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Climate change effects on terrestrial parasitic nematodes: Where are the knowledge gaps?

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Abstract

Climate change is expected to affect parasitic nematodes and hence possibly parasite-host dynamics and may have far-reaching consequences for animal health, livestock production, and ecosystem functioning. However, there has been no recent overview of current knowledge to identify how studies could contribute to a better understanding of terrestrial parasitic nematodes under changing climates. Here we screened almost 1,400 papers to review 57 experimental studies on the effects of temperature and moisture on hatching, development, survival, and behaviour of the free-living stages of terrestrial parasitic nematodes with a direct life cycle in birds and terrestrial mammals. Two major knowledge gaps are apparent. First, research should study the temperature dependency curves for hatching, development, and survival under various moisture treatments to test the interactive effect of temperature and moisture. Second, we specifically advocate for more studies that investigate how temperature, and its interaction with moisture, affect both vertical and horizontal movement of parasitic nematodes to understand infection risks. Overall, we advocate for more field experiments that test environmental effects on life-history traits and behaviour of parasitic nematodes in their free-living stages under natural and realistic circumstances. We also encourage studies to expand the range of used hosts and parasitic nematodes because 66% of results described in the available studies use sheep and cattle as hosts and 32% involve just three nematode species. This new comprehension brings attention to understudied abiotic impacts on terrestrial parasitic nematodes and will have broader implications for livestock management, wildlife conservation, and ecosystem functioning in a rapidly warming climate.

Introduction

Parasitic nematodes are a group of multicellular organisms infecting and occupying a diverse range of hosts and ecosystems (Chung & Boeri 2012; Aleuy & Kutz 2020). They are a cause of animal disease (Charlier et al. 2009; Mehlhorn 2016) and can potentially impact the dynamics of wild populations (Tompkins et al. 2002), food webs (Lafferty et al. 2006), and prey-predator interactions (Hudson et al. 1992; Hatcher et al. 2006). Parasitic nematodes and their development, survival, and distribution/migration in and on the herbage are sensitive to abiotic factors such as moisture and temperature (Crofton 1965; Stromberg 1997; O' Connor et al. 2006; Gyeltshen et al. 2022). Thus, changing environmental factors associated with climate change are expected to affect parasitic nematodes, with possible consequences for parasite-host dynamics (Kutz et al. 2009; Molnár, Kutz et al. 2013; Dobson et al. 2015). However, there has been no recent review of the knowledge gaps about the effect of temperature and moisture on direct life cycle parasitic nematodes in birds and mammals. Highlighting the knowledge gaps about abiotic effects, and particularly the interaction of moisture and temperature (O' Connor et al. 2006) on parasitic nematodes, could guide future research and provide a better understanding of the implications for livestock health management, wildlife conservation, and ecosystem functioning in a rapidly warming climate.

In this review, we focus on parasitic nematodes with a direct life cycle. This group of parasites has one definitive host and a free-living stage outside the host where they occupy highly diverse habitats (e.g., faeces and herbage; Figure 1) and may be vulnerable to changes in abiotic conditions (Molnár, Dobson *et al.* 2013; Molnár, Kutz *et al.* 2013). In an attempt to understand environmental effects on parasitic nematodes with a direct life cycle, many studies have focused on the seasonal correlation between environmental factors and nematode abundance (Chaudary *et al.* 2008; van Dijk *et al.* 2010; Halvorsen 2012; Holand *et al.* 2019) and behaviour (Langrová *et al.* 2003; Kuzmina *et al.* 2006; Santos *et al.* 2012). Although seasonality studies are important in shedding light into the role of annual environmental change, the underlying mechanisms influencing the parasite dynamics often remain unknown (Altizer *et al.* 2006) and need to be disentangled with experimental studies. Therefore, to tease apart the diverse factors and mechanisms that affect parasite—host dynamics, we have focused on reviewing experimental field and laboratory studies on the effect of

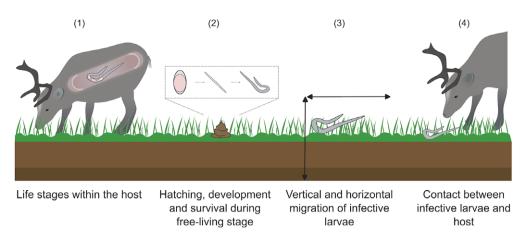


Figure 1. Schematic figure of the life cycle of parasitic nematodes with a direct life cycle that occupy highly diverse habitats (faeces, herbage, and inside the host) during the different life stages. 1) Parasitic nematodes with a direct life cycle have one definite host, in which exsheathment, reproduction, survival and egg production occurs. 2) In the free-living stage outside the host, eggs develop into infective larvae. 3) Infective larvae move out of the faeces and 4) are ingested by the host again.

abiotic conditions on parasitic nematodes with a direct life cycle in birds and mammals. Our primary aim is to identify the knowledge gaps by reviewing the literature describing studies that manipulated the effects of temperature, moisture, and precipitation on hatching, development, survival, and behaviour of parasitic nematodes in the free-living stage.

