

## Original Article

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# Investigating the shared genetic architecture of post-traumatic stress disorder and gastrointestinal tract disorders: a genome-wide cross-trait analysis

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**Abstract**

**Background.** Observational studies suggest a correlation between post-traumatic stress disorder (PTSD) and gastrointestinal tract (GIT) disorders. However, the genetic overlap, causal relationships, and underlying mechanisms between PTSD and GIT disorders were absent.

**Methods.** We obtained genome-wide association study statistics for PTSD (23 212 cases, 151 447 controls), peptic ulcer disease (PUD; 16 666 cases, 439 661 controls), gastroesophageal reflux disease (GORD; 54 854 cases, 401 473 controls), PUD and/or GORD and/or medications (PGM; 90 175 cases, 366 152 controls), irritable bowel syndrome (IBS; 28 518 cases, 426 803 controls), and inflammatory bowel disease (IBD; 7045 cases, 449 282 controls). We quantified genetic correlations, identified pleiotropic loci, and performed multi-marker analysis of genomic annotation, fast gene-based association analysis, transcriptome-wide association study analysis, and bidirectional Mendelian randomization analysis.

**Results.** PTSD globally correlates with PUD ( $r_g = 0.526$ ,  $p = 9.355 \times 10^{-7}$ ), GORD ( $r_g = 0.398$ ,  $p = 5.223 \times 10^{-9}$ ), PGM ( $r_g = 0.524$ ,  $p = 1.251 \times 10^{-15}$ ), and IBS ( $r_g = 0.419$ ,  $p = 8.825 \times 10^{-6}$ ). Cross-trait meta-analyses identify seven genome-wide significant loci between PTSD and PGM (rs13107325, rs1632855, rs1800628, rs2188100, rs3129953, rs6973700, and rs73154693); three between PTSD and GORD (rs13107325, rs1632855, and rs3132450); one between PTSD and IBS/IBD (rs4937872 and rs114969413, respectively). Proximal pleiotropic genes are mainly enriched in immune response regulatory pathways, and in brain, digestive, and immune systems. Gene-level analyses identify five candidates: *ABTI*, *BTN3A2*, *HIST1H3J*, *ZKSCAN4*, and *ZKSCAN8*. We found significant causal effects of GORD, PGM, IBS, and IBD on PTSD. We observed no reverse causality of PTSD with GIT disorders, except for GORD.

**Conclusions.** PTSD and GIT disorders share common genetic architectures. Our work offers insights into the biological mechanisms, and provides genetic basis for translational research studies.

**Introduction**

Post-traumatic stress disorder (PTSD) is a prevalent psychiatric disorder that occurs in some individuals after a traumatic event (Duncan et al., 2018). PTSD poses strong social impacts: suicide, hospitalization, and substance use (Davidson, 2000). Although scholars have now begun to prioritize PTSD in ameliorating various social burdens, fundamental questions remain on the genetic etiology of PTSD.

Gastrointestinal tract (GIT) disorders frequently co-occur with psychiatric diagnoses marked by irritability, fearfulness, hypervigilance, and physiological mobilization (Kolacz, Kovacic, & Porges, 2019). GIT-psychiatric comorbidities emerge in a range of psychiatric disorders: PTSD, anxiety, and depression (Graff, Walker, & Bernstein, 2009; Grinsvall, Tornblom, Tack, Van Oudenhove, & Simren, 2018; Hejazi & McCallum, 2014; Henningsen, Zimmermann, & Sattel, 2003). Growing evidence suggest traumatic experiences are related to GIT disorders: a history of abuse exacerbates symptoms of irritable bowel disease (IBD) (Drossman et al., 2018; Leserman, 2005); acute early-life stressors induce long-term sensory and motor digestive changes in irritable bowel syndrome (IBS) (O'Mahony, Hyland, Dinan, & Cryan, 2011; Vannucchi & Evangelista, 2018). However, whether GIT disorders are a risk factor for PTSD and vice versa are unclear.

Large-scale genome-wide association studies (GWASs) have documented a growing number of single-nucleotide polymorphisms (SNPs), genes, and susceptibility loci respectively for PTSD and GIT disorders (An et al., 2019; Nievergelt et al., 2019; Watanabe et al., 2019; Wu et al., 2021), providing a basis to evaluate the genetic etiology of PTSD and GIT disorders including gastroesophageal reflux disease (GORD), peptic ulcer disease (PUD), PUD and/or GORD and/or medications (PGM, an alternative combination of diagnosis and/or treatments for PUD and/or GORD), IBS, and IBD. Nevertheless, no study has reported their shared genetic architectures.

Here, we performed a comprehensive genome-wide cross-trait analysis to investigate the genetic overlaps and the causal relationships between PTSD and GIT disorders. We quantified the global and local genetic correlations, analyzed partitioned heritability and cell-type-specific enrichment, identified pleiotropic loci, performed functional annotations, conducted gene-level analyses, and inferred putative causal associations.

