Article



Association Between Rheumatoid Arthritis and Clonal Hematopoiesis: A Mendelian Randomization Study

Jie Zhang¹, Chun Zhou² and Shaoxing Guan³ ()

¹Department of Pharmacy, Guangdong Provincial Hospital of Chinese Medicine, The Second Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangzhou, China, ²School of Pharmaceutical Sciences; Guangdong Provincial Key Laboratory of Shock and Microcirculation, Southern Medical University, Guangzhou, China and ³NMPA Key Laboratory for Research and Evaluation of Drug Metabolism & Guangdong Provincial Key Laboratory of New Drug Screening & Guangdong-Hongkong-Macao Joint Laboratory for New Drug Screening, School of Pharmaceutical Sciences, Southern Medical University, Guangzhou, China

Abstract

Immunity activation and inflammation are the main characteristics of rheumatoid arthritis and clonal hematopoiesis. However, it remains unclear whether rheumatoid arthritis increase the risk of clonal hematopoiesis. Here, a Mendelian randomization (MR) analysis was conduct to explore the causal effects of rheumatoid arthritis on clonal hematopoiesis. Summary statistics data of rheumatoid arthritis (13,838 cases and 33,742 controls) and clonal hematopoiesis (10,203 cases and 173,918 controls) derived from a genomewide association study were selected to analyze. We selected inverse-variance weighted, MR-Egger, weighted median, simple mode, and weighted mode to evaluate the causal effect of rheumatoid arthritis on clonal hematopoiesis. The two-sample MR analysis suggested a strong causal relationship between rheumatoid arthritis and clonal hematopoiesis by inverse-variance weighted (OR = 1.002311673, 95% CI [1.000110757, 1.004517433], p = .039518447) methods. No significant pleiotropy or heterogeneity was found in the sensitivity analysis. These results supported a potentially causal relationship between rheumatoid arthritis and clonal hematopoiesis, and the exposure of rheumatoid arthritis increased the risks of clonal hematopoiesis. Our findings highlight the importance of how chronic inflammation and immune activation induced rheumatoid arthritis enhances the risks of clonal hematopoiesis, and that early intervention with rheumatoid arthritis patients might reduce the clonal hematopoiesis risks in rheumatoid arthritis patients. Moreover, our study provides clues for prediction of risk factors and potential mechanisms of clonal hematopoiesis.

Keywords: Rheumatoid arthritis; Clonal hematopoiesis; Mendelian randomization

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Clonal hematopoiesis (CH) is defined as any clonal outgrowth of hematopoietic progenitor cells (Genovese et al., 2014). The prevalence of CH depends on the sensitivity of detection methods, and increases with age (Evans & Walsh, 2023). Clonal hematopoiesis is associated with an increase in the risk of all-cause mortality, hematological malignancy and nonhematological malignancy, chronic kidney disease, and cardiovascular diseases due to accumulation of somatic mutations. Although risk factors of CH include male sex (Jaiswal et al., 2014), germline mutation (e.g., 8-bp intronic deletion in TERT; Zink et al., 2017) and cigarette smoking (Levin et al., 2022), potential causal risk factors for CH are still unclear.

Rheumatoid arthritis (RA) is one of the most common chronic inflammatory and autoimmune diseases with synovial inflammation and hyperplasia of joint tissue (McInnes & Schett, 2011; Scott et al., 2010; Smolen et al., 2016). Approximately 0.5–1.0% of the population in Europe (van der Woude & van der Helm-van Mil, 2018) and 0.32–0.36% in Asia (Tong et al., 2020) are affected by RA, which leads to joint pain, swelling and stiffness (van der

Woude & van der Helm-van Mil, 2018). Since RA has multi-organ and mulri-system involvement, the brain, heart, lung, kidney and hematologic system can be invaded by the disease exposure (Scott et al., 2010; Smolen et al., 2016). Although previous observational studies reported that the RA patients have increased risk for vertebral osteoporotic fractures (Tong et al., 2020), low bone mineral density (Lee et al., 2012), and bone fracture (Xue et al., 2017) due to abnormal inflammatory and autoimmune exposure, whether RA has an impact on CH is still unknown.