Methods

For this review, we researched articles in Web of Science (© Copyright Clarivate 2021) on the effects of temperature, precipitation, and moisture on parasitic nematodes, with the last search date on 5 December 2022. The search string employed was (Parasit*) AND (nematod* OR roundworm) AND (temperature OR moisture OR humidity OR precipitation). This search resulted in 1,391 papers. From this total, only studies that fulfilled all the following criteria were included in the subsequent analysis: a) parasite with direct life cycle, b) parasite of terrestrial mammal or bird, c) effect of temperature, moisture, precipitation, or humidity on life-history traits (hatching, development, or survival) or behaviour of parasitic nematodes in the free-living stage, and d) experimental study under laboratory or field conditions. We specifically excluded studies on the effect of abiotic factors on the soil nematode community and studies not written in English. Most of the excluded studies were on plant and insect parasitic nematodes, soil communities, or observational studies without experimental manipulation. Finally, we added one relevant study (PhD thesis) that was missed by the literature search but that was previously known (Hoar 2012) to give a total of 57 relevant papers to review. In the next section, we synthesize the results of the reviewed papers. When findings are based on evidence from three or more papers, we only refer to the oldest (first documentation) and most recent relevant paper in the text here. In parentheses, we indicate the number of papers on the specific topic with n_p, while n_r refers to the total number of results described in the different papers (as some papers describe results of multiple species or life stages). We refer the reader to the references of the reviewed studies provided in the Supplementary Material.

Results

The main findings are described below and summarized in Table 1. A complete overview of the results can be found in the Supplementary

Materials available at the journal's website (Tables S1–S13 in Appendix 1). In addition, we would like to point out that the majority of results described in the available studies use sheep and cattle as hosts (66%), and consequently, 32% of the results on environmental effects on parasitic nematodes are obtained from just three nematode species: *Teladorsagia circumcincta*, *Trichostrongylus colubriformis*, and *Haemonchus contortus* (Figures 2 and 3).

1 What are the effects of temperature and moisture on lifehistory traits of free-living parasitic nematodes?

1.1 Temperature effects on parasitic nematode life-history traits Temperature effects (studied in laboratory) on hatching $(n_p = 8)$, $n_r = 18$, Table S1) (Crofton 1965; Melville et al. 2020), development ($n_p = 14$, $n_r = 29$, Table S2) (Ciordia & Bizell 1963; Gyeltshen et al. 2022), and survival ($n_p = 15$, $n_r = 19$, Table S3) (Gardner et al. 2004; Wang et al. 2022) are generally well known. All three traits have an optimum temperature and a minimum and maximum temperature in which the trait functions. Moreover, the temperature effect is dependent on the trait (van Dijk & Morgan 2008; Gyeltshen et al. 2022), the species (Crofton, 1965; Gyeltshen et al. 2022), larval stage (Peacock et al. 2022), and sometimes substrate (Knapp-Lawitzke et al. 2016; Wang et al. 2022). Thus, conclusions on the effect of temperature cannot be drawn based on one trait or stage only, but it is important to consider the different life-history traits, larvae stages, substrates, and the corresponding temperature ranges, simultaneously. However, a minority of the studies ($n_p = 5$, $n_r = 14$, Table S1–S3) (Ciordia & Bizell 1963; Gyeltshen et al. 2022) cover both the optimum temperature and the upper and lower temperature threshold in which the trait functions.