## Methods

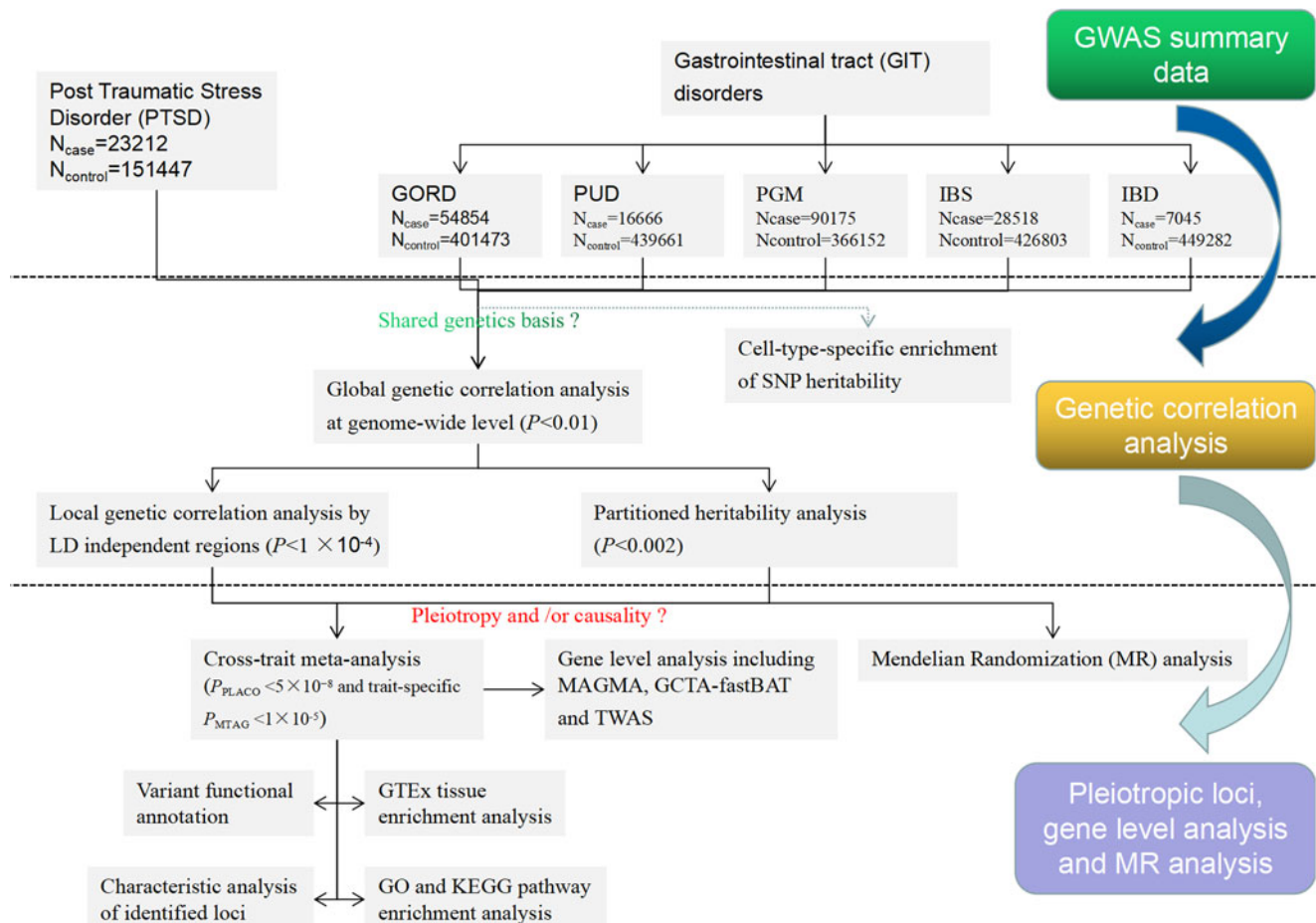
### GWAS summary statistics

Overall study design is shown in Fig. 1. GWAS data are summarized in online Supplementary Table S1. We obtained GWAS summary statistics for PTSD from the Psychiatric Genomics Consortium (PGC) comprising 23 212 cases and 151 447 controls

of European ancestry (Nievergelt et al., 2019). In the original study, PTSD assessment was based on lifetime or current PTSD, and the unaffected control individuals did not have a PTSD diagnosis; PTSD was diagnosed using various instruments and different versions of the Diagnostic and Statistical Manual of Mental Disorders; quality control was performed by PGC pipeline and dataset was imputed by the 1000 Genomes European panel. We obtained GWAS for GIT traits including PUD (16 666 cases and 439 661 controls), GORD (54 854 cases and 401 473 controls), IBS (28 518 cases and 426 803 controls), and IBD (7045 cases and 449 282 controls) from a recent publication (Wu et al., 2021). Clinically, PUD medications are indicated in GORD and gastritis. Accordingly, GWAS combining diagnosis for PGM commonly used for these disorders was conducted, and we used this GWAS data (90 175 cases and 366 152 controls) as a proxy for PUD or GORD (Wu et al., 2021). The criteria for GIT disorders are: diagnosis phenotypes from death register, self-report, hospital admission, or primary care record; treatment phenotypes based on the operation and medication-taking code. Dataset was imputed to a mixed panel of UK10K+1000 Genomes and Haplotype Reference Consortium (Bycroft et al., 2018).

### Genetic correlation

We conducted a post-GWAS global correlation analysis by linkage disequilibrium (LD) score regression (LDSC) (Bulik-Sullivan



**Figure 1.** Overall study design of genome-wide cross-trait analysis.

et al., 2015). LDSC estimates correlation between true causal effects of two traits (ranging from  $-1$  to  $1$ ). SNPs in a high LD region would have a larger  $\chi^2$ , and a similar relationship would appear when single-study statistics are replaced by the product of  $z$ -scores from two studies of traits with correlation. We performed LDSC with unconstrained intercept using pre-computed LD scores from the 1000 Genomes Project Phase 3 (European ancestry), and restricted our analysis to only haplotype map 3 (HapMap3) SNPs to minimize bias of low imputation quality (Bulik-Sullivan et al., 2015). We set Bonferroni-corrected significant threshold at a  $p$  value of  $0.01$  ( $0.05/5 = 0.01$ ).