Several researchers have found that RA affects the proliferation of hematopoietic progenitor cells in vitro, including myeloid and lymphoid cells (Colmegna et al., 2012), which is the one of main characteristics of CH. Meanwhile, CH with a typical mutation profile occurs in RA (e.g., DNMT3A and TET2 mutations; Savola et al., 2018), implying that long systemic inflammation could increase the rate of CH by accumulation of somatic mutations. However, only one previous observational study found that the prevalence of CH was not markedly increased in RA patients (Savola et al., 2018) in a very small cohort (N = 59). Thus far, it is unknown whether RA is causally associated with CH.

In the present study, we used a two-sample Mendelian randomization (MR) analysis to evaluate the causal association of RA on CH, based on a genomewide association study (GWAS), which selected single-nucleotide polymorphisms (SNPs) as

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Corresponding author: Shaoxing Guan; Email: Guanshx6@mail.sysu.edu.cn

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instrumental variables (IVs) to evaluate the potential causal effects of exposures on outcomes. Our study illustrated the link between RA and CH, highlighting potential mechanisms in genetics of the RA-CH relationship.

Materials and Methods

Study Design

In this study, we utilized a two-sample MR to evaluate the causal association between RA and CH using data based on publicly available GWAS. The genetic variants (SNPs) were used as IVs to estimate potential causal effects of RA on CH. To conduct a valid MR analyses, three important hypotheses must be confirmed during the whole process (Lawlor et al., 2008). In brief, (a) the IVs are strongly associated with RA; (b) RA and CH are independent of any known confounders; (c) the IVs affected CH only through RA. There was no participant overlap between the exposure and outcome datasets.

Data Source and IVs Selection

The summary-level statistics for RA were currently derived from the published studies (Eyre et al., 2012), which involved 13,838 cases and 33,742 controls of European participants. Summary statistics for CH were obtained from a previously published GWAS study (Kar et al., 2022) that contained 10,203 cases and 173,918 controls from Europe (https://zenodo.org/records/5893861). The SNPs were strongly associated with RA (p < 5e-8) with low linkage disequilibrium (LD, $R^2 < .001$, clumping distance = 10000 kb), and large *F* values (*F* statistics > 10). We then manually searched the PhenoScanner database (http://www.phenoscanner.medschl.cam. ac.uk/phenoscanner) to exclude variants that were associated with confounders or outcomes. Finally, the SNPs with relative common frequency in population (minor allele frequency > 0.01) were selected as IVs.

MR Analysis and Sensitivity Analysis

The MR analysis was conducted in R 4.2.0 using the TwoSampleMR v0.5.7 (https://mrcieu.github.io/TwoSampleMR/ news/index.html). Orienting the causal relationship was determined by Steiger test. Five different methods were used to assess the causal effects of CH outcomes, including weighted mode, simple mode, weighted median, Mendelian randomization-egger (MR-Egger) and inverse-variance weighted (IVW). The MR-Egger method allows for horizontal pleiotropy of instrumental SNPs, but the method is not capable of distinguishing between causal effects and pleiotropy (Burgess & Thompson, 2017), and is easily affected by 'weak instrument bias' (Pierce & Burgess, 2013). Simple mode and weighted mode methods were set as a complementary MR analysis strategy. The IVW method combines the estimates of each SNP and provides a consistent estimate of the causal effect, which is the most reliable method when there is no horizontal pleiotropy of the instrumental SNPs (Bae & Lee, 2018; Bowden et al., 2016). Additionally, MR pleiotropy residual sum and outlier (MR-PRESSO) was conducted to exclude any potential pleiotropy (https://github.com/rondolab/MR-PRESSO), and p < .05 was regarded as statistical significance. Meanwhile, the MR-Egger and IVW methods were conducted to evaluate the heterogeneity quantified using Cochran's Q-test. The horizontal pleiotropy was analyzed by the MR-Egger method. In addition, the leave-one-out analysis was used to verify whether outliers existed that affected the result.

Results

IVs for RA

A total of 2386 SNPs were found significantly associated with RA in European population ($p < 5 \times 10^8$). After clumped analysis, funnel plots analysis and calculation of *F* statistic, 15 SNPs were selected as IVs to evaluate the causal relationship between RA and CH; the detailed information of IVs is shown in Table 1. In addition, no significant association was found between these SNPs and CH or related confounders according to the Phenoscanner database (Table S1).

