


The role of carbon dioxide in nematode behaviour and physiology

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Review

Cite this article: Banerjee N, Hallem EA (2020). The role of carbon dioxide in nematode behaviour and physiology. *Parasitology* **147**, 841–854. <https://doi.org/10.1017/S0031182019001422>

Received: 11 July 2019

Revised: 4 September 2019

Accepted: 16 September 2019

First published online: 11 October 2019

Key words:

Carbon dioxide; chemotaxis; *C. elegans*; hookworms; nematodes; parasitic nematodes; sensory behaviour; *Strongyloides*

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Abstract

Carbon dioxide (CO₂) is an important sensory cue for many animals, including both parasitic and free-living nematodes. Many nematodes show context-dependent, experience-dependent and/or life-stage-dependent behavioural responses to CO₂, suggesting that CO₂ plays crucial roles throughout the nematode life cycle in multiple ethological contexts. Nematodes also show a wide range of physiological responses to CO₂. Here, we review the diverse responses of parasitic and free-living nematodes to CO₂. We also discuss the molecular, cellular and neural circuit mechanisms that mediate CO₂ detection in nematodes, and that drive context-dependent and experience-dependent responses of nematodes to CO₂.

Introduction

Carbon dioxide (CO₂) is an important sensory cue for animals across diverse phyla, including Nematoda (Lahiri and Forster, 2003; Shusterman and Avila, 2003; Bensafi *et al.*, 2007; Smallegange *et al.*, 2011; Carrillo and Hallem, 2015). While the CO₂ concentration in ambient air is approximately 0.038% (Scott, 2011), many nematodes encounter much higher levels of CO₂ in their microenvironment during the course of their life cycles. For instance, parasitic nematodes may encounter high CO₂ concentrations released from potential hosts as a byproduct of respiration or from the host feces within which they develop (Byrnes *et al.*, 1997; Buszewski *et al.*, 2007; Carrillo and Hallem, 2015). CO₂ concentrations are also high in specific tissues such as the venous bloodstream, lungs and intestine (Jensen and Jorgensen, 1994; Rotbart *et al.*, 2017), suggesting CO₂ may be an important intra-host cue for parasitic nematodes (Hawdon and Schad, 1990; Bekelaar *et al.*, 2018, 2019). Moreover, many free-living nematodes are found in rotting vegetation, where CO₂ levels are often high (Burg and Burg, 1965; Felix and Duveau, 2012). Therefore, nematodes must detect and respond appropriately to elevated CO₂ concentrations to survive, navigate through their microenvironment and propagate.

CO₂ may serve as a beneficial or detrimental cue for nematodes depending on specific circumstances (Carrillo and Hallem, 2015). For instance, in the case of parasitic nematodes, CO₂ may be necessary to promote parasite–host interactions and thus support their parasitic life cycle. For free-living nematodes, high CO₂ levels present in their natural habitats may act as signals for food, predators, pathogens or conspecifics (Carrillo and Hallem, 2015). Because CO₂ is a complex cue that can have either a positive or negative valence, it is not surprising that different species of nematodes have developed distinct behavioural and physiological responses to CO₂. Moreover, many species, both free-living and parasitic, exhibit CO₂ responses that vary with context, previous experience and/or life stage. Recent studies of the free-living model nematode *Caenorhabditis elegans* have provided insight into the cellular and molecular mechanisms that drive and modulate CO₂-evoked responses. In contrast, the mechanisms that promote the diverse responses of parasitic nematodes to CO₂ have not yet been elucidated due to the historic lack of tools required for molecular genetic studies of these worms. However, as a result of recent developments in molecular genetic techniques, we are now in a position to interrogate the neural circuits and molecular signals that promote CO₂ responses in parasitic nematodes. The findings from these studies will enhance our understanding of the role of CO₂ in sculpting parasite–host interactions and may enable the development of novel strategies to combat harmful nematode infections. Here, we review our existing knowledge of how various nematode species respond behaviourally and physiologically to CO₂. We also discuss how CO₂ responsiveness can be modulated based on context, previous experience and life stage.

Responses of mammalian-parasitic nematodes to CO₂

Introduction to parasitic nematodes of mammals

Mammalian-parasitic nematodes infect over a billion people worldwide and are a major cause of morbidity in low-resource areas (Boatin *et al.*, 2012). Infections with soil-transmitted nematodes can cause chronic gastrointestinal distress, stunted growth and cognitive impairment in children, anaemia and even fatality in infants and immunocompromised individuals (Lustigman *et al.*, 2012). Infections with vector-transmitted nematodes can cause severe symptoms such as permanent disfigurement and blindness (Lustigman *et al.*, 2012). Additionally,

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parasitic nematodes that infect livestock are an enormous source of economic burden (Jasmer *et al.*, 2003). Current treatments for infections depend on anthelmintic drugs that reduce the worm burden in heavier infections but do not prevent reinfections, with the result that reinfection is common in endemic areas (Prichard *et al.*, 2012). In addition, drug resistance resulting from mass drug administration is a major challenge for the treatment of nematode-infected livestock (Kumar *et al.*, 2013; Roeber *et al.*, 2013; Emery *et al.*, 2016; Learmount *et al.*, 2016) and is expected to be a concern for the treatment of nematode-infected humans in the near future (Keiser and Utzinger, 2008; Diawara *et al.*, 2013; Repetto *et al.*, 2018). The drugs currently available are also not sufficient to eliminate human infections in all cases, at least by following the administration schedules under practice (Repetto *et al.*, 2018).

Many of these nematodes are gastrointestinal parasites with a developmentally arrested infective larval stage that inhabits the environment and infects hosts following either skin penetration or passive ingestion, depending on the species (Gang and Hallem, 2016; Bryant and Hallem, 2018). The infective larval stages of these species respond robustly to a diverse array of host and environmental sensory cues, including CO₂ (Gang and Hallem, 2016; Bryant and Hallem, 2018). In addition, many parasitic nematodes may rely on sensory cues inside the host body, including CO₂, to re-initiate development upon host entry, direct somatic migration and establish a successful infection (Hawdon and Schad, 1990, 1992; Hawdon *et al.*, 1992; Bekelaar *et al.*, 2018, 2019).

Responses of skin-penetrating nematodes to CO₂

Skin-penetrating nematodes such as the human-parasitic hookworms *Ancylostoma duodenale* and *Necator americanus* and the human-parasitic threadworm *Strongyloides stercoralis* are gastrointestinal parasites that infect hosts as developmentally arrested third-stage larvae (iL3s) (Roberts *et al.*, 2005; Nutman, 2017; Velikkakam *et al.*, 2017). The iL3s are soil-dwelling and actively engage in host seeking using a variety of host-associated sensory cues (Gang and Hallem, 2016; Bryant and Hallem, 2018). These parasites generally have narrow host ranges, infecting only a limited number of host species (Haley, 1961; Bezubik, 1965; Nolan *et al.*, 2007; Viney and Lok, 2007; Viney and Kikuchi, 2017). After invading a host by skin penetration, the iL3s resume development inside the host, a process called activation (Stoltzfus *et al.*, 2012, 2014). The nematodes then migrate through the host body to their final destination, the small intestine, where they reside as parasitic adults (Roberts *et al.*, 2005; Nutman, 2017; Velikkakam *et al.*, 2017). The adults reproduce in the small intestine, and then the eggs or young larvae, depending on the species, exit the host body in feces. The nematodes inhabit the feces until they develop into iL3s (Roberts *et al.*, 2005; Nutman, 2017; Velikkakam *et al.*, 2017). In most species, all of the progeny of the parasitic adults develop directly into iL3s. However, *Strongyloides* species can cycle through one or a limited number of free-living generations on the feces before developmentally arresting as iL3s (Roberts *et al.*, 2005). *Strongyloides stercoralis* can also pass through multiple generations inside the same host through autoinfective cycles (Roberts *et al.*, 2005).

Many skin-penetrating nematodes show behavioural responses to CO₂. For example, iL3s of the dog hookworm *Ancylostoma caninum* display increased nictation in the presence of CO₂ (Granzer and Haas, 1991). Nictation is a specialized behaviour displayed by many parasitic nematodes in which the worm stands on its tail and waves its head in the air to facilitate attachment to mobile hosts (Granzer and Haas, 1991; Bryant and Hallem, 2018). In addition, both *Ancylostoma caninum* and *Strongyloides*

stercoralis iL3s exhibit increased movement when exposed to human breath, and this behaviour is not observed when CO₂ is removed from the breath (Sciacca *et al.*, 2002). Similarly, the human-parasitic hookworms *Ancylostoma duodenale* and *Necator americanus* display increased activity in response to CO₂ in combination with heat and/or moisture (Haas *et al.*, 2005). The similar responses of *Strongyloides stercoralis* and hookworms to CO₂ is particularly notable given their phylogenetic divergence, with *Strongyloides stercoralis* in clade IV and hookworms in clade V (Blaxter and Koutsovoulos, 2015; Blaxter *et al.*, 2016). However, these studies did not look at migration in CO₂ gradients, and whether CO₂ was an attractant or repellent was not clear.

More recent studies demonstrated that skin-penetrating iL3s of the human parasites *Strongyloides stercoralis* and *Ancylostoma ceylanicum* and the rat parasites *Strongyloides ratti* and *Nippostrongylus brasiliensis* are repelled by CO₂ in CO₂-chemotaxis assays (Fig. 1A, B) (Castelletto *et al.*, 2014; Ruiz *et al.*, 2017). A lack of attraction towards CO₂ is consistent with the route of infection of skin-penetrating nematodes, since mammalian skin surfaces emit low concentrations of CO₂ (Alkalay *et al.*, 1971). On the other hand, fecal deposits contain high levels of CO₂ resulting from aerobic respiration of fecal bacteria (Jensen and Jorgensen, 1994; de Lacy Costello *et al.*, 2014; Rotbart *et al.*, 2017), and CO₂ repulsion may drive these iL3s off of host feces and into the environment in search of new hosts.

Responses of passively ingested nematodes to CO₂

Many passively ingested gastrointestinal nematodes have a motile environmental iL3 stage that invades hosts after being swallowed. For example, iL3s of the ruminant parasite *Haemonchus contortus* inhabit the soil and infect after being swallowed by grazing animals (O'Connor *et al.*, 2006). After entering a host, the nematodes exsheath in the rumen and travel to the abomasum, where they develop into parasitic adults (Laing *et al.*, 2013). Similarly, the murine gastrointestinal parasite *Heligmosomoides polygyrus* has an iL3 stage that can infect mice either from feces during coprophagy or from the fur during grooming (Hernandez and Sukhdeo, 1995). Despite their passive route of infection, both *Haemonchus contortus* and *Heligmosomoides polygyrus* actively migrate towards host-associated sensory cues. This suggests that these species use host-associated cues to position themselves in the vicinity of potential hosts, where they are more likely to be ingested (Hernandez and Sukhdeo, 1995; Castelletto *et al.*, 2014; Ruiz *et al.*, 2017; Bryant *et al.*, 2018).

Examination of the CO₂-evoked behaviours of *Haemonchus contortus* and *Heligmosomoides polygyrus* revealed that both species show experience-dependent responses to CO₂ (Fig. 1C) (Castelletto *et al.*, 2014; Ruiz *et al.*, 2017). In the case of *Heligmosomoides polygyrus*, iL3s extracted directly from feces are repelled by CO₂, while iL3s that have been removed from feces for multiple days – a condition designed to mimic the soil environment of iL3s – are attracted to CO₂ (Ruiz *et al.*, 2017). This shift in CO₂ preference appears to occur as a result of the drop in ambient CO₂ levels experienced by the iL3s after they migrate off feces, since cultivating iL3s off feces under high CO₂ conditions (2.5% CO₂) prevents the behavioural switch. The initial repulsion from CO₂ experienced by *Heligmosomoides polygyrus* iL3s on feces may enable them to disperse off of feces and into the environment to host seek. Following a prolonged period without feces, CO₂ attraction may drive them towards new hosts or fresh host feces to increase their chances of host entry through ingestion (Ruiz *et al.*, 2017).