We calculated pairwise local correlations for PTSD and GIT disorders in 2343 pre-specified LD-independent segments using the SUPER GeNetic cOVariance Analyzer (SUPERGNOVA), which identifies locally associated small contiguous genomic regions using GWAS summary statistics and a reference panel as input data, and provides  $p$  values ( $P_{\text{SUPERGNOVA}}$ ) between pairs of traits in local regions (Zhang et al., 2021). Here, we removed SNPs with missing values, used the 1000 Genomes Project, with rare variants (minor allele frequency  $< 5\%$ ) filtered out, as the reference panel to generate the genome partition files in SUPERGNOVA.  $P_{\text{SUPERGNOVA}} < 1 \times 10^{-4}$  was used as a cut-off value to indicate significance (Chen, Wang, Huang, & Jia, 2022).

### SNP heritability and cell-type-specific enrichment

We partitioned SNP heritability of PTSD and GIT disorders respectively by stratified LDSC, to determine if SNPs sharing high heritability are clustered in the 24 main functional annotations (Finucane et al., 2015). We retrieved LD scores, regression weights, and allele frequencies from European samples (<https://alkesgroup.broadinstitute.org/LDSCORE>). We assumed the estimates to be significant if  $p$  value surpasses an  $\alpha$  level of  $0.002$ , derived by Bonferroni correction.

We performed heritability partitioning for the 396 annotations constructed by the Roadmap project for six chromatin marks ('DHS', 'H3K27ac', 'H3K36me3', 'H3K4me1', 'H3K4me3', 'H3K9ac') in a set of 88 primary cell types or tissues (Finucane et al., 2018). Each annotation corresponds to a chromatin mark in a single-cell type. These 396 cell-type-specific annotations belong to nine groups: adipose, central nervous system, digestive system, cardiovascular, musculoskeletal, and connective tissue, immune and blood, liver, pancreas, and other. We calculated annotation-specific enrichment values for each trait using LDSC, and performed visualization by hierarchical clustering. We used adjusted  $p$  value to correct for multiple testing.

### Cross-trait meta-analysis

Multi-trait analysis of GWAS (MTAG) applies generalized inverse-variance-weighted meta-analysis for multiple correlated traits, and detects genetic associations for each trait by borrowing correlations among correlated traits to boost statistical power (Guo et al., 2022; Tadros et al., 2021; Yang et al., 2021). We performed MTAG and implemented options that assume equal SNP heritability for each trait (Turley et al., 2018). We denoted the single-trait GWAS statistics as  $\text{GWAS}_{\text{PTSD}}$ ,  $\text{GWAS}_{\text{PUD}}$ ,  $\text{GWAS}_{\text{GORD}}$ ,  $\text{GWAS}_{\text{PGM}}$ ,  $\text{GWAS}_{\text{IBS}}$ , and  $\text{GWAS}_{\text{IBD}}$ , respectively, and the PTSD statistics from MTAG analysis as  $\text{MTAG}_{\text{PTSD}}$ . Using functional mapping and annotation (FUMA) platform (Watanabe, Taskesen, van Bochoven, & Posthuma, 2017), we identified genome-wide

significant SNPs of  $\text{MTAG}_{\text{PTSD}}$  ( $p < 5.0 \times 10^{-8}$ ; pairwise  $R^2 < 0.6$  in a 1 Mb window), and conducted gene-set and tissue expression analyses by multi-marker analysis of genomic annotation (MAGMA v.1.08, implemented in FUMA). For gene-set analysis, we used gene ontology (GO) gene sets from the Molecular Signatures Database (MSigDB, v.6.2). For tissue analysis, we used data from Genotype-Tissue Expression project (GTEx, v.8). For comparison, we additionally performed annotations of the significant SNPs identified from  $\text{GWAS}_{\text{PTSD}}$  using the FUMA platform.

To determine if assumption violations biased the MTAG results, we performed pleiotropic analysis under composite null hypothesis (PLACO) as a sensitivity analysis (Ray & Chatterjee, 2020). PLACO identifies pleiotropic loci by testing the composite null hypothesis that a locus is associated with zero or one of the traits (Ray et al., 2021). We prioritized independent SNPs with genome-wide significance in both MTAG and PLACO ( $P_{\text{PLACO}} < 5 \times 10^{-8}$ , and trait-specific  $P_{\text{MTAG}}$  for PTSD  $< 1 \times 10^{-5}$ ), following identification by LD clumping ( $r^2 < 0.2$  within 500-kilobase windows) in 'PLINK v1.9' (<https://www.cog-genomics.org/plink/1.9>), based on 1000 Genomes reference data (Purcell et al., 2007).

### Functional annotation

To annotate functions of the identified pleiotropic SNPs, we used Ensembl variant effect predictor and human variation annotation database (<http://www.licpathway.net/VARADB>) (Pan et al., 2021; Zerbino et al., 2018). We then queried these loci in the GWAS catalogue (<https://www.ebi.ac.uk/gwas>) to determine their identity in PTSD and GIT disorders.