MR Analysis

By performing IVW analysis (Table 2, Figure 1), we found causality of RA with CH, and RA was associated with increased risk of CH as indicated by IVW (OR = 1.002311673, 95% CI [1.000110757, 1.004517433], p = .039706) and weighted median (OR = 1.002311 673, 95% CI [1.000110757, 1.004517433], p = .039518447) methods. As for the MR-Egger regression (MR-Egger, OR = 1.001021956, 95% CI [0.99816608, 1.003886003], p = .0495823), simple mode (OR = 1.002411974, 95% CI [0.997456772, 1.007391792], p = .356 839342) and weighted mode (OR = 1.002630923, 95% CI [1.000336062, 1.004931049], p = .041255472), since the beta value of all two major MR approaches (IVW and weighted median) were ≥ 0 and the OR value was ≥ 1 , this suggests a positive causal connection between RA and CH. In addition, we found that the assumption that RA causes CH was valid, as indicated by Steiger p < .0001, using a Steiger test (Table S2).

Sensitivity Analysis

MR pleiotropy residual sum and outlier (MR-PRESSO) analysis indicated no pleiotropy and outlier SNP in this study (p = .5700). In addition, the intercepts of RA on CH was 0.0002625363 (p = .4892), indicating that IV had no horizontal pleiotropic effect (Table S3 and Figure 1). Meanwhile, no obvious heterogeneity was observed according to Q values based on MR-Egger (p = .5323473) and IVW (p = .5702701) tests (Table S3). The leave-one-out (Figure 2) and funnel plots (Figure 3) demonstrated that there were no obvious or potential outlier IVs, strengthening the robustness of our results, which made our MR results more convincing.

Discussion

This MR study aimed to investigate the causal effect of RA on CH by reanalysis GWAS summary data in European ancestry. Consistent with our hypothesis, a positive causality between RA and CH risk was found in this study (IVW, OR = 1.002311673, 95% CI [1.000110757, 1.004517433], p = .039706); Weighted median, OR = 1.002311673, 95% CI = [1.000110757, 1.004517433], p = .039518447). In the sensitivity analysis, the MR results were robust and reliable. These findings suggested that RA exposure increases the risk of CH.

Recently, MR has become a useful tool to assess the causal relationship between exposure and outcome via genetic markers. Since RA has multi-organ and multi-system involvement, several studies have revealed the causal association between RA and pre-eclampsia (Zhang et al., 2022), hepatocellular carcinoma (Zhang et al., 2023), glaucoma (Meng et al., 2023) and cardio-cerebrovascular disease (Qiu et al., 2021), implying that RA has a broad effect on various pathophysiology progresses. In this study, we found a causal association between RA and CH via a robust MR

rs932036

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SNP Chr Position Effect allele Other allele Beta SE EAF p value rs10209110 2 1E + 080.1058 0.0185 0.52 1.13E-08 G А rs13426947 2 1.9E + 08G 0.1426 0.0231 0.1909 7.19E-10 A rs2228145 1 1.5E + 08С А -0.1090.0191 0.3598 1.32E-08 0.0198 1.36E-08 rs2240339 1 1.8E + 07A G -0.1120.3469 rs2269429 6 3.2E + 07 A G -0.272 0.035 0.1074 9.27E-15 rs2476601 A G 0.4659 0.0281 0.0944 9.01E-62 1 1.1E + 08rs2812378 9 3.5E + 07G А 0.1104 0.0194 0.3161 1.31E-08 rs3130190 6 3.3E + 07G А -0.214 0.0217 0.2714 5.49E-23 rs3131875 6 3E + 07G А 0.105 0.0187 0.4602 1.82E-08 rs34536443 19 1E + 07G С -0.373 0.0511 0.9712 2.70E-13 6 С rs592229 3.2E + 07A -0.2550.019 0.3728 2.94E-41 rs6920220 6 1.4E + 08А G 0.1405 0.022 0.169 1.84E-10 rs71624119 5 5.5E + 07А G -0.207 0.0226 0.2445 5.59E-20 rs9268145 6 3.2E + 07С А 0.7064 0.0211 0.1869 1.00E-200

А

Table 1. Genetic variants used as instruments for rheumatoid arthritis.