In the case of *Haemonchus contortus*, iL3s directly removed from feces are neutral to CO₂, whereas iL3s that have been

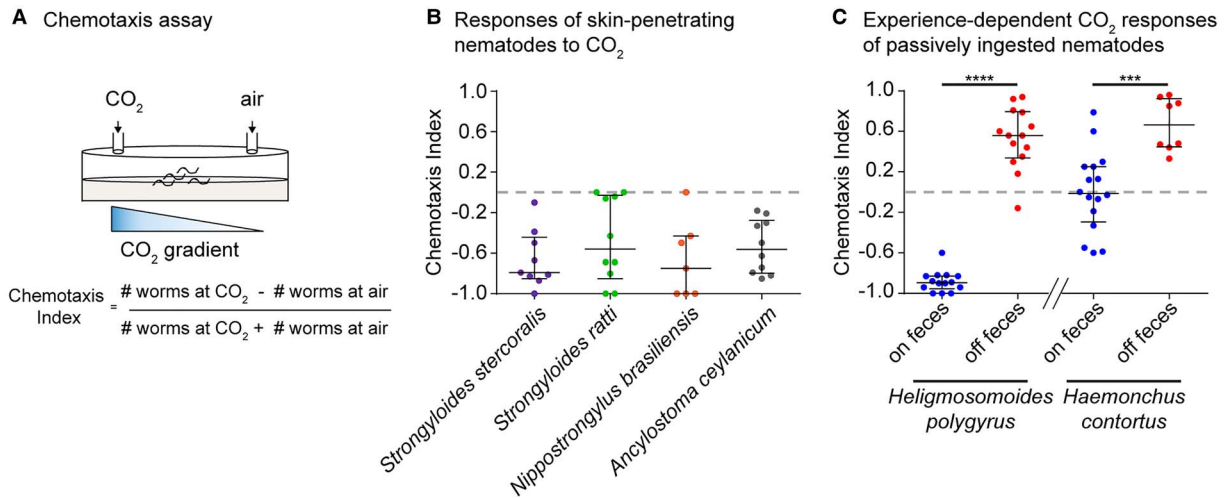


Fig. 1. Behavioural responses of mammalian-parasitic nematodes to CO₂. (A) A CO₂-chemotaxis assay. CO₂ and air are pumped into opposite sides of a 10 cm plate. Infective larvae (iL3s) are placed at the centre and allowed to migrate for 1 h. A chemotaxis index is then calculated according to the formula indicated; a positive index indicates attraction and a negative index indicates repulsion. (B) Behavioural responses of skin-penetrating iL3s to 10% CO₂. All species tested are repelled by CO₂. Data are from Castelletto *et al.* (2014) and Ruiz *et al.* (2017). (C) Experience-dependent changes in CO₂ responsiveness in the passively ingested nematodes *Heligmosomoides polygyrus* and *Haemonchus contortus*. CO₂ responses switch from repulsion (in *Heligmosomoides polygyrus*) or neutral (in *Haemonchus contortus*) to attraction following removal from host feces for days to weeks. *Heligmosomoides polygyrus* was tested with 10% CO₂; *Haemonchus contortus* was tested with 15% CO₂. Figure adapted from Ruiz *et al.* (2017). Graphs show medians and interquartile ranges. *****P* < 0.0001, ****P* < 0.001, Mann-Whitney test for each species.

removed from feces for a week or more are attracted to CO₂ (Castelletto *et al.*, 2014; Ruiz *et al.*, 2017). This experience-dependent shift in CO₂-evoked behaviour may enable the iL3s to migrate towards the mouths of grazing ruminants, whose breath emits high concentrations of CO₂ (Ruiz *et al.*, 2017). In contrast to *Heligmosomoides polygyrus* and *Haemonchus contortus*, the skin-penetrating nematodes *Ancylostoma ceylanicum*, *Strongyloides stercoralis* and *Strongyloides ratti* do not display this flexibility in their behavioural responses to CO₂. Thus, experience-dependent plasticity towards CO₂ may be unique to passively ingested nematodes (Ruiz *et al.*, 2017). However, skin-penetrating nematodes do show other forms of sensory plasticity, including experience-dependent thermal plasticity and temperature-dependent olfactory plasticity (Lee *et al.*, 2016; Bryant *et al.*, 2018). This suggests that experience-dependent responses to CO₂ may not be beneficial for skin-penetrating nematodes, likely because the skin surface of mammals emits only very low levels of CO₂ (Alkalay *et al.*, 1971).

The role of CO₂ in mammalian-parasitic nematode development and physiology

In addition to being a robust behavioural cue for parasitic nematodes, CO₂ is also an important regulator of their development and physiology. For example, CO₂ stimulates exsheathment and activation (exit from the developmentally arrested iL3 stage) in passively ingested ruminant parasites such as *Haemonchus contortus* (Rogers and Sommerville, 1960; Taylor and Whitlock, 1960; Sommerville, 1964; Bekelaar *et al.*, 2018, 2019). However, the requirement for CO₂ during exsheathment varies across species. CO₂ is an absolute requirement for the exsheathment of *Haemonchus contortus* iL3s, whereas CO₂ enhances but is not required for exsheathment in other passively ingested abomasal nematodes (Bekelaar *et al.*, 2018). In the dog hookworm *Ancylostoma caninum*, CO₂ is not required for activation but results in a slight increase in the rate of activation (Hawdon and Schad, 1990).

The role of CO₂ is not limited to exsheathment and activation. CO₂, in combination with O₂, also regulates the development of *Strongyloides ratti* into either free-living adults or iL3s (Taylor

and Weinstein, 1990). In addition, CO₂ stimulates egg hatching in the giant roundworm *Ascaris lumbricoides*, a human-parasitic species that infects when eggs containing developmentally arrested infective larvae are swallowed by hosts as a result of fecal-oral contamination (Fairbairn, 1961; Dold and Holland, 2011). Finally, CO₂ is required for the *in vitro* development of parasitic larvae in the pig roundworm *Ascaris suum* (Douvres and Urban, 1983). Thus, CO₂ influences both behaviour and development in many if not all mammalian-parasitic nematode species.

Responses of entomopathogenic nematodes to CO₂

Introduction to entomopathogenic nematodes

Entomopathogenic nematodes (EPNs) are parasites that infect and kill insects (Dillman and Sternberg, 2012). They are considered beneficial for humans due to their role as biological agents for pest control, and are likely also important for maintaining balanced ecosystems in nature. EPNs of the genera *Heterorhabditis* and *Steinernema* have been successfully employed commercially against insect agricultural pests (Liu *et al.*, 2000; Grewal *et al.*, 2005; Dillman and Sternberg, 2012; Labaude and Griffin, 2018). The geographical distribution of EPNs spans all continents except Antarctica (Hominick, 2002). Some EPNs, such as *Steinernema carpocapsae* and *Heterorhabditis bacteriophora*, are generalists that can infect many different insects; in contrast, other EPNs have very narrow host ranges (Peters, 1996). For example, the specialist *Steinernema scapterisci* specifically infects mole crickets, and the specialist *Steinernema diaprepesi* specifically infects the larval stages of the citrus pest *Diaprepes abbreviatus* (Nguyen and Smart, 1991; Nguyen and Hunt, 2007; Ali *et al.*, 2010). EPNs infect only as third-stage larvae called infective juveniles (IJs); the IJ stage of EPNs is equivalent to the iL3 stage of mammalian-parasitic nematodes (Dillman *et al.*, 2012a). IJs enter their insect hosts through a body orifice such as the mouth, spiracles or anus; IJs of some species can also penetrate directly through the cuticle (Bedding and Molyneux, 1982; Kaya and Gaugler, 1993). The IJs then enter the insect haemocoel and release a bacterial symbiont from their intestine (Bedding and

Molyneux, 1982; Kaya and Gaugler, 1993). Toxins secreted by the nematode and the bacteria kill the insect, typically within 48 h (Kaya and Gaugler, 1993; Lu *et al.*, 2017; Chang *et al.*, 2019). The nematodes then feed on the insect cadaver and complete their parasitic life cycle. The nematodes can cycle through multiple generations in the host cadaver until resources are depleted, at which point new IJs form and disperse into the environment to seek out new hosts (Kaya and Gaugler, 1993).

The role of CO₂ in the host-seeking behaviours of EPNs

The host-seeking strategies of EPNs vary across species. Some species are considered ‘cruisers’ that actively migrate towards stationary hosts, other species are considered ‘ambushers’ that remain relatively stationary and nictate to facilitate attachment to mobile hosts, and still other species use an intermediate strategy (Campbell and Gauger, 1993; Lewis, 2002; Lewis *et al.*, 2006). However, both ambushers and cruisers are capable of migrating towards host-emitted chemosensory cues, suggesting that all EPNs engage in chemosensory-driven navigation towards hosts (Schmidt and All, 1979; Pye and Burman, 1981; O’Halloran and Burnell, 2003; Hallem *et al.*, 2011a; Dillman *et al.*, 2012b; Castelletto *et al.*, 2014; Lee *et al.*, 2016). Some EPNs in the genus *Steinernema* also engage in a unique jumping behaviour where the IJ stands on its tail and then propels itself into the air, presumably to facilitate host attachment as well as transport to new niches (Campbell and Kaya, 1999). Jumping can be stimulated by exposure to host-emitted chemosensory cues (Campbell and Kaya, 1999, 2000; Hallem *et al.*, 2011a; Dillman *et al.*, 2012b).

Many EPNs, including *Heterorhabditis bacteriophora*, *Steinernema carpocapsae*, *Steinernema riobrave*, *Steinernema scapterisci* and *Steinernema glaseri*, are attracted to CO₂ (Fig. 2A) (Gaugler *et al.*, 1980, 1991; Lewis *et al.*, 1993; Robinson, 1995; Hallem *et al.*, 2011a; Dillman *et al.*, 2012b; Lee *et al.*, 2016). This group includes both specialists and generalists, and both ambushers and cruisers. In addition, CO₂ stimulates jumping in *Steinernema carpocapsae*, *Steinernema riobrave* and *Steinernema scapterisci* IJs at concentrations as low as 0.08% (approximately two times higher than atmospheric levels), suggesting that jumping is highly sensitive to environmental CO₂ (Hallem *et al.*, 2011a; Dillman *et al.*, 2012b). Both the attractive responses of EPNs towards the odour of live insect hosts and jumping responses to host odour are decreased when CO₂ is chemically removed using a soda lime filter, illustrating the importance of CO₂ for host seeking (Gaugler *et al.*, 1991; Dillman *et al.*, 2012b). However, the extent to which host attraction is reduced in the absence of CO₂ varies across different EPN–host combinations (Dillman *et al.*, 2012b). Thus, EPNs use CO₂ in combination with host-specific olfactory cues to migrate towards insects. CO₂ also acts synergistically with plant root volatiles to attract some EPNs to plants infested with insects (Turlings *et al.*, 2012).

Like some mammalian-parasitic nematodes, some EPNs exhibit plasticity in their olfactory responses to CO₂. For example, the IJs of *Steinernema scapterisci* are repelled by CO₂ immediately after emergence from the insect host, but their response shifts to robust attraction over the course of weeks (Fig. 2B) (Lee *et al.*, 2016). In addition, the rate at which this shift occurs varies depending on the cultivation temperature of the IJs, with the shift occurring more rapidly in IJs cultured at 15 °C relative to IJs cultured at 25 °C (Lee *et al.*, 2016). The change in CO₂ preference correlates with a corresponding change in responses to host odours, including the odour of its natural host, the mole cricket (Lee *et al.*, 2016). The strong repulsion of newly emerged *Steinernema scapterisci* IJs to CO₂ and host odours may serve as a dispersal mechanism to drive them towards new niches. The

mechanisms that drive the change in CO₂ preference following host emergence in *Steinernema scapterisci* have not yet been elucidated. An intriguing possibility is that the CO₂ repulsion exhibited by IJs immediately following host emergence could result from the elevated levels of CO₂ experienced inside the decaying insect cadaver, similar to the way in which CO₂ repulsion in *Heligmosomoides polygyrus* iL3s results from the elevated levels of CO₂ experienced on mammalian feces (Ruiz *et al.*, 2017). However, additional experiments will be necessary to determine whether the change in CO₂ preference over time (or age) in *Steinernema scapterisci* is in fact regulated by ambient CO₂ levels.

Responses of plant-parasitic nematodes to CO₂

Introduction to plant-parasitic nematodes

Plant-parasitic nematodes (PPNs) are a major cause of agricultural crop damage throughout the world. It has been estimated that PPNs are responsible for approximately 100 billion dollars of crop loss per year worldwide (Jasmer *et al.*, 2003; Wrather *et al.*, 2003). Of over 4100 species of PPNs that have been identified (Decraemer and Hunt, 2006), the ones that cause the most severe economic loss are the nematodes that infect the roots of major agricultural crops (Bernard *et al.*, 2017). These PPNs prevent water and nutrient uptake by plant roots, which results in greatly reduced crop quality and yield (Bernard *et al.*, 2017).

Responses of PPNs to CO₂

CO₂ is ubiquitously produced by the roots of plants. Several studies have demonstrated an important role for CO₂ in mediating attraction of PPNs to their host plants. For example, the stem nematode *Ditylenchus dipsaci*, which infects onion and garlic, migrates towards CO₂ (Klingler, 1972; Viglierchio, 1990). Many other PPNs, including species from the genera *Ditylenchus*, *Meloidogyne*, *Heterodera* and *Pratylenchus*, are also attracted to CO₂ (Johnson and Viglierchio, 1961; Prot, 1980; McCallum and Dusenbery, 1992; Robinson, 1995). In the case of *Meloidogyne incognita*, attraction to tomato root volatiles appears to be due to the presence of O₂ and CO₂ in the volatile mix (McCallum and Dusenbery, 1992). However, a more recent study found that for *Meloidogyne hapla*, the attractant is not CO₂ itself but rather the low pH environment created by dissolved CO₂ (Wang *et al.*, 2009). In the case of the pine wilt nematode *Bursaphelenchus xylophilus*, the fourth-stage juveniles (J_{IVS}) are repelled by CO₂. CO₂ repulsion by J_{IVS} plays an important role in dispersal from its insect vector, the pine sawyer beetle, into the pine tree (Wu *et al.*, 2019). *Bursaphelenchus xylophilus* J_{IVS} enter the beetle tracheal system, where they are transported by the beetle to new pine trees. As the beetle matures and feeds on the pine tree, CO₂ levels in the beetle tracheal system increase. Once CO₂ levels reach a certain concentration, CO₂ repulsion drives the J_{IVS} out of the beetle spiracles and into the pine tree (Wu *et al.*, 2019). Thus, the responses of PPNs to CO₂ vary greatly across species. A better understanding of how other PPNs respond to CO₂ may enable the development of new biocontrol strategies.