We assessed GO biological process functional categories and Kyoto encyclopedia of genes and genomes pathways for enriched pleiotropic SNPs between PTSD and five GIT traits. We performed tissue enrichment using 'GENE2FUNC' function in the FUMA platform with 30 general tissue types from GTEx v.8 (GTEx Consortium et al., 2017; Watanabe et al., 2017). We applied Benjamini-Hochberg procedure to correct for multiple testing.

### Gene-level analysis

Using methods with different assumptions to obtain overlapped signals can avoid the risk of false discoveries. Thus, we applied three parallel gene-level analyses with distinct principles: MAGMA, genome-wide complex trait analysis-fast set-based association analysis (GCTA-fastBAT), and transcriptome-wide association study (TWAS) analysis.

MAGMA uses a multiple regression structure to allow generalized analysis of continuous properties of genes and simultaneous analysis of multiple gene-sets and other gene properties (de Leeuw, Mooij, Heskes, & Posthuma, 2015). Here, we submitted summary statistics for  $\text{MTAG}_{\text{PTSD}}$  in MAGMA to analyze all protein-coding genes to which at least one SNP is annotated within a 0 kb interval. We applied Bonferroni correction for multiple testing based on the number of genes tested ( $P_{\text{MAGMA}} = 2.66 \times 10^{-6}$ ).

GCTA-fastBAT analyzes human complex traits using summary-level data from GWAS and LD data from a reference sample with individual-level genotypes (Bakshi et al., 2016). Here, we conducted a gene-based analysis by GCTA-fastBAT using  $\text{MTAG}_{\text{PTSD}}$  for all 24 763 genes and LD information from 1000 Genomes Project Phase 3. We only analyzed the SNPs located within the gene to examine the gene-trait associations.

We set Bonferroni-corrected significance at  $P_{\text{fastBAT}} < 2.02 \times 10^{-6}$  (0.05/24 763).

TWAS integrates GWAS and expression quantitative trait locus (eQTL) data to identify tissue-specific gene–trait associations (Hu et al., 2019). We used MTAG<sub>PTSD</sub> and the unified test for molecular signatures (UTMOST) program combined with generalized Berk–Jones (GBJ) models in a two-stage TWAS analysis (<https://github.com/Joker-Jerome/UTMOST>). We used GBJ models with transcriptome data derived from 44 different tissues (GTEx, v.6). We set Bonferroni-corrected significance at  $P_{\text{TWAS}} < 3.24 \times 10^{-6}$  (0.05/15 430).

### Causal inference

We conducted Mendelian randomization (MR) analysis with R packages ‘TwoSampleMR’ and ‘MR-PRESSO’. We used five MR methods: inverse-variance weighted (IVW) (Burgess, Butterworth, & Thompson, 2013), MR–Egger (Burgess & Thompson, 2017), weighted median (Bowden, Davey Smith, Haycock, & Burgess, 2016), weighted mode (Hartwig, Davey Smith, & Bowden, 2017), and MR-PRESSO (Verbanck, Chen, Neale, & Do, 2018), with different assumptions in the extent and nature of horizontal pleiotropy. We applied the IVW method as our primary approach (Burgess, Scott, Timpson, Davey Smith, & Thompson, 2015), and we considered relationships with consistent evidence of causality in all MR methods to be more reliable and noteworthy.

IVW assumes if an uncorrelated pleiotropy exists, a mean of zero adds noise only to regressions with multiplicative random effects. MR–Egger allows the presence of directional (i.e. non-zero mean) uncorrelated pleiotropy, and adds an intercept to the IVW regression to exclude confounding. Weighted median measures the median rather than the mean of SNP ratio, and identifies true causality if  $\leq 50\%$  of the weights are from invalid SNPs. Weighted model divides SNPs into groups based on estimated effects, assesses evidence of causality using only the group with the most SNPs, essentially relaxing the hypothesis, and identifies true effects even if most tools are invalid. In these MR analyses, we used independent SNPs (LD clumping  $r^2 < 0.05$  within 1000-kb windows using PLINK v1.9) associated with the ‘exposure’ trait ( $p < 1 \times 10^{-5}$ ) as instrumental variables, and merged with SNPs from the ‘outcome’ trait in Harmonizing data step. To avoid sample overlapping effect, we used the function ‘power\_prune’ from R package ‘TwoSampleMR’ in Harmonizing data step.

## Results

### Genetic correlations

With unconstrained intercept, we found positive global correlations of PTSD with PUD ( $r_g = 0.5261$ ,  $p = 9.3549 \times 10^{-7}$ ), GORD ( $r_g = 0.3977$ ,  $p = 5.2225 \times 10^{-9}$ ), PGM ( $r_g = 0.5236$ ,  $p = 1.2506 \times 10^{-15}$ ), IBS ( $r_g = 0.4185$ ,  $p = 8.8245 \times 10^{-6}$ ), and a marginal correlation with IBD ( $r_g = 0.179$ ,  $p = 0.0558$ ) (Table 1). We identified no statistically significant local correlation of PTSD with GIT disorders in specific genomic regions (online Supplementary Data files S1–S5).