Note: SNP, single nucleotide polymorphism; Chr, chromosome; EAF, effect allele frequency.

Т

2.6E + 07

Table 2. Mendelian randomization estimates of rheumatoid arthritis associated with the risk of clonal hematopoiesis

Methods	IVs (<i>n</i>)	Beta	SE	OR	OR (lower limit of 95% CI)	OR (upper limit of 95% CI)	<i>p</i> value
MR Egger	15	0.001021	0.00146	1.00102	0.998166	1.003886	0.4958
Weighted median	15	0.002309	0.00112	1.00231	1.000111	1.004517	0.0395
Inverse variance weighted	15	0.00184	0.00089	1.00184	1.000087	1.003601	0.0397
Simple mode	15	0.002409	0.00253	1.00241	0.997457	1.007392	0.3568
Weighted mode	15	0.002627	0.00117	1.00263	1.000336	1.004931	0.0413

0.1274

0.02

Note: MR, Mendelian randomization; IVs, instrumental variables. Bold type indicates significance.



Figure 1. The scatterplot depicts the causal relationship between rheumatoid arthritis and clonal hematopoiesis.

Note: MR, Mendelian randomization; SNP, single nucleotide polymorphism.

analysis, and that the exposure of RA increases the risk of CH. However, Savola and colleagues reported that RA had a minimum effect on the risk of CH in only 59 RA patients based on Illumina's TruSeq Custom Amplicon technology, which only covered 34

tumor suppressor genes and/or mutational hotspots (Savola et al., 2018). Meanwhile, the diagnosis of CH varied according to the approach and sensitivity of detestation (Evans & Walsh, 2023). For example, initial studies found the frequency of CH was 10% of individuals over the age of 70 in their white blood cells using whole exome sequencing (Jaiswal et al., 2014); recent studies with deeper sequencing technologies found that CH is ubiquitous by middle age or earlier (Watson et al., 2020; Young et al., 2016). Moreover, genetic drift and fitness shaping the genetic diversity were not detected in the observational study, which are the main characteristics of CH (Watson et al., 2020). More importantly, Savola and colleagues (2018) also found that CH with a typical mutation profile occurs in RA patients, implying that the reactions induced by RA might have an impact on the development of CH. Taken together, further studies with large cohorts and deeper sequencing technologies will be invaluable in the future.

Previous studies demonstrated that chronic inflammation and immune activation may cause mutagenesis via DNA damage by a series of immune-inflammation reactions (Nakad & Schumacher, 2016). Autoimmune response and chronic inflammation are the main features of RA (Smolen et al., 2016), which can lead to abundant mutation accumulation in patients with RA. The cellular composition in RA includes innate immune cells and adaptive immune cells involved with a matrix regulatory, aggressive

F

16.6929

18.7988

16.2698

17.1452

42.6297

15.7418

54.9939

16.3698

23.372

93.2454

16.5848

47.5956

20.5361

531.383

2.00E-10

0.3032

114.628



Figure 2. Leave-one-out analysis. Each dot in the forest plot represents the Mendelian randomization estimate (using inverse-variance weighted) excluding that particular instrumental variable.



Figure 3. Funnel plots of individual variant effects for the instrumental variables. Note: MR, Mendelian randomization.

inflammatory and invasive phenotype, which fails to detect DNA lesions, and thus leads to DNA damage and mutation accumulations (Shao, 2018; Smolen et al., 2007). Meanwhile, outgrowths of mutated blood cells, including two enzymes involved in DNA methylation (DNMT3A and TET2) and a chromatin regulator (ASXL1) as well as cellular growth signaling (GNAS, GNB1, JAK2, CBL) and the DNA damage response (PPM1D, TP53), are the main drivers mutation of CH (Jaiswal & Ebert, 2019). In addition, the mechanism and function of these mutations in CH remains unknown. Collectively, the exposure of RA for years may cause accumulation of mutation induced by chronic inflammation that leads to development of CH, and therefore more studies to consider this should be conducted in the future.