Responses of free-living nematodes to CO₂

Introduction to free-living nematodes

Free-living nematodes are found in a wide range of ecological habitats. These include various types of soil, sediment and organic matter, as well as marine and freshwater environments. Free-living nematodes use a wide variety of sensory cues to navigate their environment in search of food and mates, and to escape

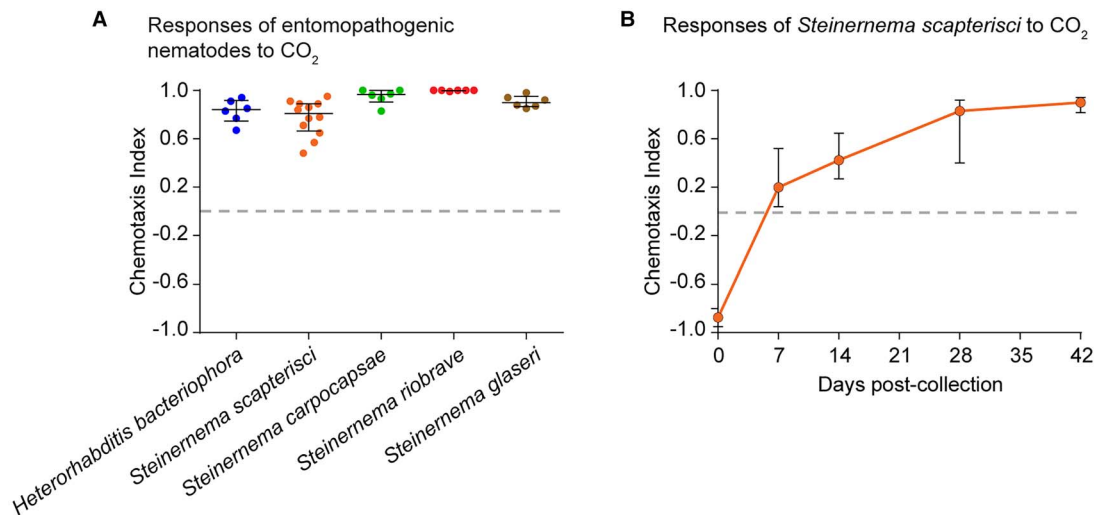


Fig. 2. Behavioural responses of entomopathogenic nematodes (EPNs) to CO₂. (A) Behavioural responses of the infective juveniles (IJs) of various EPN species to CO₂ in a chemotaxis assay (Fig. 1A). All EPNs tested showed attraction to 2.5% CO₂. Data are from Dillman *et al.* (2012b) and Hallem *et al.* (2011a). (B) Age-dependent changes in the CO₂ preferences of *Steinernema scapterisci* IJs. IJs were grown at room temperature (approximately 22 °C) and then incubated at 15 °C until the indicated time points (days post-collection). CO₂ responsiveness changes from repulsion to attraction with age. IJs were tested with 1% CO₂. Data are from Lee *et al.* (2016). The graph shows medians and interquartile ranges.

from predators and pathogens. CO₂ is universally present in terrestrial and aquatic habitats, and may serve as an important cue for survival and propagation of these nematodes. The most well-studied free-living nematode is the model worm *Caenorhabditis elegans*. *C. elegans* is commonly found in microbe-rich environments such as those of fallen rotting fruits (Felix and Duveau, 2012), where CO₂ is produced as one of many microbial byproducts. Consequently, *C. elegans* displays several behavioural and physiological responses to CO₂.

Behavioural responses of *C. elegans* to carbon dioxide

The first studies of CO₂ responsiveness in *C. elegans* demonstrated that these worms undergo rapid changes in locomotion in response to changes in CO₂ concentrations (Dusenbery, 1985). These responses are characterized by an overall decrease in movement and an increase in turning frequency (Dusenbery, 1985). A more recent study examining the effects of acute CO₂ exposure found that freely moving well-fed adults reverse rapidly when their head is exposed to high levels of CO₂, indicating that CO₂ is an aversive cue for well-fed *C. elegans* adults (Hallem and Sternberg, 2008). In addition, well-fed *C. elegans* adults avoid high CO₂ areas when allowed to migrate along a CO₂ gradient in a CO₂-chemotaxis assay (Fig. 3A) (Bretscher *et al.*, 2008). For well-fed adults, CO₂ may indicate the presence of potential predators or pathogens, and repulsion from CO₂ may function as an escape mechanism.

As is the case for some parasitic nematodes, CO₂ responsiveness in *C. elegans* is subject to experience-dependent plasticity. In the case of *C. elegans*, one of the factors that influences CO₂ responsiveness is the worm's nutritional status. Depriving adults of food suppresses CO₂ avoidance behaviour (Bretscher *et al.*, 2008; Hallem and Sternberg, 2008). Moreover, as *C. elegans* adults are starved, CO₂ response shifts from repulsion to attraction (Fig. 3A) (Rengarajan *et al.*, 2019). The effects of starvation can be reversed when the animals are re-exposed to food (Rengarajan *et al.*, 2019). The shift from CO₂ repulsion to CO₂ attraction in starved worms may be beneficial for survival, since starved animals must find food in order to survive and bacterial food emits CO₂. CO₂ attraction by starved animals may also reflect an increased tolerance for risk taking; CO₂ attraction is

an inherently risky behaviour, since both predators and pathogens of *C. elegans* emit CO₂ (Felix and Duveau, 2012; Brandt and Ringstad, 2015; Schulenburg and Felix, 2017).

CO₂ responsiveness in *C. elegans* adults is also modulated by recently experienced environmental CO₂ and oxygen (O₂) levels, as well as immediate O₂ context. For example, animals exposed to elevated CO₂ levels (2.5% CO₂) become robustly attracted to CO₂ over the course of hours in a reversible manner (Fig. 3A) (Guillermin *et al.*, 2017). Prior exposure to low O₂ levels also suppresses CO₂ avoidance in adults, an effect that depends on the hypoxia inducible factor gene *hif-1* (Fig. 3B) (Bretscher *et al.*, 2008). In addition, whether animals have been pre-exposed to low O₂ affects their responsiveness to CO₂ stimuli under some conditions (Fenk and de Bono, 2017). CO₂ responsiveness is also modulated by ambient O₂ levels such that animals assayed under low O₂ conditions are more strongly repelled by CO₂ than animals assayed under high O₂ conditions (Carrillo *et al.*, 2013; Kodama-Namba *et al.*, 2013). *C. elegans* prefers O₂ concentrations below atmospheric (Gray *et al.*, 2004); however, when exposed to opposing gradients of O₂ and CO₂, the avoidance response to high CO₂ dominates over the avoidance response to high O₂ (Bretscher *et al.*, 2008). In nature, *C. elegans* is found in rotting vegetation (Felix and Braendle, 2010; Schulenburg and Felix, 2017), where both O₂ and CO₂ concentrations fluctuate. Moreover, both O₂- and CO₂-sensing pathways control foraging behaviour (Bendesky *et al.*, 2011; Milward *et al.*, 2011; Juozaityte *et al.*, 2017). Thus, the interplay between O₂- and CO₂-evoked behaviours likely contributes to the ability of *C. elegans* to navigate the complex organic environments it inhabits.

CO₂ responsiveness is also modulated by the presence or absence of food, and prior temperature experience (Bretscher *et al.*, 2008, 2011; Kodama-Namba *et al.*, 2013). In the case of temperature-dependent modulation of CO₂ responsiveness, animals cultivated at 22 °C show enhanced repulsion to 1% CO₂ when assayed at 15 °C compared with 22 °C (Kodama-Namba *et al.*, 2013), suggesting an interaction between recent temperature experience and CO₂ sensing. The ecological significance of this interaction is not yet clear, but it suggests that CO₂ preferences may vary on a diurnal cycle as the ambient temperature fluctuates. Thus, CO₂-evoked behaviours are regulated by multiple sensory

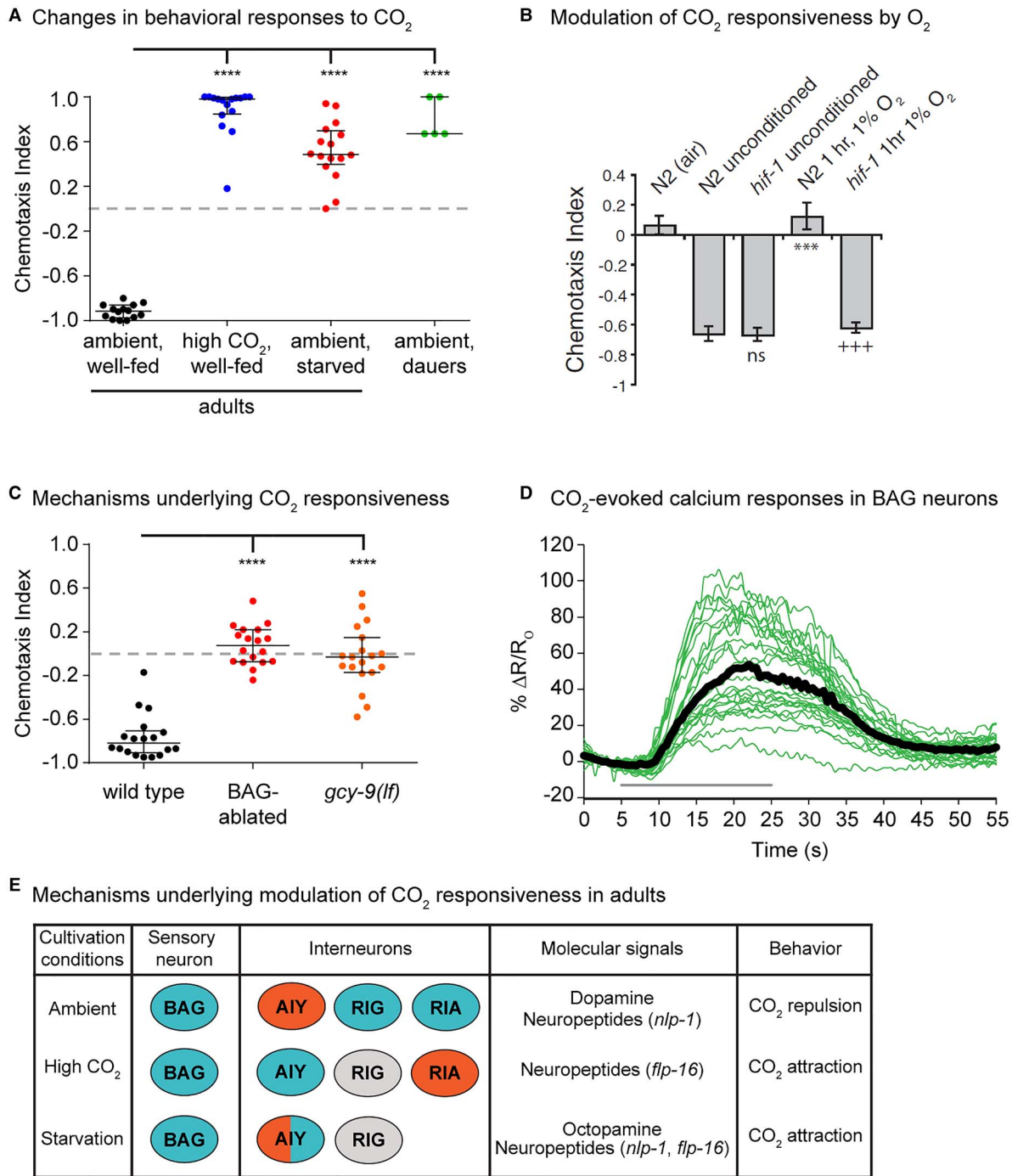


Fig. 3. Behavioural responses of *C. elegans* to CO₂. (A) Responses of wild-type *C. elegans* adults and dauers to CO₂ in a chemotaxis assay (Fig. 1A). Dauers are developmentally arrested third-stage larvae that are similar to parasitic iL3s and IJs (Hotez *et al.*, 1993; Viney *et al.*, 2005; Crook, 2014). Animals were either well-fed adults cultivated at ambient CO₂, well-fed adults cultivated at high CO₂, starved adults cultivated at ambient CO₂ or dauer larvae cultivated at ambient CO₂. Adults were tested in a 20 min assay; dauer larvae were tested in a 1 h assay. Responses shown are to 2.5% CO₂ (for adults cultivated at high CO₂) or 10% CO₂ (for all other conditions). For the high CO₂ condition, adults were cultivated at 2.5% CO₂ for one generation prior to the assay. For the starvation condition, adults were starved for 3 h prior to the assay. Data are from Guillermin *et al.* (2017), Rengarajan *et al.* (2019) and Hallem *et al.* (2011a). *****P* < 0.0001, one-way ANOVA with Dunnett's post-test. The graph shows medians and interquartile ranges. (B) Previously experienced hypoxic conditions modulate CO₂ responsiveness in *C. elegans* adults. Animals cultivated at low (1%) oxygen for 1 h prior to assays showed decreased CO₂ avoidance. This change is mediated by hypoxia inducible factor-1 (HIF-1), since *hif-1* mutants are not affected by prior O₂ exposure. ns, not significant relative to N2 (wild-type); ****P* < 0.001 relative to N2; +++*P* < 0.001 relative to N2 exposed to 1% O₂ for 1 h. The N2 (air) condition represents a control condition in which animals were not exposed to a CO₂ gradient. The graph shows means and SEMs. Figure is from Bretscher *et al.* (2008), copyright 2008 National Academy of Sciences. (C) The BAG sensory neurons and GCY-9 are required for CO₂ repulsion in *C. elegans*. BAG-ablated animals and *gcy-9* loss-of-function (*lf*) mutants are neutral to CO₂. *****P* < 0.0001, one-way ANOVA with Dunnett's post-test. The graph shows medians and interquartile ranges. Data are from Carrillo *et al.* (2013). (D) Calcium activity in BAG neurons in response to 10% CO₂, as measured using the ratio-metric calcium indicator yellowameleon 3.60 (YC3.60). Green traces show responses of individual neurons; black line shows median response. Data are from Rengarajan *et al.* (2019). (E) A model for experience-dependent modulation of CO₂ responsiveness in *C. elegans* adults. Under ambient conditions, CO₂ repulsion is mediated by activation of the RIA and RIG interneurons and inhibition of the AIY interneurons, and by both dopamine and neuropeptide signalling involving the neuropeptide gene *nlp-1*. CO₂ attraction in animals cultivated at high CO₂ is mediated by activation of AIY, inhibition of RIA and silencing of RIG, as well as neuropeptide signalling involving the neuropeptide gene *flp-16*. Finally, CO₂ attraction in starved adults is mediated by silencing of RIG and by a change in AIY responses such that activation and inhibition are observed with approximately equal frequency. Octopamine signalling and neuropeptide signalling via the neuropeptide genes *nlp-1* and *flp-16* also promote CO₂ attraction during starvation. Blue = excitatory activity, orange = inhibitory activity, grey = silencing of activity. Figure is adapted from Guillermin *et al.* (2017) and Rengarajan *et al.* (2019).

modalities, resulting in both context-dependent and experience-dependent responses to CO₂.