### Partitioned SNP heritability and cell-type-specific enrichment

In partitioned SNP heritability analysis, we found significant enrichments of the functional categories ‘conserved’, ‘FetalDHS’,

**Table 1.** Genome-wide global correlations between PTSD and GIT disorders by unconstrained LD score regression

Trait 1	Trait 2	Unconstrained		
		$r_g$	$r_{g\_se}$	$p$
PTSD	PUD	0.526	0.107	$9.355 \times 10^{-7}$
	PGM	0.524	0.066	$1.251 \times 10^{-15}$
	GORD	0.398	0.068	$5.223 \times 10^{-9}$
	IBS	0.419	0.094	$8.825 \times 10^{-6}$
	IBD	0.179	0.094	0.056

PTSD, post-traumatic stress disorder; PUD, peptic ulcer disease; PGM, PUD and/or GORD and/or medications; GORD, gastroesophageal reflux disease; IBS, irritable bowel syndrome; IBD, inflammatory bowel disease;  $r_g$ , genetic correlation, se, standard error; GIT, gastrointestinal tract.

‘H3K27ac’, ‘H3K4me1’, ‘H3K4me3’, ‘Intron’, ‘SuperEnhancer’, ‘TSS’, and ‘WeakEnhancer’ for PGM, ‘conserved’ for GORD and IBS, and ‘Enhancer’, ‘super Enhancer’, ‘TFBS’, and ‘H3K27ac’ for IBD. We observed no functional categories enrichment for PUD and PTSD (online Supplementary Table S2).

Partitioning SNP heritability using 396 cell-type-specific annotations, we found significant enrichments for GORD and IBS in the central nervous and digestive systems: brain dorsolateral prefrontal cortex, brain inferior temporal lobe, and fetal stomach. We observed significant enrichments for IBD in the blood and immune system: primary natural killer cells and primary T helper 17 cells. We found insignificant enrichments for PUD, while enrichments are clustered closely to GORD/IBS at each chromatin mark in the central nervous and digestive systems. Comparing enrichments for all GIT disorders, we observed differential patterns, where only IBD is enriched in the immune system (online Supplementary Data files S6–S11).

### Cross-trait meta-analysis and pleiotropic loci

In meta-analysis of GWAS<sub>PTSD</sub>, GWAS<sub>PUD</sub>, GWAS<sub>GORD</sub>, GWAS<sub>PGM</sub>, GWAS<sub>IBS</sub>, and GWAS<sub>IBD</sub> using MTAG, we obtained a total of 6 443 162 SNPs, and we identified 541 genome-wide significant SNPs in MTAG<sub>PTSD</sub> with FUMA annotations (online Supplementary Data file S12). The Manhattan plots for GWAS<sub>PTSD</sub> and MTAG<sub>PTSD</sub> are shown in online Supplementary Fig. S1. We found the number of risk loci increases from two in GWAS<sub>PTSD</sub> to seven in MTAG<sub>PTSD</sub>, and the number of lead SNPs ( $P_{\text{MTAG}} < 5 \times 10^{-8}$  and  $R^2 < 0.1$ ) increases from two in GWAS<sub>PTSD</sub> to eight in MTAG<sub>PTSD</sub> (online Supplementary Tables S3 and S4). In MAGMA analysis for MTAG<sub>PTSD</sub>, we identified significant gene-sets linked to regulation of biosynthetic process and regulation of gene expression function, and we associated MTAG<sub>PTSD</sub> expressions in brain tissues (brain frontal cortex BA9, brain anterior cingulate cortex BA24, brain cortex, brain cerebellar hemisphere, brain cerebellum, brain hypothalamus) (online Supplementary Data files S13 and S14).

Using both MTAG and PLACO, we identified seven loci with a genome-wide significance in the meta-analysis of PTSD and PGM (rs13107325, rs1632855, rs1800628, rs2188100, rs3129953, rs6973700, and rs73154693); three in the meta-analysis of PTSD and GORD (rs13107325, rs1632855, and rs3132450); and one in the meta-analysis of PTSD and IBS/IBD (rs4937872 and rs114969413, respectively) (online Supplementary Tables S5 and

S6). We observed no genome-wide significant locus in the meta-analysis of PTSD and PUD. We queried each locus for pleiotropic associations with other traits via GWAS catalogue, and identified proximal genes: *SLC39A8*, *MUC21*, *MUC22*, *TNF*, *LTB*, *FAM209A/FAM209B*, and *PRRC2A* (online Supplementary Tables S7 and S8). In functional and tissue enrichment analysis of proximal genes, we found that PTSD shares ‘immune response related regulation’ with PGM and IBD, and ‘protein processing’ and ‘regulation of acute inflammatory response’ with IBD (online Supplementary Tables S9 and S10). Using GTEx data, we found significant enriched expression of PTSD- and PGM-shared genes in blood, and PTSD- and IBD-shared genes in brain and liver (online Supplementary Figs S2 and S3). We found no statistically significant enrichment for other traits.