This study had some limitations. First, only 15 SNPs were selected as IVs in this study with a relatively limited effect on RA as indicated by R^2 (range [%]: 0.05286–15.1647). Therefore, other important factors besides RA that can affect the risk of CH should not be ignored in the next studies, such as smoking (Levin et al., 2022). Second, the MR results only involved European ancestry populations in this study, and whether a casual association exists between RA on CH needs to be clarified by further investigations. Third, previous studies reported that male and elderly individuals

have an increased risk of suffering CH, but the published summary GWAS data did not provide this information to adjust the bias. Fourth, the SNPs used for analysis may be correlated with other traits due to genetic polymorphisms and thus generate confounding bias, which may affect causal inference. Fifth, the strength of the IV depends on the sample size of the GWAS, and a larger scale GWAS is required to determine more genetic variation for MR. Finally, our study found the risk of CH increased under the exposure of RA for years, rather than for a specific time in life. Thus, additional explorations of the effect sizes between the specific time of RA and CH risks are needed. In addition, RA is a binary exposure (Ding et al., 2024), which might generate biased estimates and yield a spurious causal estimate (Burgess & Labrecque, 2018) between RA and CH. Therefore, the exclusion-restriction assumption was done in our study.

Conclusion

In conclusion, these results support a potentially causal relationship between RA and CH, and the exposure of RA increases the risks of CH. Our findings highlight the importance of chronic inflammation and immune activation induced RA on the risks of CH, and that early intervention for RA patients might reduce the CH risks in RA patients. Moreover, our study provides clues for prediction of risk factors and potential mechanisms of CH.

Supplementary material. To view supplementary material for this article, please visit https://doi.org/10.1017/thg.2024.24.

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Competing interests. None.

References

- Bae, S. C., & Lee, Y. H. (2018). Vitamin D level and risk of systemic lupus erythematosus and rheumatoid arthritis: A Mendelian randomization. *Clinical Rheumatology*, 37, 2415–2421. https://doi.org/10.1007/s10067-018-4152-9
- Bowden, J., Davey Smith, G., Haycock, P. C., & Burgess, S. (2016). Consistent estimation in mendelian randomization with some invalid instruments using a weighted median estimator. *Genetic Epidemiology*, 40, 304–314. https://doi. org/10.1002/gepi.21965
- Burgess, S., & Labrecque, J. A. (2018). Mendelian randomization with a binary exposure variable: interpretation and presentation of causal estimates. *European Journal of Epidemiology*, 33, 947–952. https://doi.org/10.1007/ s10654-018-0424-6
- Burgess, S., & Thompson, S. G. (2017). Interpreting findings from Mendelian randomization using the MR-Egger method. *European Journal of Epidemiology*, 32, 377–389. https://doi.org/10.1007/s10654-017-0255-x
- Colmegna, I., Pryshchep, S., Oishi, H., Goronzy, J. J., & Weyand, C. M. (2012). Dampened ERK signaling in hematopoietic progenitor cells in rheumatoid arthritis. *Clinical Immunology*, 143, 73–82. https://doi.org/10. 1016/j.clim.2012.01.007
- Ding, K., Jiang, W., Zhangwang, J., Li, J., & Lei, M. (2024). The effect of rheumatoid arthritis on features associated with sarcopenia: A Mendelian randomization study. *Calcified Tissue International*, 114, 286–294. https:// doi.org/10.1007/s00223-023-01178-w
- Evans, M. A., & Walsh, K. (2023). Clonal hematopoiesis, somatic mosaicism, and age-associated disease. *Physiological Reviews*, 103, 649–716. https://doi. org/10.1152/physrev.00004.2022
- Eyre, S., Bowes, J., Diogo, D., Lee, A., Barton, A., Martin, P., Zhernakova, A., Stahl, E., Viatte, S., McAllister, K., Amos, C. I., Padyukov, L., Toes, R. E., Huizinga, T. W., Wijmenga, C., Trynka, G., Franke, L., Westra, H. J.,

Alfredsson, L., ... Worthington, J. (2012). High-density genetic mapping identifies new susceptibility loci for rheumatoid arthritis. *Nature Genetics*, 44, 1336–1340. https://doi.org/10.1038/ng.2462