CO₂ can also elicit behavioural changes in *C. elegans* that are independent of changes in locomotion. For example, exposing *C. elegans* to high CO₂ levels (5% CO₂) inhibits egg-laying behaviour, at least transiently (Fenk and de Bono, 2015). Modulation of egg-laying behaviour by high CO₂ levels may prevent animals from exposing their progeny to unfamiliar environmental conditions. Feeding behaviour is also altered by CO₂ such that brief exposure to high CO₂ levels causes an acute reduction in pharyngeal pumping (Sharabi *et al.*, 2009). Well-fed animals stop pharyngeal pumping when exposed to a lower concentration of CO₂ than starved animals, suggesting that this behavioural change is dependent on the nutritional state of the animal (Sharabi *et al.*, 2009).

Finally, *C. elegans* exhibits life-stage-specific responses to CO₂. *C. elegans* dauers, which are developmentally arrested third-stage larvae that are similar to the iL3 and IJ stages of parasitic nematodes (Hotez *et al.*, 1993; Viney *et al.*, 2005; Crook, 2014), are attracted to CO₂ (Fig. 3A) (Hallem *et al.*, 2011a). Dauer larvae form when food is limited or environmental conditions are otherwise unfavourable (Hu, 2007), and CO₂ attraction by dauers may enable them to locate bacterial food sources. In addition, dauers associate with invertebrates such as slugs, snails and isopods, which act as carriers that facilitate their dispersal to new niches (Baird, 1999; Caswell-Chen *et al.*, 2005; Lee *et al.*, 2012; Schulenburg and Felix, 2017). Thus, CO₂ attraction may also enable dauers to locate and associate with invertebrate carriers.

Effects of CO₂ on *C. elegans* development and physiology

High levels of CO₂ can also elicit physiological changes in *C. elegans*. Cultivating animals at CO₂ levels above 9% CO₂ decreases brood size and significantly slows the rate of embryonic development (Sharabi *et al.*, 2009). This developmental delay is not caused by any obvious reduction in the overall health of the embryos or the adults that develop from these embryos. In addition, chronic exposure (>4 days) to 19% CO₂ causes defects in overall body muscle morphology, resulting in long-lasting movement defects (Sharabi *et al.*, 2009). Nevertheless, cultivation at 19% CO₂ extends lifespan, an effect that appears to be independent of the inhibitory effect of CO₂ on egg laying (Sharabi *et al.*, 2009). The lifespan extension induced by exposure to high CO₂ also may be independent of the dietary restriction pathway, since *eat-2* mutants, which show reduced pharyngeal pumping, show increased lifespan at high CO₂ (Sharabi *et al.*, 2009). Thus, CO₂ has diverse effects on *C. elegans* development, physiology and behaviour.

Behavioural and physiological responses of other free-living nematodes to CO₂

Other free-living nematodes show diverse responses to CO₂. For example, the free-living marine nematode *Adoncholaimus thalassophygas* is attracted to CO₂ (Riemann and Schrage, 1988). This effect is not due to a general decrease in the pH of the medium, since the addition of hydrochloric acid did not elicit a similar attractive response (Riemann and Schrage, 1988). CO₂ is emitted from bacteria present in decaying carcasses found in sediments and may act as a food signal for these nematodes. The necromenic nematode *Pristionchus pacificus*, which represents an evolutionary intermediate between free-living and parasitic nematodes, displays acute CO₂ avoidance (Hallem and Sternberg, 2008). In contrast, the free-living nematodes *Caenorhabditis briggsae*, *Caenorhabditis angaria* and *Panagrellus redivivus* do not respond

to CO₂ in acute avoidance assays (Hallem and Sternberg, 2008). However, ambient CO₂ concentrations play a prominent role in regulating reproduction in *Panagrellus redivivus*. Under low O₂ conditions, the brood size of *Panagrellus redivivus* increases in response to an increase in CO₂ concentration from 0 to 5% (Hansen and Buecher, 1970). The effects of CO₂ on nematode physiology also vary greatly across species. At the extreme, nematodes have been isolated from volcanic gas vents, where CO₂ levels can reach 100%; these nematodes can survive under 100% CO₂ conditions for at least 5 days (Pilz and Hohberg, 2015). Thus, CO₂ has species-specific effects on nematode behaviour and physiology.

Cellular and molecular mechanisms of CO₂ responsiveness in nematodes

Cellular mechanisms of CO₂ responsiveness in *C. elegans*

The primary CO₂-sensing neurons in *C. elegans* are the paired BAG neurons in the head. Ablation of the BAG neurons abolishes both CO₂ avoidance in adults (Hallem and Sternberg, 2008) and CO₂ attraction in dauers (Fig. 3C) (Hallem *et al.*, 2011a). Exposing animals to varying concentrations of CO₂ produces dose-dependent calcium activity in the BAG neurons *in vivo* (Fig. 3D) (Hallem *et al.*, 2011b). In addition, isolated BAG neurons derived from *C. elegans* embryos respond to CO₂ *in vitro*, suggesting that BAG neurons are intrinsically sensitive to CO₂ (Smith *et al.*, 2013). The calcium responses in isolated BAG neurons in culture are independent of carbonic anhydrase activity, indicating that these neurons can sense molecular CO₂. Moreover, a majority of isolated BAG neurons do not respond to pH, although responses to low pH are detectable in some isolated BAG neurons (Smith *et al.*, 2013). However, the role of BAG neurons is not limited to promoting CO₂ responses. The BAG neurons also sense O₂ (Zimmer *et al.*, 2009). Furthermore, they play a role in establishing food odour preferences and in foraging behaviour (Harris *et al.*, 2014; Juozaityte *et al.*, 2017). In addition to BAG neurons, other sensory neurons including ASE, AFD, AWC, ASJ, ASK, ASH and ADL also exhibit CO₂-evoked calcium activity and contribute to CO₂ responsiveness (Bretscher *et al.*, 2011; Fenk and de Bono, 2015). The interneurons AIY, RIG, RIA and AIZ act downstream of BAG neurons to mediate CO₂-evoked behaviour (Kodama-Namba *et al.*, 2013; Guillermin *et al.*, 2017). The AIA interneurons also show CO₂-evoked activity and are involved in CO₂ responsiveness (Fenk and de Bono, 2015). In well-fed adults, CO₂ repulsion is correlated with activation of RIA, RIG and AIZ, and inhibition of AIY (Fig. 3E) (Guillermin *et al.*, 2017).

Molecular mechanisms of CO₂ responsiveness in *C. elegans*

The detection of CO₂ by the BAG neurons requires the receptor-type guanylate cyclase GCY-9. Animals with loss-of-function mutations in *gcy-9* are insensitive to CO₂ in behavioural assays (Fig. 3C) (Hallem *et al.*, 2011b). Moreover, CO₂-evoked calcium transients in the BAG neurons require GCY-9, and ectopic expression of GCY-9 confers CO₂ sensitivity to other sensory neurons (Hallem *et al.*, 2011b; Brandt *et al.*, 2012; Carrillo *et al.*, 2013). The expression of GCY-9 in the BAG neurons requires the E26 transformation-specific (ETS)-domain transcription factor ETS-5, and *ets-5* mutants fail to avoid CO₂ (Guillermin *et al.*, 2011; Brandt *et al.*, 2012). Both ETS-5 and the SoxD transcription factor EGL-13 are also required more generally for normal differentiation of the BAG neurons (Guillermin *et al.*, 2011; Brandt *et al.*, 2012; Petersen *et al.*, 2013). The Toll-like receptor TOL-1 is also required for normal BAG neuron differentiation,

and *tol-1* mutants are defective in pathogen avoidance behaviour as a result (Brandt and Ringstad, 2015).

In addition to GCY-9, the cGMP signalling pathway that mediates CO₂ detection consists of the cGMP-gated cation channel TAX-2/TAX-4 (Bretscher *et al.*, 2008; Hallem and Sternberg, 2008). Glutamate signalling and neuropeptide signalling are also required for BAG-mediated responses to CO₂ (Guillermin *et al.*, 2017). BAG neurons are glutamatergic (Serrano-Saiz *et al.*, 2013), and well-fed adults lacking the vesicular glutamate transporter EAT-4 show neutral responses to CO₂ (Guillermin *et al.*, 2017). BAG neurons also release neuropeptides, and well-fed adults lacking the BAG-expressed FMRamide-like neuropeptide FLP-17 do not respond to CO₂ (Guillermin *et al.*, 2017). In addition, several other signalling molecules, including the calcineurin subunits TAX-6 and CNB-1, the regulator of G-protein signalling RGS-3 and the nuclear hormone receptor NHR-49 are known to regulate CO₂ response (Hallem and Sternberg, 2008). The microRNA *mir-791* is also required for the normal CO₂-evoked calcium activity of the BAG neurons (Drexel *et al.*, 2016).

Mechanisms underlying the context-dependent modulation of CO₂ responsiveness by O₂

The extent to which CO₂ responsiveness is regulated by ambient O₂ levels depends on the neuropeptide Y receptor NPR-1 (McGrath *et al.*, 2009; Carrillo *et al.*, 2013; Kodama-Namba *et al.*, 2013). The laboratory wild-type (N2) strain of *C. elegans* contains a gain-of-function mutation in the *npr-1* gene that confers CO₂ avoidance on well-fed adults regardless of ambient O₂ levels. However, animals containing loss-of-function (*lf*) mutations in *npr-1* and animals carrying the natural variant of *npr-1* avoid CO₂ under low O₂ conditions but do not respond to CO₂ at normal atmospheric O₂ levels (21% O₂) (Carrillo *et al.*, 2013; Kodama-Namba *et al.*, 2013). The gain-of-function NPR-1 variant in N2 animals suppresses the activity of the O₂-sensing URX neurons to promote CO₂ avoidance regardless of ambient O₂ levels. In animals containing an *npr-1(lf)* mutation or a natural variant of *npr-1*, the URX neurons are tonically active under high O₂ conditions and inhibit CO₂ avoidance at high O₂. The RIA interneurons appear to act downstream of URX to partially mediate its effects on the CO₂ circuit (Kodama-Namba *et al.*, 2013). In addition, the neuroglobin gene *glb-5* also acts *via* the URX neurons to modulate CO₂ responsiveness as a function of ambient O₂ levels (McGrath *et al.*, 2009; Kodama-Namba *et al.*, 2013).

Mechanisms underlying the experience-dependent modulation of CO₂ responsiveness

The mechanisms underlying experience-dependent modulation of CO₂ responsiveness in *C. elegans* have been elucidated in some detail. The shift in CO₂ response from repulsion to attraction when animals are moved from low CO₂ to high CO₂ cultivation conditions results from the differential activity of a single set of interneurons downstream of the BAG sensory neurons (Guillermin *et al.*, 2017). In animals that were previously cultivated at low CO₂, CO₂ exposure inhibits the AIY interneurons and activates the RIA and RIG interneurons. In contrast, in animals that have been cultivated at high CO₂, CO₂ exposure activates AIY and inhibits RIA. Moreover, RIG is silenced such that it no longer responds to CO₂ (Fig. 3E). Thus, CO₂ response is not determined by whether an 'attractive' or 'repulsive' pathway is activated; rather, it is determined by experience-dependent modulation of interneuron activity in a single pathway (Guillermin *et al.*, 2017). A number of neuropeptides also differentially modulate CO₂ responsiveness in animals cultured under high vs low CO₂ conditions (Fig. 3E) (Guillermin *et al.*, 2017).

The shift from CO₂ repulsion to CO₂ attraction that occurs during starvation also arises due to the differential activities of the AIY and RIG interneurons (Rengarajan *et al.*, 2019). In starved animals, RIG is silenced and AIY shows stochastic responses such that CO₂ evokes activating and inhibiting responses with approximately equal frequency (Fig. 3E). At the molecular level, whether CO₂ is attractive or repulsive is regulated by biogenic amine signalling. Dopamine promotes CO₂ avoidance in well-fed animals by promoting activation of RIG and inhibition of AIY, while octopamine promotes CO₂ attraction in starved animals by promoting activation of AIY (Fig. 3E) (Rengarajan *et al.*, 2019). Thus, the CO₂ circuit is modulated during starvation by opposing biogenic amine signals. Neuropeptide signalling also regulates CO₂ responsiveness during starvation (Fig. 3E) (Rengarajan *et al.*, 2019). Finally, CO₂ attraction in dauer larvae is less well understood but is regulated at least in part by neuropeptide signalling (Lee *et al.*, 2017).

Molecular and cellular mechanisms underlying other CO₂-evoked behaviours

Some of the molecular and cellular mechanisms that mediate the effects of CO₂ on other behaviours in *C. elegans* have also been elucidated. For instance, CO₂-evoked activity in the AWC sensory neurons triggers a cGMP signalling pathway that ultimately inhibits the activity of the HSN neurons, resulting in the inhibition of egg-laying behaviour (Fenk and de Bono, 2015). Antagonistic effects of the BAG neurons and the URX neurons regulate lifespan in *C. elegans*, resulting in increased longevity in BAG-ablated animals (Liu and Cai, 2013). Mutations in the c-Jun N-terminal kinase (JNK) signalling pathway genes *jnk-1* and *kgb-2* suppress CO₂-induced fertility defects, indicating that JNK signalling may be involved in regulating fertility in response to CO₂ (Vadasz *et al.*, 2012).

