a population to a particular CA-MRSA strain type.

Similar risk factors, identified for both CA-MRSA and CA-MSSA infections in our study, and recently

in Denmark [34], suggest that standard hygienic measures and proper treatment guidelines are likely to be beneficial in controlling both CA-MRSA and CA-MSSA. Similar strategies have previously been successful in limiting CA-MRSA transmission in prison inmates [9] and sports teams [35]. To address this issue in northern Saskatchewan, treatment algorithms and many educational materials, such as hand-washing posters, patient pamphlets, community presentations, radio advertisements, podcasts, and newly developed educational tools for school-aged children (Germs Away and a computer flash animation game) have been provided throughout many northern communities and schools in Saskatchewan. These materials are all freely available ([www.narp.ca](http://www.narp.ca)) and will hopefully aid in increasing awareness of the importance of proper antimicrobial usage and hygiene in diminishing the spread of disease.

## ACKNOWLEDGEMENTS

Funding was provided by the Canadian Institutes of Health Research and the Public Health Agency of Canada.

## DECLARATION OF INTEREST

None.

## REFERENCES

1. Low DE, *et al.* Methicillin-resistant *Staphylococcus aureus*-Ontario. *Canadian Disease Weekly Report* 1981; **7**: 249–250.
2. Simor AE, *et al.* The evolution of methicillin-resistant *Staphylococcus aureus* in Canadian hospitals: 5 years of national surveillance. *Canadian Medical Association Journal* 2001; **165**: 21–26.
3. Simor AE, *et al.* Laboratory characterization of methicillin-resistant *Staphylococcus aureus* in Canadian hospitals: Results of 5 years of national surveillance, 1995–1999. *Journal of Infectious Diseases* 2002; **186**: 652–660.
4. Aiello AE, *et al.* Methicillin-resistant *Staphylococcus aureus* among US prisoners and military personnel: review and recommendations for future studies. *Lancet Infectious Diseases* 2006; **6**: 335–341.
5. Al-Rawahi GN, *et al.* Methicillin-resistant *Staphylococcus aureus* nasal carriage among injection drug users: six years later. *Journal of Clinical Microbiology* 2008; **46**: 477–479.
6. Benjamin HJ, Nikore V, Takagishi J. Practical management: Community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA): the latest sports epidemic. *Clinical Journal of Sports Medicine* 2007; **17**: 393–397.
7. Gilbert M, *et al.* Outbreak in Alberta of community-acquired (USA300) methicillin-resistant *Staphylococcus aureus* in people with a history of drug use, homelessness or incarceration. *Canadian Medical Association Journal* 2006; **175**: 149–154.
8. Lo WT, *et al.* Nasal carriage of a single clone of community-acquired methicillin-resistant *Staphylococcus aureus* among kindergarten attendees in northern Taiwan. *BMC Infectious Diseases* 2007; **7**: 51.
9. Main CL, *et al.* Outbreaks of infection caused by community-acquired methicillin-resistant *Staphylococcus aureus* in a Canadian correctional facility. *Canadian Journal of Infectious Diseases and Medical Microbiology* 2005; **16**: 343–348.
10. Mulvey MR, *et al.* Community-associated methicillin-resistant *Staphylococcus aureus*, Canada. *Emerging Infectious Diseases* 2005; **11**: 844–850.
11. Ofner-Agostini M, *et al.* Methicillin-resistant *Staphylococcus aureus* in Canadian aboriginal people. *Infection Control and Hospital Epidemiology* 2006; **27**: 204–207.
12. Adam H, McGeer A, Simor A. Fatal case of post-influenza, community-associated MRSA pneumonia in an Ontario teenager with subsequent familial transmission. *Canadian Communicable Disease Report* 2007; **33**: 45–48.
13. Castaldo ET, Yang EY. Severe sepsis attributable to community-associated methicillin-resistant *Staphylococcus aureus*: an emerging fatal problem. *The American Surgeon* 2007; **73**: 684–688.
14. Vayalumkal JV, *et al.* Necrotizing pneumonia and septic shock: Suspecting CA-MRSA in patients presenting to Canadian emergency departments. *Journal of the Canadian Association of Emergency Physicians* 2007; **9**: 300–303.
15. Seybold U, *et al.* Emergence of community-associated methicillin-resistant *Staphylococcus aureus* USA300 genotype as a major cause of health care-associated blood stream infections. *Clinical Infectious Diseases* 2006; **42**: 647–656.
16. D'Agata EM, *et al.* Modeling the invasion of community-acquired methicillin-resistant *Staphylococcus aureus* into hospitals. *Clinical Infectious Diseases* 2009; **48**: 274–284.
17. Embil J, *et al.* Methicillin-resistant *Staphylococcus aureus* in tertiary care institutions on the Canadian prairies 1990–1992. *Infection Control and Hospital Epidemiology* 1994; **15**: 646–651.
18. Shahin R, *et al.* Methicillin-resistant *Staphylococcus aureus* carriage in a child care center following a case of disease. Toronto child care center study group. *Archives of Pediatric and Adolescent Medicine* 1999; **153**: 864–868.
19. Christianson S, *et al.* Comparative genomics of Canadian epidemic lineages of methicillin-resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology* 2007; **45**: 1904–1911.