**Gene-level analysis**

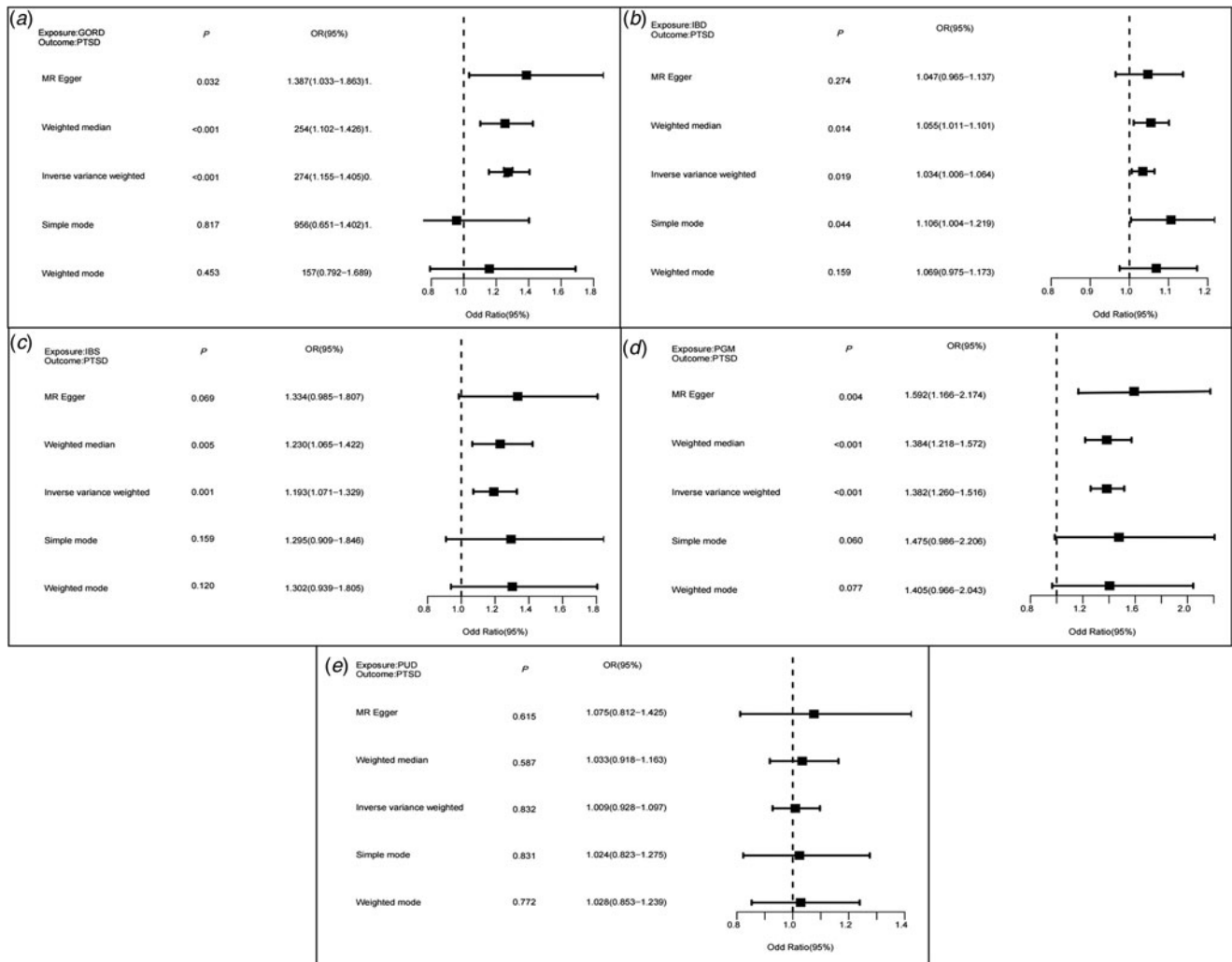
In MAGMA, we observed 35 PTSD-related genes that survive correction for multiple testing; in GCTA-fastBAT, we found 93 genes at the significant level ( $P_{\text{fastBAT}} < 2.02 \times 10^{-6}$ ); in TWAS

combining eQTL data from 44 tissues, we revealed 31 genes with Bonferroni-corrected significance (online Supplementary Tables S11–S13).

We checked the overlapping genes identified by MAGMA, GCTA-fastBAT, and TWAS, and identified five PTSD-associated candidates: *ABT1* ( $P_{\text{MAGMA}} = 7.76 \times 10^{-7}$ ,  $P_{\text{fastBAT}} = 1.38 \times 10^{-8}$ ,  $P_{\text{TWAS}} = 1.57 \times 10^{-6}$ ), *BTN3A2* ( $P_{\text{MAGMA}} = 2.60 \times 10^{-8}$ ,  $P_{\text{fastBAT}} = 7.52 \times 10^{-10}$ ,  $P_{\text{TWAS}} = 9.94 \times 10^{-8}$ ), *HIST1H3J* ( $P_{\text{MAGMA}} = 1.25 \times 10^{-8}$ ,  $P_{\text{fastBAT}} = 1.15 \times 10^{-6}$ ,  $P_{\text{TWAS}} = 1.87 \times 10^{-8}$ ), *ZKSCAN4* ( $P_{\text{MAGMA}} = 4.22 \times 10^{-7}$ ,  $P_{\text{fastBAT}} = 8.89 \times 10^{-8}$ ,  $P_{\text{TWAS}} = 2.95 \times 10^{-12}$ ), and *ZKSCAN8* ( $P_{\text{MAGMA}} = 4.72 \times 10^{-8}$ ,  $P_{\text{fastBAT}} = 2.60 \times 10^{-8}$ ,  $P_{\text{TWAS}} = 5.71 \times 10^{-7}$ ). For the proximal genes of pleiotropic loci identified by our cross-trait meta-analysis, we found *PRRC2A* (MAGMA and GCTA-fastBAT), *MUC21* and *MUC22* (GCTA-fastBAT) are significant in our gene-level analysis.

**Causal associations**

In IVW MR analysis, we found significant causal effects of PGM, GORD, IBS, and IBD on PTSD (Fig. 2), estimates remain



**Figure 2.** MR analyses for causal effects of GIT disorders on PTSD. Boxes represent the point estimates of causal effects, and error bars represent 95% confidence intervals. Inverse-variance-weighted approach was adopted as the primary analysis. PTSD, post-traumatic stress disorder; PUD, peptic ulcer disease; PGM, PUD and/or GORD and/or medications; GORD, gastroesophageal reflux disease; IBS, irritable bowel syndrome; IBD, inflammatory bowel disease; GIT, gastrointestinal tract.