Unanswered questions regarding CO₂ responsiveness in C. elegans

Although the mechanisms underlying CO₂ responsiveness in *C. elegans* have been elucidated in appreciable detail, several questions remain unexplored. For example, more information is needed to fully understand how the differential flow of information from BAG neurons to downstream interneurons generates experience-dependent plasticity of CO₂-evoked behaviour. One intriguing possibility is that the BAG neurons express or release different neurotransmitters or neuropeptides in response to CO₂ under varying conditions. Consistent with this possibility, the BAG neurons modulate the expression of FLP-19 neuropeptides as a function of their CO₂-evoked activity (Rojo Romanos *et al.*, 2017). In addition, the interneurons that act downstream of other CO₂-sensing neurons have not been identified. Finally, the CO₂ microcircuit that drives CO₂ attraction in dauers remains poorly understood, although it appears to involve dauer-specific, gap-junction-mediated signalling between the BAG neurons and the downstream AIB interneurons (Bhattacharya *et al.*, 2019). In future studies, it will be interesting to determine whether the same set of neurons or a distinct set of neurons promotes CO₂ attraction in dauers. A better understanding of the neural circuits and signalling pathways that regulate CO₂ responsiveness as a function of experience, context and life stage will provide important insights into how a single sensory cue can give rise to diverse behavioural responses in an ethologically-appropriate manner.

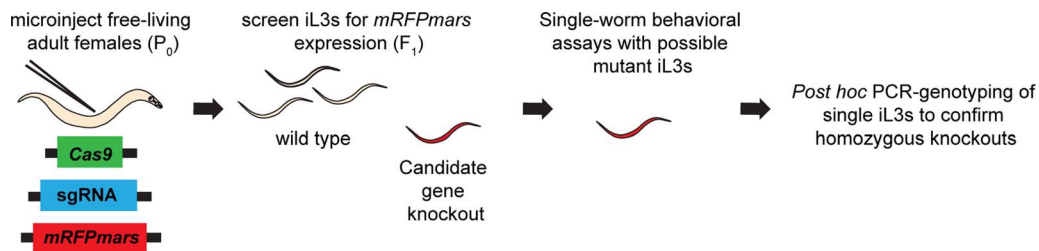


Fig. 4. Targeted mutagenesis in *Strongyloides stercoralis*. (A) Strategy for CRISPR/Cas9-mediated targeted mutagenesis in *Strongyloides stercoralis*. Plasmid vectors encoding Cas9, the single guide RNA (sgRNA) for the gene of interest and a repair template for homology-directed repair encoding an mRFPmars reporter are introduced into *Strongyloides stercoralis* free-living adult females (P₀) by gonadal microinjection. The iL3 progeny (F₁) from microinjected females are screened for mRFPmars expression, indicative of a possible disruption of the gene of interest. iL3s are then tested in single-worm chemotaxis assays and genotyped post hoc for homozygous disruption of the gene of interest. Figure is adapted from Gang *et al.* (2017).

Mechanisms underlying CO₂ responsiveness in other nematodes

The anatomy and function of nematode sensory neurons are generally conserved across species (Ashton *et al.*, 1995, 1999; Lopez *et al.*, 2000; Li *et al.*, 2000a, 2000b, 2001; Bhopale *et al.*, 2001; Forbes *et al.*, 2004; Ketschek *et al.*, 2004; Ashton *et al.*, 2007; Bumbarger *et al.*, 2007; Srinivasan *et al.*, 2008; Bumbarger *et al.*, 2009; Zhu *et al.*, 2011; Hallem *et al.*, 2011a), making it possible to use knowledge of CO₂ responsiveness in *C. elegans* as a starting point for launching investigations into the mechanisms of CO₂ responsiveness in parasitic nematodes. In the case of both the necromenic nematode *Pristionchus pacificus* and the EPNs *Heterorhabditis bacteriophora* and *Steinernema carpocapsae*, BAG neurons were identified on the basis of conserved neuroanatomical position and shown to be required for behavioural responses to CO₂ by laser ablation analyses (Hallem and Sternberg, 2008; Hallem *et al.*, 2011a). BAG-ablated *Pristionchus pacificus* adults do not show acute CO₂ avoidance, and BAG-ablated *Heterorhabditis bacteriophora* and *Steinernema carpocapsae* IJs do not show CO₂ attraction (Hallem *et al.*, 2011a). In addition, CO₂-evoked jumping behaviour in *Steinernema carpocapsae* requires the BAG neurons (Hallem *et al.*, 2011a). Thus, the neural circuits that mediate CO₂ response are at least partly conserved across nematode species. However, the interneurons that operate downstream of BAG neurons to mediate CO₂ responsiveness in other nematode species have not yet been identified. Moreover, nothing is currently known about the neural circuits and molecular signals that promote CO₂ responsiveness in mammalian-parasitic nematodes. In future studies, it will also be interesting to determine whether similar or distinct mechanisms operate in *C. elegans* and parasitic nematodes to modulate CO₂ responses depending on context, previous experience or life stage.

Directions for future research

A major focus going forward will be on elucidating the cellular and molecular mechanisms underlying CO₂ responsiveness in mammalian-parasitic nematodes. The identification of the neural mechanisms that drive or regulate the CO₂ responses of mammalian-parasitic nematodes both inside and outside the host could lead to the identification of new drug targets or new strategies for nematode control. Until recently, investigations into the mechanisms underlying sensory behaviours in parasitic nematodes were limited to laser ablation analysis due to the dearth of resources and tools required for the genetic manipulation of these parasites. Laser ablation analysis has been used to establish the function of a number of different sensory neurons in mammalian-parasitic nematodes, including *Strongyloides*

stercoralis, hookworms and *Haemonchus contortus* (Ashton *et al.*, 1998; Lopez *et al.*, 2000; Li *et al.*, 2000b; Bhopale *et al.*, 2001; Forbes *et al.*, 2004; Ketschek *et al.*, 2004; Nolan *et al.*, 2004; Ashton *et al.*, 2007). However, several recent advances have facilitated the study of gene function and the genetic basis of sensory behaviours in these parasites. High-quality genome sequences of several nematode species have been identified and are readily accessible (Mitreva *et al.*, 2007; Brindley *et al.*, 2009; Hunt *et al.*, 2016; Howe *et al.*, 2017; International Helminth Genomes, 2019). Transcriptomic data are also now available for many parasitic nematode species (Jex *et al.*, 2019), which may significantly advance the study of gene expression and the identification of novel signalling pathways that drive sensory behaviours.

The most genetically tractable parasitic nematodes are *Strongyloides stercoralis* and *Strongyloides ratti*. *Strongyloides stercoralis* and *Strongyloides ratti* are more readily amenable to genetic manipulation than other parasitic nematodes because they can undergo one free-living generation (Viney, 1999, 2006; Lok, 2007). Foreign DNA can be introduced by gonadal microinjection into free-living adults using techniques based on those originally developed for *C. elegans* (Evans, 2006). Most other mammalian-parasitic nematodes lack a free-living generation, which makes it difficult to introduce foreign DNA into these worms. *Strongyloides stercoralis* is a human parasite that infects approximately 370 million people worldwide (Page *et al.*, 2018) and is therefore of direct interest as a human pathogen; additionally, *Strongyloides stercoralis* is of interest as a model for other human-parasitic nematodes such as hookworms that cannot be genetically manipulated.

Transgenic nematodes can be generated by introducing plasmid DNA containing exogenous genes; these genes are then expressed as extrachromosomal arrays in the F₁ progeny of the microinjected adults (Lok and Massey, 2002; Li *et al.*, 2006, 2011; Junio *et al.*, 2008; Lok and Artis, 2008; Lok, 2012; Shao *et al.*, 2012; Lok *et al.*, 2017; Shao *et al.*, 2017). This technique can be potentially used to express any gene of choice, including those required for genetic ablation or silencing of neurons (Schiavo *et al.*, 1992; Qi *et al.*, 2012; Williams *et al.*, 2013; Pokala *et al.*, 2014) and those required for monitoring neuronal activity (Nagai *et al.*, 2004; Chen *et al.*, 2013; Dana *et al.*, 2016). The targeted expression of exogenous genes in *Strongyloides* has been aided by the identification of several promoters that drive expression in single cells or subsets of cells (Junio *et al.*, 2008; Stoltzfus *et al.*, 2012; Bryant *et al.*, 2018). However, whereas extrachromosomal arrays in *C. elegans* are stably expressed across generations, extrachromosomal arrays in *Strongyloides* are silenced after the F₁ generation by as-yet-unknown mechanisms (Junio *et al.*, 2008; Li *et al.*, 2011). Persistent expression across generations can be achieved in *Strongyloides* by methods that promote genomic integration of transgenes, such as transposon-mediated

random integration (Shao *et al.*, 2012; Lok, 2013) and CRISPR/Cas9-mediated targeted integration (Gang *et al.*, 2017).

Methods for disrupting gene function are also now available for *Strongyloides stercoralis* and *Strongyloides ratti*. The recent development of an approach for CRISPR/Cas9-mediated targeted gene disruption in these species provided the first insights into the genetic mechanisms that drive sensory behaviours (Fig. 4) (Gang *et al.*, 2017; Lok *et al.*, 2017; Bryant *et al.*, 2018). For example, knockout of the gene encoding the cyclic-nucleotide-gated channel subunit TAX-4 severely disrupts the thermosensory behaviour of *Strongyloides stercoralis* infective larvae (Bryant *et al.*, 2018). RNA interference (RNAi) has also now been successfully applied to *Strongyloides ratti*. RNAi approaches using both dsRNA and siRNA have been used to study the effects of transcriptional knockdown of genes in several parasitic nematode species, although with varying efficacy (Geldhof *et al.*, 2006; Kotze and Bagnall, 2006; Visser *et al.*, 2006; Kang and Hong, 2008; Lendner *et al.*, 2008; Viney and Thompson, 2008; Samarasinghe *et al.*, 2011; Britton *et al.*, 2012; Zawadzki *et al.*, 2012; Tzelos, 2014). In the case of *Strongyloides ratti*, a recent study demonstrated the first successful knockdown of multiple mRNAs using an siRNA approach (Dulovic and Streit, 2019). In addition, chemical mutagenesis has been used to perform unbiased forward genetic screens to generate dominant non-targeted mutations in *Strongyloides ratti* iL3s, although mapping the locations of these mutations has not been possible yet (Viney *et al.*, 2002; Guo *et al.*, 2015).

Using a combination of the above approaches, it should be possible to identify the neural mechanisms and molecular pathways that are involved in driving behavioural and physiological responses of *Strongyloides stercoralis* to CO₂. For example, it will be interesting to determine whether the BAG neurons, which sense CO₂ and promote behavioural responses to CO₂ in *C. elegans*, play a similar role in *Strongyloides stercoralis*. It will also be important to elucidate the neural circuitry that operates downstream of the CO₂-sensing neurons to mediate or modulate CO₂-evoked behaviours in *Strongyloides stercoralis*. An intriguing possibility is that while sensory neuron function may be generally conserved across species, interneuron function may be less well conserved and may instead reflect species-specific behavioural and physiological responses to CO₂. In addition, through the systematic screening of candidate genes known to be involved in CO₂ responsiveness in *C. elegans*, it might be possible to uncover molecular signals that regulate parasite–host interactions or that are required for successful parasitism. In the long run, a better understanding of the molecular and cellular bases of CO₂-evoked behaviours in parasitic nematodes may lead to new avenues for nematode control. It may also shed light on some of the unique sensory mechanisms that operate in parasitic nematodes to shape parasite-specific behavioural responses.

Acknowledgments. We thank Astra Bryant, Michelle Castelletto, Elisa Rojas-Palato, Felicitas Ruiz and Breanna Walsh for their insightful comments on the manuscript.

Financial support. This work was supported by the National Institutes of Health (1DP2DC014596 and 1R01DC017959 to E.A.H., and 1F32AI147617 to N.B.). E.A.H. is also supported by a Burroughs Wellcome Fund Investigators in the Pathogenesis of Disease Award and a Howard Hughes Medical Institute Faculty Scholar Award.

References

Ali JG, Alborn HT and Stelinski LL (2010) Subterranean herbivore-induced volatiles released by citrus roots upon feeding by *Diaprepes abbreviatus* recruit entomopathogenic nematodes. *Journal of Chemical Ecology* **36**, 361–368.