- Genovese, G., Kähler, A. K., Handsaker, R. E., Lindberg, J., Rose, S. A., Bakhoum, S. F., Chambert, K., Mick, E., Neale, B. M., Fromer, M., Purcell, S. M., Svantesson, O., Landén, M., Höglund, M., Lehmann, S., Gabriel, S. B., Moran, J. L., Lander, E. S., Sullivan, P. F., ... McCarroll, S. A. (2014). Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *New England Journal of Medicine*, 371, 2477–2487. https://doi.org/ 10.1056/NEJMoa1409405
- Jaiswal, S., & Ebert, B. L. (2019). Clonal hematopoiesis in human aging and disease. Science, 366, eaan4673. https://doi.org/10.1126/science.aan4673
- Jaiswal, S., Fontanillas, P., Flannick, J., Manning, A., Grauman, P. V., Mar, B. G., Lindsley, R. C., Mermel, C. H., Burtt, N., Chavez, A., Higgins, J. M., Moltchanov, V., Kuo, F. C., Kluk, M. J., Henderson, B., Kinnunen, L., Koistinen, H. A., Ladenvall, C., Getz, G., ... Ebert, B. L. (2014). Age-related clonal hematopoiesis associated with adverse outcomes. *New England Journal* of *Medicine*, 371, 2488–2498. https://doi.org/10.1056/NEJMoa1408617
- Kar, S. P., Quiros, P. M., Gu, M., Jiang, T., Mitchell, J., Langdon, R., Iyer, V., Barcena, C., Vijayabaskar, M. S., Fabre, M. A., Carter, P., Petrovski, S., Burgess, S., & Vassiliou, G. S. (2022). Genome-wide analyses of 200,453 individuals yield new insights into the causes and consequences of clonal hematopoiesis. *Nature Genetics*, 54, 1155–1166. https://doi.org/10.1038/ s41588-022-01121-z
- Lawlor, D. A., Harbord, R. M., Sterne, J. A., Timpson, N., & Davey Smith, G. (2008). Mendelian randomization: Using genes as instruments for making causal inferences in epidemiology. *Statistics in Medicine*, 27, 1133–1163. https://doi.org/10.1002/sim.3034
- Lee, S. G., Park, Y. E., Park, S. H., Kim, T. K., Choi, H. J., Lee, S. J., Kim, S. I., Lee, S. H., Kim, G. T., Lee, J. W., Lee, J. H., & Baek, S. H. (2012). Increased frequency of osteoporosis and BMD below the expected range for age among South Korean women with rheumatoid arthritis. *International Journal of Rheumatic Diseases*, 15, 289–296. https://doi.org/10.1111/j.1756-185X.2012. 01729.x
- Levin, M. G., Nakao, T., Zekavat, S. M., Koyama, S., Bick, A. G., Niroula, A., Ebert, B., Damrauer, S. M., & Natarajan, P. (2022). Genetics of smoking and risk of clonal hematopoiesis. *Scientific Reports*, 12, 7248. https://doi.org/ 10.1038/s41598-022-09604-z
- McInnes, I. B., & Schett, G. (2011). The pathogenesis of rheumatoid arthritis. New England Journal of Medicine, 365, 2205–2219. https://doi.org/10.1056/ NEJMra1004965
- Meng, Y., Tan, Z., Su, Y., Li, L., & Chen, C. (2023). Causal association between common rheumatic diseases and glaucoma: A Mendelian randomization study. *Frontiers in Immunology*, 14, 1227138. https://doi.org/10.3389/fimmu. 2023.1227138
- Nakad, R., & Schumacher, B. (2016). DNA damage response and immune defense: Links and mechanisms. *Frontiers in Genetics*, 7, 147. https://doi.org/ 10.3389/fgene.2016.00147
- Pierce, B. L., & Burgess, S. (2013). Efficient design for Mendelian randomization studies: Subsample and 2-sample instrumental variable estimators. *American Journal of Epidemiology*, 178, 1177–1184. https://doi. org/10.1093/aje/kwt084