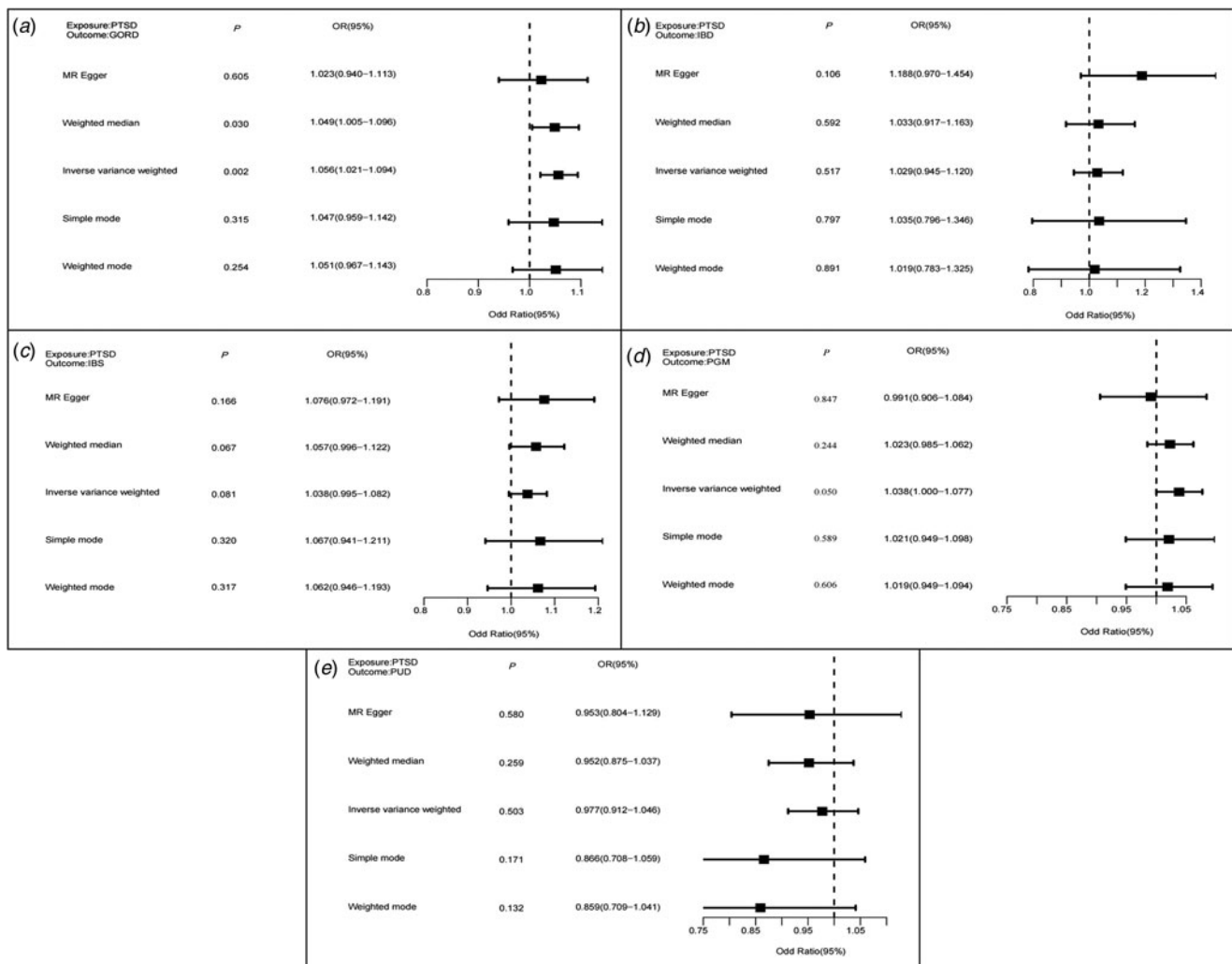
directionally consistent in MR-Egger and weighted median despite larger statistical uncertainties, and MR-PRESSO sensitivity analysis supports a causal association of four GIT disorders with PTSD (online Supplementary Table S14). In reverse-direction MR analysis, we found no statistically significant causal effects of PTSD on GIT disorders, except for GORD (Fig. 3 and online Supplementary Table S14).

## Discussion

With large-scale GWAS summary data and multiple statistical approaches, we present a comprehensive assessment of shared genetic architecture between PTSD and GIT disorders. We identified significant genetic correlations between all five GIT disorders with PTSD in global correlation analysis, and we did not find a local correlation at statistical significance, suggesting that current analyses are underpowered to detect correlation in specific genomic regions between PTSD and GIT disorders. Significant genetic correlation reflects not only a common genetic etiology (biological pleiotropy), but also suggests a potential causal effect (vertical pleiotropy). In agreement, we observed causal

associations of four GIT disorders (IBD, IBS, PGM, and GORD) with PTSD using IVW and MR-PRESSO, and we found the estimates remain directionally consistent in MR-Egger and weighted median approaches.