- Alkalay I, Suetsugu S, Constantine H and Stein M (1971) Carbon dioxide elimination across human skin. *American Journal of Physiology* **220**, 1434–1436.
- Ashton FT, Bhopale VM, Fine AE and Schad GA (1995) Sensory neuroanatomy of a skin-penetrating nematode parasite: *Strongyloides stercoralis*. I. Amphidial neurons. *Journal of Comparative Neurology* **357**, 281–295.
- Ashton FT, Bhopale VM, Holt D, Smith G and Schad GA (1998) Developmental switching in the parasitic nematode *Strongyloides stercoralis* is controlled by the ASF and ASI amphidial neurons. *Journal of Parasitology* **84**, 691–695.
- Ashton FT, Li J and Schad GA (1999) Chemo- and thermosensory neurons: structure and function in animal parasitic nematodes. *Veterinary Parasitology* **84**, 297–316.
- Ashton FT, Zhu X, Boston R, Lok JB and Schad GA (2007) *Strongyloides stercoralis*: amphidial neuron pair ASJ triggers significant resumption of development by infective larvae under host-mimicking *in vitro* conditions. *Experimental Parasitology* **115**, 92–97.
- Baird SE (1999) Natural and experimental associations of *Caenorhabditis remanei* with *Trachelipus rathkii* and other terrestrial isopods. *Nematology* **1**, 471–475.
- Bedding RA and Molyneux AS (1982) Penetration of insect cuticle by infective juveniles of *Heterorhabditis* spp. (Heterorhabditidae: Nematoda). *Nematologica* **28**, 354–359.
- Bekelaar K, Waghorn T, Tavendale M, McKenzie C and Leathwick D (2018) Carbon dioxide is an absolute requirement for exsheathment of some, but not all, abomasal nematode species. *Parasitology Research* **117**, 3675–3678.
- Bekelaar K, Waghorn T, Tavendale M, McKenzie C and Leathwick D (2019) Abomasal nematode species differ in their *in vitro* response to exsheathment triggers. *Parasitology Research* **118**, 707–710.
- Bendesky A, Tsunozaki M, Rockman MV, Kruglyak L and Bargmann CI (2011) Catecholamine receptor polymorphisms affect decision-making in *C. elegans*. *Nature* **472**, 313–318.
- Bensafi M, Frasnelli J, Reden J and Hummel T (2007) The neural representation of odor is modulated by the presence of a trigeminal stimulus during odor encoding. *Clinical Neurophysiology* **118**, 696–701.
- Bernard GC, Egnin M and Bonsi C (2017) The impact of plant-parasitic nematodes on agriculture and methods of control. In Shah MM and Mahamood M (eds), *Nematology: Concepts, Control, and Diagnosis*. London, UK: IntechOpen, pp. 121–151. doi: 10.5772/intechopen.68958.
- Bezubik B (1965) Failure to establish infection in rats and guinea pigs exposed to the larvae of *Strongyloides papillosus*. *Acta Parasitologica* **13**, 349–354.
- Bhattacharya A, Aghayeva U, Berghoff EG and Hobert O (2019) Plasticity of the electrical connectome of *C. elegans*. *Cell* **176**, 1174–1189.
- Bhopale VM, Kupprion EK, Ashton FT, Boston R and Schad GA (2001) *Ancylostoma caninum*: the finger cell neurons mediate thermotactic behavior by infective larvae of the dog hookworm. *Experimental Parasitology* **97**, 70–76.
- Blaxter M and Koutsovoulos G (2015) The evolution of parasitism in Nematoda. *Parasitology* **142**(suppl. 1), S26–S39.
- Blaxter M, Koutsovoulos G, Jones M, Kumar S and Elsworth B (2016) Phylogenomics of Nematoda. In Olson PD, Hughes J and Cotton JA (eds), *Next Generation Systematics*. Cambridge, UK: Cambridge University Press, pp. 62–82.
- Boatin BA, Basanez MG, Prichard RK, Awadzi K, Barakat RM, Garcia HH, Gazzinelli A, Grant WN, McCarthy JS, N'Goran EK, Osei-Atweneboana MY, Sripa B, Yang FJ and Lustigman S (2012) A research agenda for helminth diseases of humans: towards control and elimination. *PLoS Neglected Tropical Diseases* **6**, e1547.
- Brandt JP and Ringstad N (2015) Toll-like receptor signaling promotes development and function of sensory neurons required for a *C. elegans* pathogen-avoidance behavior. *Current Biology* **25**, 2228–2237.
- Brandt JP, Aziz-Zaman S, Juozaityte V, Martinez-Velazquez LA, Petersen JG, Pockock R and Ringstad N (2012) A single gene target of an ETS-family transcription factor determines neuronal CO₂-chemosensitivity. *PLoS ONE* **7**, e34014.
- Bretscher AJ, Busch KE and de Bono M (2008) A carbon dioxide avoidance behavior is integrated with responses to ambient oxygen and food in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **105**, 8044–8049.
- Bretscher AJ, Kodama-Namba E, Busch KE, Murphy RJ, Soltesz Z, Laurent P and de Bono M (2011) Temperature, oxygen, and salt-sensing

- neurons in *C. elegans* are carbon dioxide sensors that control avoidance behavior. *Neuron* **69**, 1099–1113.
- Brindley PJ, Mitreva M, Ghedin E and Lustigman S (2009) Helminth genomics: the implications for human health. *PLoS Neglected Tropical Diseases* **3**, e538.
- Britton C, Samarasinghe B and Knox DP (2012) Ups and downs of RNA interference in parasitic nematodes. *Experimental Parasitology* **132**, 56–61.
- Bryant AS and Hallem EA (2018) Terror in the dirt: sensory determinants of host seeking in soil-transmitted mammalian-parasitic nematodes. *International Journal for Parasitology: Drugs and Drug Resistance* **8**, 496–510.
- Bryant AS, Ruiz F, Gang SS, Castelletto ML, Lopez JB and Hallem EA (2018) A critical role for thermosensation in host seeking by skin-penetrating nematodes. *Current Biology* **28**, 2338–2347.
- Bumbarger DJ, Crum J, Ellisman MH and Baldwin JG (2007) Three-dimensional fine structural reconstruction of the nose sensory structures of *Acrobelles complexus* compared to *Caenorhabditis elegans* (Nematoda: rhabditida). *Journal of Morphology* **268**, 649–663.
- Bumbarger DJ, Wijeratne S, Carter C, Crum J, Ellisman MH and Baldwin JG (2009) Three-dimensional reconstruction of the amphid sensilla in the microbial feeding nematode, *Acrobelles complexus* (Nematoda: rhabditida). *Journal of Comparative Neurology* **512**, 271–281.
- Burg SP and Burg EA (1965) Gas exchange in fruits. *Physiologia Plantarum* **18**, 870–884.
- Buszewski B, Keszy M, Ligor T and Amann A (2007) Human exhaled air analytics: biomarkers of diseases. *Biomedical Chromatography* **21**, 553–566.
- Byrnes CA, Dinarevic S, Shinebourne EA, Barnes PJ and Bush A (1997) Exhaled nitric oxide measurements in normal and asthmatic children. *Pediatric Pulmonology* **24**, 312–318.
- Campbell JF and Gauger R (1993) Nictation behaviour and its ecological implications in the host search strategies of entomopathogenic nematodes (Heterorhabditidae and Steinernematidae). *Behaviour* **126**, 155–169.
- Campbell JF and Kaya HK (1999) How and why a parasitic nematode jumps. *Nature* **397**, 485–486.
- Campbell JF and Kaya HK (2000) Influence of insect-associated cues on the jumping behavior of entomopathogenic nematodes (*Steinernema* spp.). *Behaviour* **137**, 591–609.
- Carrillo MA and Hallem EA (2015) Gas sensing in nematodes. *Molecular Neurobiology* **51**, 919–931.
- Carrillo MA, Guillermin ML, Rengarajan S, Okubo R and Hallem EA (2013) O₂-sensing neurons control CO₂ response in *C. elegans*. *Journal of Neuroscience* **33**, 9675–9683.
- Castelletto ML, Gang SS, Okubo RP, Tselikova AA, Nolan TJ, Platzer EG, Lok JB and Hallem EA (2014) Diverse host-seeking behaviors of skin-penetrating nematodes. *PLoS Pathogens* **10**, e1004305.
- Caswell-Chen EP, Chen J, Lewis EE, Douhan GW, Nadler SA and Carey JR (2005) Revising the standard wisdom of *C. elegans* natural history: ecology of longevity. *Science of Aging Knowledge Environment* **2005**, pe30.
- Chang DZ, Serra L, Lu D, Mortazavi A and Dillman AR (2019) A core set of venom proteins is released by entomopathogenic nematodes in the genus *Steinernema*. *PLoS Pathogens* **15**, e1007626.
- Chen TW, Wardill TJ, Sun Y, Pulver SR, Renninger SL, Baohan A, Schreiter ER, Kerr RA, Orger MB, Jayaraman V, Looger LL, Svoboda K and Kim DS (2013) Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**, 295–300.
- Crook M (2014) The dauer hypothesis and the evolution of parasitism: 20 years on and still going strong. *International Journal for Parasitology* **44**, 1–8.
- Dana H, Mohar B, Sun Y, Narayan S, Gordus A, Hasseman JP, Tsegaye G, Holt GT, Hu A, Walpita D, Patel R, Macklin JJ, Bargmann CI, Ahrens MB, Schreiter ER, Jayaraman V, Looger LL, Svoboda K and Kim DS (2016) Sensitive red protein calcium indicators for imaging neural activity. *eLife* **5**: e12727. doi: 10.7554/eLife.12727.
- Decraemer W and Hunt D (2006) Structure and classification. In Perry R and Moens M (eds), *Plant Nematology*. Oxfordshire: CAB International, pp. 3–32.
- de Lacy Costello B, Amann A, Al-Kateb H, Flynn C, Filipiak W, Khalid T, Osborne D and Ratcliffe NM (2014) A review of the volatiles from the healthy human body. *Journal of Breath Research* **8**, 014001.
- Diawara A, Schwenkenbecher JM, Kaplan RM and Prichard RK (2013) Molecular and biological diagnostic tests for monitoring benzimidazole resistance in human soil-transmitted helminths. *American Journal of Tropical Medicine and Hygiene* **88**, 1052–1061.
- Dillman AR and Sternberg PW (2012) Entomopathogenic nematodes. *Current Biology* **22**, R430–R431.
- Dillman AR, Chaston JM, Adams BJ, Ciche TA, Goodrich-Blair H, Stock SP and Sternberg PW (2012a) An entomopathogenic nematode by any other name. *PLoS Pathogens* **8**, e1002527.
- Dillman AR, Guillermin ML, Lee JH, Kim B, Sternberg PW and Hallem EA (2012b) Olfaction shapes host-parasite interactions in parasitic nematodes. *Proceedings of the National Academy of Sciences USA* **109**, E2324–E2333.
- Dold C and Holland CV (2011) *Ascaris* and ascariasis. *Microbes and Infections* **13**, 632–637.
- Douvres FW and Urban JF (1983) Factors contributing to the *in vitro* development of *Ascaris suum* from second-stage larvae to mature adults. *Journal of Parasitology* **69**, 549–558.
- Drexler T, Mahofsky K, Latham R, Zimmer M and Cochella L (2016) Neuron type-specific miRNA represses two broadly expressed genes to modulate an avoidance behavior in *C. elegans*. *Genes and Development* **30**, 2042–2047.
- Dulovic A and Streit A (2019) RNAi-mediated knockdown of *daf-12* in the model parasitic nematode *Strongyloides ratti*. *PLoS Pathogens* **15**, e1007705.
- Dusenbery DB (1985) Video camera-computer tracking of nematode *Caenorhabditis elegans* to record behavioral responses. *Journal of Chemical Ecology* **11**, 1239–1247.
- Emery DL, Hunt PW and Le Jambre LF (2016) *Haemonchus contortus*: the then and now, and where to from here? *International Journal for Parasitology* **46**, 755–769.
- Evans TC (2006) Transformation and microinjection. In The *C. elegans* Research Community (eds). *WormBook*, pp. 1–15. <http://www.wormbook.org>.
- Fairbairn D (1961) The *in vitro* hatching of *Ascaris lumbricoides* eggs. *Canadian Journal of Zoology* **39**, 153–162.
- Felix MA and Braendle C (2010) The natural history of *Caenorhabditis elegans*. *Current Biology* **20**, R965–R969.
- Felix MA and Duvéau F (2012) Population dynamics and habitat sharing of natural populations of *Caenorhabditis elegans* and *C. briggsae*. *BMC Biology* **10**, 59.
- Fenk LA and de Bono M (2015) Environmental CO₂ inhibits *Caenorhabditis elegans* egg-laying by modulating olfactory neurons and evokes widespread changes in neural activity. *Proceedings of the National Academy of Sciences USA* **112**, E3525–E3534.
- Fenk LA and de Bono M (2017) Memory of recent oxygen experience switches pheromone valence in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **114**, 4195–4200.
- Forbes WM, Ashton FT, Boston R, Zhu X and Schad GA (2004) Chemoattraction and chemorepulsion of *Strongyloides stercoralis* infective larvae on a sodium chloride gradient is mediated by amphidial neuron pairs ASE and ASH, respectively. *Veterinary Parasitology* **120**, 189–198.
- Gang SS and Hallem EA (2016) Mechanisms of host seeking by parasitic nematodes. *Molecular and Biochemical Parasitology* **208**, 23–32.
- Gang SS, Castelletto ML, Bryant AS, Yang E, Mancuso N, Lopez JB, Pellegrini M and Hallem EA (2017) Targeted mutagenesis in a human-parasitic nematode. *PLoS Pathogens* **13**, e1006675.
- Gaugler R, LeBeck L, Nakagaki B and Boush GM (1980) Orientation of the entomogenous nematode *Neoplectana carpocapsae* to carbon dioxide. *Environmental Entomology* **9**, 649–652.
- Gaugler R, Campbell JF and Gupta P (1991) Characterization and basis of enhanced host-finding in a genetically improved strain of *Steinernema carpocapsae*. *Journal of Invertebrate Pathology* **57**, 234–241.
- Geldhof P, Murray L, Couthier A, Gilleard JS, McLauchlan G, Knox DP and Britton C (2006) Testing the efficacy of RNA interference in *Haemonchus contortus*. *International Journal for Parasitology* **36**, 801–810.
- Granzer M and Haas W (1991) Host-finding and host recognition of infective *Ancylostoma caninum* larvae. *International Journal for Parasitology* **21**, 429–440.
- Gray JM, Karow DS, Lu H, Chang AJ, Chang JS, Ellis RE, Marletta MA and Bargmann CI (2004) Oxygen sensation and social feeding mediated by a *C. elegans* guanylate cyclase homologue. *Nature* **430**, 317–322.
- Grewal PS, Ehlers R-U and Shapiro-Ilan DI (2005) *Nematodes as Biocontrol Agents*. Cambridge: CABI Publishing.
- Guillermin ML, Castelletto ML and Hallem EA (2011) Differentiation of carbon dioxide-sensing neurons in *Caenorhabditis elegans* requires the ETS-5 transcription factor. *Genetics* **189**, 1327–1339.
- Guillermin ML, Carrillo MA and Hallem EA (2017) A single set of interneurons drives opposite behaviors in *C. elegans*. *Current Biology* **27**, 2630–2639.