In addition, some species (e.g., *Nematodirus follicollis* and *Nematodirus battus*) require 'chilling' before hatching (van Dijk & Morgan 2009; Oliver *et al.* 2016) and/or show higher hatching rates after chilling (van Dijk & Morgan 2008). Some parasitic nematode species develop to infective larvae within the eggs before they hatch. Interestingly, within this group of nematodes there are species that need a minimum of 'chilling units' before hatching can occur, as, for example, in *Nematodirus filicollis* (Oliver *et al.* 2016). Chilling experiments aim to study these specific chilling requirements, (e.g., specific chilling temperature and chilling duration). Temperature could also impact developmental paths, for instance,

		Life-history traits	Behaviour
Temperature	Lab	Hatching, development, and survival occur within a range with an optimum (Table S1–3).	There is a temperature range and optimum for L3 larvae migration (Table S10).
		Some species require a chilling treatment before hatching (Table S1).	Temperature did not affect migration out of faeces in a greenhouse setting and non-significantly affected migration into soil and grass (Table S10).
		Temperature could change developmental paths (Table S4).	Light, temperature, and vegetation type might have interacting effects on migration of L3 larvae (Table S10)
		Fluctuating temperatures could yield different outcomes compared to constant temperatures (Table S5).	Temperature is important for thermokinetic behaviour and olfactory responses of skin-penetrating nematodes (Table S11).
	Field	The few studies showed development threshold and positive effect of thermal energy accumulation on L3 larvae recovery from faeces (Table S6).	
Precipitation & moisture	Lab	Positive effects of simulated rainfall & faecal moisture content on development and recovery rate of L3 larvae (Table S7).	There was a minimum and optimum faecal moisture content and required minimum of simulated rainfall for migration out of faeces, with increased migration with increasing rainfall (Table S12).
		Low relative humidity (in salt solutions) resulted in high egg mortality, but drought stress had no significant impact on the recovery of L3 larvae from soil or grass (Table S7).	Simulated rainfall and evaporation affected migration of L3 larvae to deeper soil layers, but drought stress had a non- significant negative effect on L3 larvae migration in soil (Table S12).
			Larvae needed water to migrate out of faeces but not to move into herbage (Table S12).
	Field	The few studies showed variable results: 1) confirmed importance of soil moisture, 2) no long-term effect of wetting of pasture on infection in host, 3) no relation between egg viability and soil texture, moisture, and sun exposure (Table S8).	Lack of movement of eggs on dry soils, and slow and limited movement on wet sandy soils, but limited nr. of studies (Table S13).
Temperature & moisture	Lab	The limited number of studies that tested for interacting effects suggested a lack of interaction between temperature and moisture on life-history traits of parasitic nematodes (Table S9).	The limited number of studies that combined moisture and temperature did not test for interacting effects.

Table 1. The main findings of the reviewed literature on temperature and moisture effects on life-history traits and behaviour of parasitic nematodes of birds and mammals. For more detailed results, we refer the reader to the corresponding text and the supplementary Tables S1–S13 in Appendix 1

to the infective larval stage or adult stage ($n_p = 4$, $n_r = 4$, Table S4) (Viney 1996; Aleuy *et al.* 2019).

Most laboratory studies keep each temperature treatment level constant over time (all above mentioned and Table S1-S4). However, the effects of fluctuating temperatures (which is more representative of the natural environment) on life-history traits yield different, but nonuniversal, outcomes compared to constant temperatures ($n_p = 5$, $n_r = 6$, Table S5) (Saunders *et al.* 2000b; Hernandez et al. 2013) and thus require more attention. Also, only a limited number of studies were conducted in the field with contrasting climatic conditions ($n_p = 2$, $n_r = 3$, Table S6) (Hoar 2012; Hernandez et al. 2013). These studies confirmed a temperature threshold of development of a parasitic nematode in the lower central Canadian Arctic (Hoar 2012) and a positive effect of thermal energy accumulation on the recovery of infective L3 larvae from faeces in Scotland with a temperate climate (Hernandez et al. 2013). Since recovery was not specified in this last study, we assume Hernandez et al. (2013) used recovery as a proxy for the result of hatching, development, and survival combined.