- Qiu, S., Li, M., Jin, S., Lu, H., & Hu, Y. (2021). Rheumatoid arthritis and cardio-cerebrovascular disease: A Mendelian randomization study. *Frontiers* in Genetics, 12, 745224. https://doi.org/10.3389/fgene.2021.745224
- Savola, P., Lundgren, S., Keränen, M. A. I., Almusa, H., Ellonen, P., Leirisalo-Repo, M., Kelkka, T., & Mustjoki, S. (2018). Clonal hematopoiesis in patients with rheumatoid arthritis. *Blood Cancer Journal*, 8, 69. https://doi. org/10.1038/s41408-018-0107-2
- Scott, D. L., Wolfe, F., & Huizinga, T. W. (2010). Rheumatoid arthritis. *Lancet*, 376, 1094–1108. https://doi.org/10.1016/s0140-6736(10)60826-4
- Shao, L. (2018). DNA damage response signals transduce stress from rheumatoid arthritis risk factors into T cell dysfunction. *Frontiers in Immunology*, 9, 3055. https://doi.org/10.3389/fimmu.2018.03055
- Smolen, J. S., Aletaha, D., Koeller, M., Weisman, M. H., & Emery, P. (2007). New therapies for treatment of rheumatoid arthritis. *Lancet*, 370, 1861–1874. https://doi.org/10.1016/s0140-6736(07)60784-3
- Smolen, J. S., Aletaha, D., & McInnes, I. B. (2016). Rheumatoid arthritis. Lancet, 388, 2023–2038. https://doi.org/10.1016/s0140-6736(16)30173-8
- Tong, J. J., Xu, S. Q., Zong, H. X., Pan, M. J., Teng, Y. Z., & Xu, J. H. (2020). Prevalence and risk factors associated with vertebral osteoporotic fractures in patients with rheumatoid arthritis. *Clinical Rheumatology*, 39, 357–364. https://doi.org/10.1007/s10067-019-04787-9
- van der Woude, D., & van der Helm-van Mil, A. H. M. (2018). Update on the epidemiology, risk factors, and disease outcomes of rheumatoid arthritis. *Best Practice & Research: Clinical Rheumatology*, 32, 174–187. https://doi.org/10. 1016/j.berh.2018.10.005
- Watson, C. J., Papula, A. L., Poon, G. Y. P., Wong, W. H., Young, A. L., Druley, T. E., Fisher, D. S., & Blundell, J. R. (2020). The evolutionary dynamics and fitness landscape of clonal hematopoiesis. *Science*, 367, 1449–1454. https://doi.org/10.1126/science.aay9333
- Xue, A. L., Wu, S. Y., Jiang, L., Feng, A. M., Guo, H. F., & Zhao, P. (2017). Bone fracture risk in patients with rheumatoid arthritis: A meta-analysis. *Medicine*, 96, e6983. https://doi.org/10.1097/md.000000000006983
- Young, A. L., Challen, G. A., Birmann, B. M., & Druley, T. E. (2016). Clonal haematopoiesis harbouring AML-associated mutations is ubiquitous in healthy adults. *Nature Communications*, 7, 12484. https://doi.org/10.1038/ ncomms12484
- Zhang, D., Hu, Y., Guo, W., Song, Y., Yang, L., Yang, S., Ou, T., Liu, Y., & Zhang, Y. (2022). Mendelian randomization study reveals a causal relationship between rheumatoid arthritis and risk for pre-eclampsia. *Frontiers in Immunology*, 13, 1080980. https://doi.org/10.3389/fimmu.2022. 1080980
- Zhang, Y., Zhang, Y., He, P., Ge, F., Huo, Z., & Qiao, G. (2023). The genetic liability to rheumatoid arthritis may decrease hepatocellular carcinoma risk in East Asian population: a Mendelian randomization study. *Arthritis Research & Therapy*, 25, 49. https://doi.org/10.1186/s13075-023-03029-3
- Zink, F., Stacey, S. N., Norddahl, G. L., Frigge, M. L., Magnusson, O. T., Jonsdottir, I., Thorgeirsson, T. E., Sigurdsson, A., Gudjonsson, S. A., Gudmundsson, J., Jonasson, J. G., Tryggvadottir, L., Jonsson, T., Helgason, A., Gylfason, A., Sulem, P., Rafnar, T., Thorsteinsdottir, U., Gudbjartsson, D. F., ... Stefansson, K. (2017). Clonal hematopoiesis, with and without candidate driver mutations, is common in the elderly. *Blood*, 130, 742–752. https://doi.org/10.1182/blood-2017-02-769869