- Guo L, Chang Z, Dieterich C and Streit A (2015) A protocol for chemical mutagenesis in *Strongyloides ratti*. *Experimental Parasitology* **158**, 2–7.
- Haas W, Haberl B, Idris SI, Kallert D, Kersten S and Stiegeler P (2005) Behavioural strategies used by the hookworms *Necator americanus* and *Ancylostoma duodenale* to find, recognize and invade the human host. *Parasitology Research* **95**, 30–39.
- Haley AJ (1961) Biology of the rat nematode *Nippostrongylus brasiliensis* (Travassos, 1914). I. Systematics, hosts and geographic distribution. *Journal of Parasitology* **47**, 727–732.
- Hallem EA and Sternberg PW (2008) Acute carbon dioxide avoidance in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **105**, 8038–8043.
- Hallem EA, Dillman AR, Hong AV, Zhang Y, Yano JM, DeMarco SF and Sternberg PW (2011a) A sensory code for host seeking in parasitic nematodes. *Current Biology* **21**, 377–383.
- Hallem EA, Spencer WC, McWhirter RD, Zeller G, Henz SR, Ratsch G, Miller DM, Horvitz HR, Sternberg PW and Ringstad N (2011b) Receptor-type guanylate cyclase is required for carbon dioxide sensation by *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **108**, 254–259.
- Hansen EL and Buecher EJ (1970) Biochemical approach to systematic studies with axenic nematodes. *Journal of Nematology* **2**, 1–6.
- Harris G, Shen Y, Ha H, Donato A, Wallis S, Zhang X and Zhang Y (2014) Dissecting the signaling mechanisms underlying recognition and preference of food odors. *Journal of Neuroscience* **34**, 9389–9403.
- Hawdon JM and Schad GA (1990) Serum-stimulated feeding *in vitro* by third-stage infective larvae of the canine hookworm *Ancylostoma caninum*. *Journal of Parasitology* **76**, 394–398.
- Hawdon JM and Schad GA (1992) *Ancylostoma caninum*: reduced glutathione stimulates feeding by third-stage infective larvae. *Experimental Parasitology* **75**, 40–46.
- Hawdon JM, Volk SW, Pritchard DI and Schad GA (1992) Resumption of feeding *in vitro* by hookworm third-stage larvae: a comparative study. *Journal of Parasitology* **78**, 1036–1040.
- Hernandez AD and Sukhdeo MVK (1995) Host grooming and the transmission strategy of *Heligmosomoides polygyrus*. *Journal of Parasitology* **81**, 865–869.
- Hominick WM (2002) Biogeography. In Gaugler R (ed.), *Entomopathogenic Nematology*. New York: CABI Publishing, pp. 115–143.
- Hotez P, Hawdon J and Schad GA (1993) Hookworm larval infectivity, arrest and amphiparatensis: the *Caenorhabditis elegans* Daf-c paradigm. *Parasitology Today* **9**, 23–26.
- Howe KL, Bolt BJ, Shafie M, Kersey P and Berriman M (2017) Wormbase ParaSite – a comprehensive resource for helminth genomics. *Molecular and Biochemical Parasitology* **215**, 2–10.
- Hu PJ (2007) Dauer. In The *C. elegans* Research Community (eds). *WormBook*, pp. 1–19. <http://www.wormbook.org>.
- Hunt VL, Tsai IJ, Coghlan A, Reid AJ, Holroyd N, Foth BJ, Tracey A, Cotton JA, Stanley EJ, Beasley H, Bennett HM, Brooks K, Harsha B, Kajitani R, Kulkarni A, Harbecke D, Nagayasu E, Nichol S, Ogura Y, Quail MA, Randle N, Xia D, Brattig NW, Soblik H, Ribeiro DM, Sanchez-Flores A, Hayashi T, Itoh T, Denver DR, Grant W, Stoltzfus JD, Lok JB, Murayama H, Wastling J, Streit A, Kikuchi T, Viney M and Berriman M (2016) The genomic basis of parasitism in the *Strongyloides* clade of nematodes. *Nature Genetics* **48**, 299–307.
- International Helminth Genomes C (2019) Comparative genomics of the major parasitic worms. *Nature Genetics* **51**, 163–174.
- Jasmer DP, Govere A and Smant G (2003) Parasitic nematode interactions with mammals and plants. *Annual Review of Phytopathology* **41**, 245–270.
- Jensen BB and Jorgensen H (1994) Effect of dietary fiber on microbial activity and microbial gas production in various regions of the gastrointestinal tract of pigs. *Applied and Environmental Microbiology* **60**, 1897–1904.
- Jex AR, Gasser RB and Schwarz EM (2019) Transcriptomic resources for parasitic nematodes of veterinary importance. *Trends in Parasitology* **35**, 72–84.
- Johnson RN and Viglierchio DR (1961) The accumulation of plant parasitic nematode larvae around carbon dioxide and oxygen. *Proceedings of the Helminthological Society of Washington* **28**, 171–174.
- Junio AB, Li X, Massey Jr HC, Nolan TJ, Todd Lamitina S, Sundaram MV and Lok JB (2008) *Strongyloides stercoralis*: cell- and tissue-specific transgene expression and co-transformation with vector constructs incorporating a common multifunctional 3' UTR. *Experimental Parasitology* **118**, 253–265.
- Juozaityte V, Pladevall-Morera D, Podolska A, Norgaard S, Neumann B and Pocock R (2017) The ETS-5 transcription factor regulates activity states in *Caenorhabditis elegans* by controlling satiety. *Proceedings of the National Academy of Sciences USA* **114**, E1651–E1658.
- Kang S and Hong YS (2008) RNA interference in infectious tropical diseases. *Korean Journal of Parasitology* **46**, 1–15.
- Kaya HK and Gaugler R (1993) Entomopathogenic nematodes. *Annual Review of Entomology* **38**, 181–206.
- Keiser J and Utzinger J (2008) Efficacy of current drugs against soil-transmitted helminth infections: systematic review and meta-analysis. *Journal of the American Medical Association* **299**, 1937–1948.
- Ketschek AR, Joseph R, Boston R, Ashton FT and Schad GA (2004) Amphidial neurons ADL and ASH initiate sodium dodecyl sulphate avoidance responses in the infective larva of the dog hookworm *Ancylostoma caninum*. *International Journal for Parasitology* **34**, 1333–1336.
- Klingler J (1972) The effect of single and combined heat and CO₂ stimuli at different ambient temperatures on the behavior of two plant-parasitic nematodes. *Journal of Nematology* **4**, 95–100.
- Kodama-Namba E, Fenk LA, Bretscher AJ, Gross E, Busch KE and de Bono M (2013) Cross-modulation of homeostatic responses to temperature, oxygen and carbon dioxide in *C. elegans*. *PLoS Genetics* **9**, e1004011.
- Kotze AC and Bagnall NH (2006) RNA interference in *Haemonchus contortus*: suppression of b-tubulin gene expression in L3, L4 and adult worms *in vitro*. *Molecular and Biochemical Parasitology* **145**, 101–110.
- Kumar N, Rao TK, Varghese A and Rathor VS (2013) Internal parasite management in grazing livestock. *Journal of Parasitic Diseases* **37**, 151–157.
- Labaude S and Griffin CT (2018) Transmission success of entomopathogenic nematodes used in pest control. *Insects* **9**, 72.
- Lahiri S and Forster II RE (2003) CO₂/H⁺ sensing: peripheral and central chemoreception. *International Journal of Biochemistry and Cell Biology* **35**, 1413–1435.
- Laing R, Kikuchi T, Martinelli A, Tsai IJ, Beech RN, Redman E, Holroyd N, Bartley DJ, Beasley H, Britton C, Curran D, Devaney E, Gilabert A, Hunt M, Jackson F, Johnston SL, Kryukov I, Li K, Morrison AA, Reid AJ, Sargison N, Saunders GI, Wasmuth JD, Wolstenholme A, Berriman M, Gilleard JS and Cotton JA (2013) The genome and transcriptome of *Haemonchus contortus*, a key model parasite for drug and vaccine discovery. *Genome Biology* **14**, R88.
- Learmount J, Stephens N, Boughtflower V, Barrecheguren A and Rickell K (2016) The development of anthelmintic resistance with best practice control of nematodes on commercial sheep farms in the UK. *Veterinary Parasitology* **229**, 9–14.
- Lee H, Choi MK, Lee D, Kim HS, Hwang H, Kim H, Park S, Paik YK and Lee J (2012) Nictation, a dispersal behavior of the nematode *Caenorhabditis elegans*, is regulated by IL2 neurons. *Nature Neuroscience* **15**, 107–112.
- Lee J, Dillman AR and Hallem EA (2016) Temperature-dependent changes in the host-seeking behaviors of parasitic nematodes. *BMC Biology* **14**, 36.
- Lee JS, Shih PY, Schaedel ON, Quintero-Cadena P, Rogers AK and Sternberg PW (2017) FMRFamide-like peptides expand the behavioral repertoire of a densely connected nervous system. *Proceedings of the National Academy of Sciences USA* **114**, E10726–E10735.
- Lendner M, Doligalska M, Lucius R and Hartmann S (2008) Attempts to establish RNA interference in the parasitic nematode *Heligmosomoides polygyrus*. *Molecular and Biochemical Parasitology* **161**, 21–31.
- Lewis EE (2002) Behavioral ecology. In Gaugler R (ed.), *Entomopathogenic Nematology*. New York: CAB International, pp. 205–223.
- Lewis EE, Gauger R and Harrison R (1993) Response of cruiser and ambusher entomopathogenic nematodes (steinernematidae) to host volatile cues. *Canadian Journal of Zoology* **71**, 765–769.
- Lewis EE, Campbell J, Griffin C, Kaya H and Peters A (2006) Behavioral ecology of entomopathogenic nematodes. *Biological Control* **38**, 66–79.
- Li J, Ashton FT, Gamble HR and Schad GA (2000a) Sensory neuroanatomy of a passively ingested nematode parasite, *Haemonchus contortus*: amphidial neurons of the first stage larva. *Journal of Comparative Neurology* **417**, 299–314.
- Li J, Zhu X, Boston R, Ashton FT, Gamble HR and Schad GA (2000b) Thermotaxis and thermosensory neurons in infective larvae of *Haemonchus contortus*, a passively ingested nematode parasite. *Journal of Comparative Neurology* **424**, 58–73.
- Li J, Zhu X, Ashton FT, Gamble HR and Schad GA (2001) Sensory neuroanatomy of a passively ingested nematode parasite, *Haemonchus contortus*: amphidial neurons of the third-stage larva. *Journal of Parasitology* **87**, 65–72.