In the cross-trait meta-analysis of PTSD, we found a genome-wide significant locus rs13107325 for both PGM and GORD. This independent SNP is mapped to the gene *SLC39A8*, encoding a metal cation transporter Zrt-/Irt-related protein (ZIP8) in all vertebrates. Missense mutation in *SLC39A8* leads to reduced zinc transport, which in turn perturbs electrophysiological recordings of neurons, resulting in a significant reduction in spontaneous excitatory postsynaptic currents and reduced surface expression of glutamate receptors (Kalappa, Anderson, Goldberg, Lippard, & Tzounopoulos, 2015; Tseng *et al.*, 2021). A recent review recognizes *SLC39A8* exhibits significant pleiotropic effects associated with clinical diseases in virtually every organ, tissue, and cell type (Nebert & Liu, 2019). Among the central nervous system diseases, *SLC39A8* is associated with Parkinson's disease and schizophrenia (Pickrell *et al.*, 2016; Schizophrenia Working Group of the Psychiatric Genomes Consortium, 2014), and the latter is possibly related to putamen gray matter volume mediated by



**Figure 3.** MR analyses for causal effects of PTSD on GIT disorders. Boxes represent the point estimates of causal effects, and error bars represent 95% confidence intervals. Inverse-variance-weighted approach was adopted as the primary analysis. PTSD, post-traumatic stress disorder; PUD, peptic ulcer disease; PGM, PUD and/or GORD and/or medications; GORD, gastroesophageal reflux disease; IBS, irritable bowel syndrome; IBD, inflammatory bowel disease; GIT, gastrointestinal tract.

*SLC39A8* variants and expression (Luo et al., 2019). In a phenome-wide approach, functional *SLC39A8* variants are correlated with brain-related traits including delirium and dementia (McCoy, Pellegrini, & Perlis, 2019). In the immune system, reduced *SLC39A8* expression decreases blood–brain barrier integrity, increases interleukin-6/interleukin-1 $\beta$  and nuclear factor kappa B expression following stimulation of tumor necrosis factor  $\alpha$ , suggesting that ZIP8 dysfunction may exacerbate immune and inflammatory signaling (Kebir et al., 2007; Liu et al., 2013; Melia et al., 2019). In a gastrointestinal system, a *SLC39A8* variant is associated with Crohn disease and altered colonic mucosal microbiota (Li et al., 2016).

Among the seven loci identified in our meta-analysis of PTSD and PGM, rs6973700 is mapped to mitotic spindle-assembly checkpoint gene *MAD1L1*, which correlates with psychiatric susceptibility (Liu et al., 2021). For the meta-analysis of PTSD and IBS, we found that an independent SNP rs4937872, mapped to the neural cell adhesion molecule (*NCAM1*) gene, is associated with multiple psychiatric disorders (Shiwaku et al., 2022; Wendt et al., 2022). Among genes identified in our gene-level analysis, *BTN3A2* encodes a member of the immunoglobulin superfamily in the juxta-telomeric region of the major histocompatibility class 1 locus (Wu et al., 2019). *BTN3A2* is significantly up-regulated in induced pluripotent stem cells derived neurons from schizophrenia patients and gastric tumors (Wu et al., 2019; Zhu et al., 2017). Thus, these genes might serve as potential candidates for both PTSD and GIT disorders.

In the pathway-based analysis for PTSD and GIT disorders (PGM and IBD), we observed a significantly enriched pathway ‘regulation of immune response’, which plays a critical role in the brain–gut axis (Clark & Mach, 2016; Roth, Zadeh, Vekariya, Ge, & Mohamadzadeh, 2021). Immune dissonance represents a key element both in the pathogenesis of PTSD complications and in tissue repair (Jiang, 2008), while the adaptive immune response plays a major role in the pathogenesis of GIT disorders (Jacenic & Fichna, 2020). Analyzing SNP heritability enrichments for PTSD and GIT disorders, we found that GORD and IBS are mainly enriched in the central nervous and digestive systems, which is consistent with pathway-based analysis involving the brain–gut axis; and IBD is mainly enriched in the immune system, which emphasizes the vital role of immune response in PTSD and GIT disorders.

### Limitations

We acknowledge several limitations of our study. First, to avoid bias from population stratification, we restricted all data to European ancestry, undermining generalization in other ethnic groups. Second, despite the large sample sizes of the consortium-based meta-analysis, the number of participants and SNPs differ among studies, limiting the direct comparison between different GIT disorders. Third, data from the UK biobank constitutes approximately 35% of the effective sample size of PTSD GWAS summary statistics, leading to considerable sample overlap in our study. Accordingly, we performed LDSC without a constrained intercept and used ‘power\_prune’ function in MR analysis to minimize bias from sample overlap.

In conclusion, our study provides genetic insights into the association of PTSD and GIT disorders, and demonstrates a shared genetic susceptibility. We observed causal effects of GORD, PGM, IBS, and IBD on PTSD, and identified several genomic loci and genes shared by PTSD and GIT disorders,

particularly the *SLC39A8* gene (or isoforms), allowing translational and research implications for PTSD, GIT disorders and their co-morbidities.

**Supplementary material.** The supplementary material for this article can be found at <https://doi.org/10.1017/S0033291723001423>.

**Data.** All data are available in the main text or in the Supplementary materials.

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**Conflict of interest.** The authors declare no competing interest.

**Ethical standards.** The authors assert that all procedures contributing to this work comply with the ethical standards of the relevant national and institutional committees on human experimentation and with the Helsinki Declaration of 1975, as revised in 2008, and the relevant national and institutional guides on the care and use of laboratory animals.

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