20. **Al-Rawahi GN, et al.** Community-associated CMRSA-10 (USA-300) is the predominant strain among methicillin-resistant *Staphylococcus aureus* strains causing skin and soft tissue infections in patients presenting to the emergency department of a Canadian tertiary care hospital. *Journal of Emergency Medicine*. Published online: 4 March 2008. doi:10.1016/j.jemermed.2007.09.030.
21. **Kurbis AC, Wylie JL.** Community-based cluster of methicillin-resistant *Staphylococcus aureus* in Manitoba. *Canadian Journal of Infectious Diseases* 2001; **12**: 149–152.
22. **Wylie JL, Nowicki DL.** Molecular epidemiology of community- and health care-associated methicillin-resistant *Staphylococcus aureus* in Manitoba, Canada. *Journal of Clinical Microbiology* 2005; **43**: 2830–2836.
23. **Irvine J, et al.** Dissemination of community-associated methicillin-resistant *Staphylococcus aureus* CMRSA7 (USA400) in northern Saskatchewan, Canada. Sixth International Conference on Emerging Infectious Diseases, 16–19 March 2008, Atlanta, Georgia.
24. **Daloo A, et al.** Investigation of community-associated methicillin-resistant *Staphylococcus aureus* in a remote northern community, Nunavut, Canada. *Canadian Communicable Disease Report* 2008; **34**: 1–7.
25. **McDonald RR, et al.** Development of a triplex real-time PCR assay for detection of Panton-Valentine leukocidin toxin genes in clinical isolates of methicillin-resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology* 2005; **43**: 6147–6149.
26. **Naimi TS, et al.** Comparison of community- and health care-associated methicillin-resistant *Staphylococcus aureus* infection. *Journal of the American Medical Association* 2003; **290**: 2976–2984.
27. **Mulvey MR, et al.** Development of a Canadian standardized protocol for subtyping methicillin-resistant *Staphylococcus aureus* using pulsed-field gel electrophoresis. *Journal of Clinical Microbiology* 2001; **39**: 3481–3485.
28. **Harmsen D, et al.** Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for *spa* repeat determination and database management. *Journal of Clinical Microbiology* 2003; **41**: 5442–5448.
29. **Anthony RM, et al.** Use of the polymerase chain reaction for rapid detection of high-level mupirocin resistance in staphylococci. *European Journal of Clinical Microbiology and Infectious Diseases* 1999; **18**: 30–34.
30. **Clinical and Laboratory Standards Institute.** Performance standards for antimicrobial susceptibility testing; 17th informational supplement. CLSI document M100-S17. Clinical and Laboratory Standards Institute, 2007. Wayne, PA.
31. **Skov R, Frimodt-Moller N, Espersen F.** Correlation of MIC methods and tentative interpretive criteria for disk diffusion susceptibility testing using NCCLS methodology for fusidic acid. *Diagnostic Microbiology and Infectious Diseases* 2001; **40**: 111–116.
32. **Walker ES, et al.** A decline in mupirocin resistance in methicillin-resistant *Staphylococcus aureus* accompanied administrative control of prescriptions. *Journal of Clinical Microbiology* 2004; **42**: 2792–2795.
33. **Golding GR, et al.** A preliminary guideline for the assignment of methicillin-resistant *Staphylococcus aureus* to a Canadian pulsed-field gel electrophoresis epidemic type using *spa* typing. *Canadian Journal of Infectious Disease and Medical Microbiology* 2008; **19**: 273–281.
34. **Bocher S, et al.** Methicillin-resistant *Staphylococcus aureus*: risk factors associated with community-onset infections in Denmark. *Clinical Microbiology and Infection* 2008; **14**: 942–948.
35. **Romano R, Lu D, Holtom P.** Outbreak of community-acquired methicillin-resistant *Staphylococcus aureus* skin infections among a collegiate football team. *Journal of Athletic Training* 2006; **41**: 141–145.