- Li X, Massey Jr HC, Nolan TJ, Schad GA, Kraus K, Sundaram M and Lok JB (2006) Successful transgenesis of the parasitic nematode *Strongyloides stercoralis* requires endogenous non-coding control elements. *International Journal for Parasitology* **36**, 671–679.
- Li X, Shao H, Junio A, Nolan TJ, Massey Jr HC, Pearce EJ, Viney ME and Lok JB (2011) Transgenesis in the parasitic nematode *Strongyloides ratti*. *Molecular and Biochemical Parasitology* **179**, 114–119.
- Liu T and Cai D (2013) Counterbalance between BAG and URX neurons via guanylate cyclases controls lifespan homeostasis in *C. elegans*. *EMBO Journal* **32**, 1529–1542.
- Liu J, Poinar Jr GO and Berry RE (2000) Control of insect pests with entomopathogenic nematodes: the impact of molecular biology and phylogenetic reconstruction. *Annual Review of Entomology* **45**, 287–306.
- Lok JB (2007) *Strongyloides stercoralis*: a model for translational research on parasitic nematode biology. In *The C. elegans Research Community* (eds). *WormBook*, pp. 1–18. <http://www.wormbook.org>.
- Lok J (2013) Piggybac: a vehicle for integrative DNA transformation of parasitic nematodes. *Mobile Genetic Elements* **3**, e24417.
- Lok JB (2012) Nucleic acid transfection and transgenesis in parasitic nematodes. *Parasitology* **139**, 574–588.
- Lok JB and Artis D (2008) Transgenesis and neuronal ablation in parasitic nematodes: revolutionary new tools to dissect host-parasite interactions. *Parasite Immunology* **30**, 203–214.
- Lok JB and Massey Jr HC (2002) Transgene expression in *Strongyloides stercoralis* following gonadal microinjection of DNA constructs. *Molecular and Biochemical Parasitology* **119**, 279–284.
- Lok JB, Shao H, Massey HC and Li X (2017) Transgenesis in *Strongyloides* and related parasitic nematodes: historical perspectives, current functional genomic applications and progress towards gene disruption and editing. *Parasitology* **144**, 327–342.
- Lopez PM, Boston R, Ashton FT and Schad GA (2000) The neurons of class ALD mediate thermotaxis in the parasitic nematode, *Strongyloides stercoralis*. *International Journal for Parasitology* **30**, 1115–1121.
- Lu D, Macchietto M, Chang D, Barros MM, Baldwin J, Mortazavi A and Dillman AR (2017) Activated entomopathogenic nematode infective juveniles release lethal venom proteins. *PLoS Pathogens* **13**, e1006302.
- Lustigman S, Prichard RK, Gazzinelli A, Grant WN, Boatman BA, McCarthy JS and Basanez MG (2012) A research agenda for helminth diseases of humans: the problem of helminthiasis. *PLoS Neglected Tropical Diseases* **6**, e1582.
- McCallum ME and Dusenbery DB (1992) Computer tracking as a behavioral GC detector: nematode responses to vapor of host roots. *Journal of Chemical Ecology* **18**, 585–592.
- McGrath PT, Rockman MV, Zimmer M, Jang H, Macosko EZ, Kruglyak L and Bargmann CI (2009) Quantitative mapping of a digenic behavioral trait implicates globin variation in *C. elegans* sensory behaviors. *Neuron* **61**, 692–699.
- Milward K, Busch KE, Murphy RJ, de Bono M and Olofsson B (2011) Neuronal and molecular substrates for optimal foraging in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **108**, 20672–20677.
- Mitreva M, Zarlenga DS, McCarter JP and Jasmer DP (2007) Parasitic nematodes – from genomes to control. *Veterinary Parasitology* **148**, 31–42.
- Nagai T, Yamada S, Tominaga T, Ichikawa M and Miyawaki A (2004) Expanded dynamic range of fluorescent indicators for Ca²⁺ by circularly permuted yellow fluorescent proteins. *Proceedings of the National Academy of Sciences USA* **101**, 10554–10559.
- Nguyen KB and Hunt DJ eds (2007) *Entomopathogenic Nematodes: Systematics, Phylogeny and Bacterial Symbionts*. Leiden, The Netherlands: Brill.
- Nguyen KB and Smart GC (1991) Pathogenicity of *Steinernema scapterisci* to selected invertebrate hosts. *Journal of Nematology* **23**, 7–11.
- Nolan TJ, Brenes M, Ashton FT, Zhu X, Forbes WM, Boston R and Schad GA (2004) The amphidial neuron pair ALD controls the temperature-sensitive choice of alternative developmental pathways in the parasitic nematode, *Strongyloides stercoralis*. *Parasitology* **129**, 753–759.
- Nolan TJ, Zhu X, Ketschek A, Cole J, Grant W, Lok JB and Schad GA (2007) The sugar glider (*Petaurus breviceps*): a laboratory host for the nematode *Parastrongyloides trichosuri*. *Journal of Parasitology* **93**, 1084–1089.
- Nutman TB (2017) Human infection with *Strongyloides stercoralis* and other related *Strongyloides* species. *Parasitology* **144**, 263–273.
- O'Connor LJ, Walkden-Brown SW and Kahn LP (2006) Ecology of the free-living stages of major trichostrongylid parasites of sheep. *Veterinary Parasitology* **142**, 1–15.
- O'Halloran DM and Burnell AM (2003) An investigation of chemotaxis in the insect parasitic nematode *Heterorhabditis bacteriophora*. *Parasitology* **127**, 375–385.
- Page W, Judd JA and Bradbury RS (2018) The unique life cycle of *Strongyloides stercoralis* and implications for public health action. *Tropical Medicine and Infectious Disease* **3**, 53.
- Peters A (1996) The natural host range of *Steinernema* and *Heterorhabditis* spp. and their impact on insect populations. *Biocontrol Science and Technology* **6**, 389–402.
- Petersen JG, Romanos TR, Juozaityte V, Riveiro AR, Hum I, Traunmuller L, Zimmer M and Pocock R (2013) EGL-13/SoxD specifies distinct O₂ and CO₂ sensory neuron fates in *Caenorhabditis elegans*. *PLoS Genetics* **9**, e1003511.
- Pilz M and Hohberg K (2015) Laboratory experiments on nematodes from natural CO₂ springs reveal species-specific tolerance to extreme CO₂ concentrations. *Soil Biology and Biochemistry* **88**, 165–168.
- Pokala N, Liu Q, Gordus A and Bargmann CI (2014) Inducible and titratable silencing of *Caenorhabditis elegans* neurons *in vivo* with histamine-gated chloride channels. *Proceedings of the National Academy of Sciences USA* **111**, 2770–2775.
- Prichard RK, Basanez MG, Boatman BA, McCarthy JS, Garcia HH, Yang GJ, Sripa B and Lustigman S (2012) A research agenda for helminth diseases of humans: intervention for control and elimination. *PLoS Neglected Tropical Diseases* **6**, e1549.
- Prot JC (1980) Migration of plant-parasitic nematodes towards plant roots. *Revue de Nematology* **3**, 305–318.
- Pye AE and Burman M (1981) *Neoaplectana carpocapsae*: nematode accumulations on chemical and bacterial gradients. *Experimental Parasitology* **51**, 13–20.
- Qi YB, Garren EJ, Shu X, Tsien RY and Jin Y (2012) Photo-inducible cell ablation in *Caenorhabditis elegans* using the genetically encoded singlet oxygen generating protein miniSOG. *Proceedings of the National Academy of Sciences of the USA* **109**, 7499–7504.
- Rengarajan S, Yankura KA, Guillermin ML, Fung W and Hallem EA (2019) Feeding state sculpts a circuit for sensory valence in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the USA* **116**, 1776–1781.
- Repetto SA, Ruybal P, Batalla E, Lopez C, Fridman V, Sierra M, Radisic M, Bravo PM, Risso MG, Gonzalez Cappa SM and Alba Soto CD (2018) Strongyloidiasis outside endemic areas: long-term parasitological and clinical follow-up after ivermectin treatment. *Journal of Biochemistry and Molecular Biology* **66**, 1558–1565.
- Riemann F and Schrage M (1988) Carbon dioxide as an attractant for the free-living marine nematode *Adoncholaimus thalassophygas*. *Marine Biology* **98**, 81–85.
- Roberts LS, Janovy J and Schmidt P (2005) *Foundations of Parasitology*, 7th Edn. New York City, USA: McGraw-Hill.
- Robinson AF (1995) Optimal release rates for attracting *Meloidogyne incognita*, *Rotylenchulus reniformis*, and other nematodes to carbon dioxide in sand. *Journal of Nematology* **27**, 42–50.
- Roeber F, Jex AR and Gasser RB (2013) Impact of gastrointestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance – an Australian perspective. *Parasites & Vectors* **6**, 153.
- Rogers WP and Sommerville RI (1960) The physiology of the second ecdysis of parasitic nematodes. *Parasitology* **50**, 329–348.
- Rojo Romanos T, Petersen JG and Pocock R (2017) Control of neuropeptide expression by parallel activity-dependent pathways in *Caenorhabditis elegans*. *Scientific Reports* **7**, 38734.
- Rotbart A, Yao CK, Ha N, Chrispa MD, Muir JG, Gibson PR, Kalantar-Zadeh K and Ou JZ (2017) Designing an *in-vitro* gas profiling system for human faecal samples. *Sensors and Actuators. B, Chemical* **238**, 754–764.
- Ruiz F, Castelletto ML, Gang SS and Hallem EA (2017) Experience-dependent olfactory behaviors of the parasitic nematode *Heligmosomoides polygyrus*. *PLoS Pathogens* **13**, e1006709.
- Samarasinghe B, Knox DP and Britton C (2011) Factors affecting susceptibility to RNA interference in *Haemonchus contortus* and *in vivo* silencing of an H11 aminopeptidase gene. *International Journal for Parasitology* **41**, 51–59.

- Schiavo G, Benfenati F, Poulain B, Rossetto O, Polverino de Laureto P, DasGupta BR and Montecucco C (1992) Tetanus and botulinum-B neurotoxins block neurotransmitter release by proteolytic cleavage of synaptobrevin. *Nature* **359**, 832–835.
- Schmidt J and All JN (1979) Attraction of *Neoaplectana carpocapsae* (Nematoda: Steinernematidae) to common excretory products of insects. *Environmental Entomology* **8**, 55–61.
- Schulenburg H and Felix MA (2017) The natural biotic environment of *Caenorhabditis elegans*. *Genetics* **206**, 55–86.
- Sciaccia J, Forbes WM, Ashton FT, Lombardini E, Gamble HR and Schad GA (2002) Response to carbon dioxide by the infective larvae of three species of parasitic nematodes. *Parasitology International* **51**, 53–62.
- Scott K (2011) Out of thin air: sensory detection of oxygen and carbon dioxide. *Neuron* **69**, 194–202.
- Serrano-Saiz E, Poole RJ, Felton T, Zhang F, De La Cruz ED and Hobert O (2013) Modular control of glutamatergic neuronal identity in *C. elegans* by distinct homeodomain proteins. *Cell* **155**, 659–673.
- Shao H, Li X, Nolan TJ, Massey Jr HC, Pearce EJ and Lok JB (2012) Transposon-mediated chromosomal integration of transgenes in the parasitic nematode *Strongyloides ratti* and establishment of stable transgenic lines. *PLoS Pathogens* **8**, e1002871.
- Shao HG, Li XS and Lok JB (2017) Heritable genetic transformation of *Strongyloides stercoralis* by microinjection of plasmid DNA constructs into the male germline. *International Journal for Parasitology* **47**, 511–515.
- Sharabi K, Hurwitz A, Simon AJ, Beitel GJ, Morimoto RI, Rechavi G, Sznajder JI and Gruenbaum Y (2009) Elevated CO₂ levels affect development, motility, and fertility and extend life span in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences USA* **106**, 4024–4029.
- Shusterman D and Avila PC (2003) Real-time monitoring of nasal mucosal pH during carbon dioxide stimulation: implications for stimulus dynamics. *Chemical Senses* **28**, 595–601.
- Smallegange RC, Verhulst NO and Takken W (2011) Sweaty skin: an invitation to bite? *Trends in Parasitology* **27**, 143–148.
- Smith ES, Martinez-Velazquez L and Ringstad N (2013) A chemoreceptor that detects molecular carbon dioxide. *Journal of Biological Chemistry* **288**, 37071–37081.
- Sommerville RI (1964) Effect of carbon dioxide on the development of third-stage larvae of *Haemonchus contortus* *in vitro*. *Nature* **202**, 316–317.
- Srinivasan J, Durak O and Sternberg PW (2008) Evolution of a polymodal sensory response network. *BMC Biology* **6**, 52.
- Stoltzfus JD, Massey Jr HC, Nolan TJ, Griffith SD and Lok JB (2012) *Strongyloides stercoralis* age-1: a potential regulator of infective larval development in a parasitic nematode. *PLoS ONE* **7**, e38587.
- Stoltzfus JD, Bart SM and Lok JB (2014) cGMP and NHR signaling co-regulate expression of insulin-like peptides and developmental activation of infective larvae in *Strongyloides stercoralis*. *PLoS Pathogens* **10**, e1004235.
- Taylor KA and Weinstein PP (1990) The effect of oxygen and carbon dioxide on the development of the free-living stages of *Strongyloides ratti* in axenic culture. *Journal of Parasitology* **76**, 545–551.
- Taylor A and Whitlock JH (1960) The exsheathing stimulus for infective larvae of *Haemonchus contortus*. *Cornell Veterinarian* **50**, 339–344.
- Turlings TC, Hiltbold I and Rasmann S (2012) The importance of root-produced volatiles as foraging cues for entomopathogenic nematodes. *Plant and Soil* **358**, 51–60.
- Tzelos T (2014) *RNA interference in parasitic nematodes – from genome to control* (PhD thesis). University of Edinburgh, Edinburgh, UK.
- Vadasz I, Dada LA, Briva A, Helenius IT, Sharabi K, Welch LC, Kelly AM, Grzesik BA, Budinger GR, Liu J, Seeger W, Beitel GJ, Gruenbaum Y and Sznajder JI (2012) Evolutionary conserved role of c-Jun-N-terminal kinase in CO₂-induced epithelial dysfunction. *PLoS ONE* **7**, e46696.
- Velikkakam T, Fiuza JA and Gaze ST (2017) Overview of hookworm infection in humans. In Singh SK (ed.), *Neglected Tropical Diseases – South Asia*. Cham, Switzerland: Springer International Publishing, pp. 121–135.
- Viglierchio DR (1990) Carbon dioxide sensing by *Panagrellus silusiae* and *Ditylenchus dipsaci*. *Revue de Nématologie* **13**, 425–432.
- Viney ME (1999) Exploiting the life cycle of *Strongyloides ratti*. *Parasitology Today* **15**, 231–235.
- Viney ME (2006) The biology and genomics of *Strongyloides*. *Medical Microbiology and Immunology* **195**, 49–54.
- Viney M and Kikuchi T (2017) *Strongyloides ratti* and *S. venezuelensis* – rodent models of *Strongyloides* infection. *Parasitology* **144**, 285–294.
- Viney ME and Lok JB (2007) *Strongyloides* spp. In The *C. elegans* Research Community (eds). *WormBook*, pp. 1–15. <http://www.wormbook.org>.
- Viney ME and Thompson FJ (2008) Two hypotheses to explain why RNA interference does not work in animal parasitic nematodes. *International Journal for Parasitology* **38**, 43–47.
- Viney ME, Green LD, Brooks JA and Grant WN (2002) Chemical mutagenesis of the parasitic nematode *Strongyloides ratti* to isolate ivermectin resistant mutants. *International Journal for Parasitology* **32**, 1677–1682.
- Viney ME, Thompson FJ and Crook M (2005) TGF- β and the evolution of nematode parasitism. *International Journal for Parasitology* **35**, 1473–1475.
- Visser A, Geldhof P, de Maere V, Knox DP, Vercruyse J and Claerebout E (2006) Efficacy and specificity of RNA interference in larval life-stages of *Ostertagia ostertagi*. *Parasitology* **133**, 777–783.
- Wang C, Bruening G and Williamson VM (2009) Determination of preferred pH for root-knot nematode aggregation using pluronic F-127 gel. *Journal of Chemical Ecology* **35**, 1242–1251.
- Williams DC, Bejjani RE, Ramirez PM, Coakley S, Kim SA, Lee H, Wen Q, Samuel A, Lu H, Hilliard MA and Hammarlund M (2013) Rapid and permanent neuronal inactivation *in vivo* via subcellular generation of reactive oxygen with the use of KillerRed. *Cell Reports* **5**, 553–563.
- Wrather JA, Koenning SR and Anderson TR (2003) Effect of diseases on soybean yields in the United States and Ontario (1999–2002). *Plant Health Progress*. doi: 10.1094/PHP-2003-0325-01-RV.
- Wu Y, Wickham JD, Zhao L and Sun J (2019) CO₂ drives the pine wood nematode off its insect vector. *Current Biology* **29**, R619–R620.
- Zawadzki JL, Kotze AC, Fritz JA, Johnson NM, Hemsworth JE, Hines BM and Behm CA (2012) Silencing of essential genes by RNA interference in *Haemonchus contortus*. *Parasitology* **139**, 613–629.
- Zhu H, Li J, Nolan TJ, Schad GA and Lok JB (2011) Sensory neuroanatomy of *Parastrongyloides trichosuri*, a nematode parasite of mammals: amphidial neurons of the first-stage larva. *Journal of Comparative Neurology* **519**, 2493–2507.
- Zimmer M, Gray JM, Pokala N, Chang AJ, Karow DS, Marletta MA, Hudson ML, Morton DB, Chronis N and Bargmann CI (2009) Neurons detect increases and decreases in oxygen levels using distinct guanylate cyclases. *Neuron* **61**, 865–879.