1.2 Precipitation and moisture effects on parasitic nematode lifehistory traits

The effect of moisture on life-history traits under laboratory conditions ($n_p = 10$, $n_r = 29$, Table S7) (Rossanigo & Gruner 1995; Gyeltshen *et al.* 2022) has been less extensively studied compared

with the effect of temperature. Nonetheless, these studies showed positive effects of simulated rainfall (O'Connor et al. 2007; Khadijah, Kahn, Walkden-Brown et al. 2013), faecal moisture content (FMC) /soil moisture (O'Connor et al. 2007; Gyeltshen et al. 2022), and the ratio of precipitation to evaporation rates (O'Connor et al. 2007) on development and recovery rate (here used as proxy for developmental success) of L3 larvae from soil. FMC was also positively correlated with body length of L3 larvae (of four different nematode species; Rossanigo & Gruner, 1996). Larvae recovery was also affected by simulated rainfall timing (Khadijah, Kahn, Walkden-Brown et al. 2013a, 2013b) but in opposite ways according to the distribution (split vs. single) of simulated rain (O'Connor et al. 2007, 2008). Low relative humidity resulted in high mortality of eggs of Nematodirus battus (van Dijk & Morgan 2012), butdrought stress had no significant impact on the recovery of L3 larvae (Cooperia oncophora) from soil or grass (proxy for survival; Knapp-Lawitzke et al. 2016). As with temperature, only limited studies were conducted in the field ($n_p = 3$, $n_r = 4$, Table S8) (Khadijah, Kahn, Walkden-Brown et al. 2013c; May et al. 2022). These field studies showed differing results but were also conducted in the field under different climatic conditions. Soil moisture was identified as important for translation (a term used by the authors to describe the process of development and transition from egg to parasite stage within the host) of parasitic nematodes in sheep in New South Wales (NSW), Australia (Khadijah, Kahn,

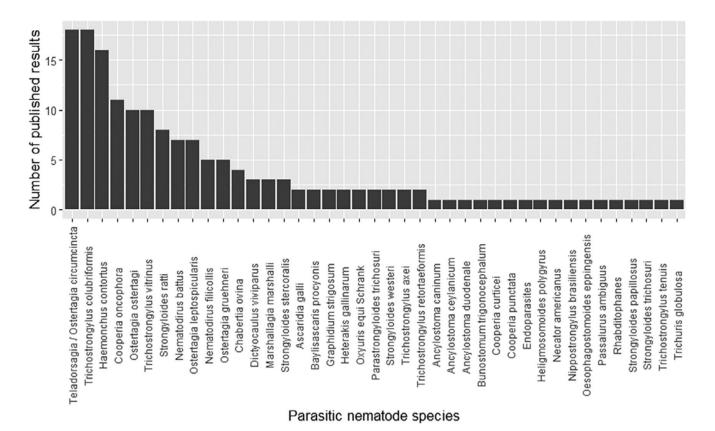


Figure 2. Bar graph of the parasitic nematode species that were used in the reviewed studies on the x-axis. On the y-axis, the number of published results on environmental effects on parasitic nematodes that were obtained from the corresponding parasitic nematode species, ordered from most studied to least studied species.

Walkden-Brown *et al.* 2013c). Nonetheless, wetting of pasture did not have long-term effects on infection of endoparasites (including nematodes) in sheep and cattle (measured by larvae and egg count in faeces) in Germany (May *et al.* 2022). In addition, a field study in Texas showed that viability of eggs (of *Baylisascaris procyonis* that can have both direct and indirect life cycle with racoon as host) was not related to soil texture, moisture, and sun exposure (Ogdee *et al.* 2016).

1.3 Interacting temperature and moisture effects on parasitic nematode life-history traits

Only two studies ($n_p = 2$, $n_r = 5$, Table S9) (Rossanigo & Gruner 1995; McCarthy et al. 2022) tested for an explicit interaction effect between temperature and moisture, and neither found one. McCarthy et al. (2022) found that lungworm (Dictyocaulus viviparus) L3 larvae mortality was similar in paired wet and dry conditions measured at three different temperatures; so, by inference, the effect of moisture on L3 larvae mortality did not depend on temperature. A study of strongyles in sheep, cattle, and deer found that the optimal faecal moisture content for development was independent of temperature (Rossanigo & Gruner 1995). Thus, a limited number of studies report a lack of interacting abiotic factors on life-history traits of parasitic nematodes. Three other studies focused on the combination or additive effects of abiotic factors on life-history traits without testing for possible interactions (Table S2, S3, S7; Rossanigo & Gruner 1996; Knapp-Lawitzke et al. 2016; Gyeltshen et al. 2022). For instance, an extensive laboratory study on the effect of temperature and faecal water content (FWC) on development averaged FWC across the different temperatures, instead of considering the interaction between FWC and temperature, and merely suggested that temperature negatively affected FWC (Gyeltshen et al. 2022). Rossanigo & Gruner (1996) found that L3 larvae body size increased with faecal moisture content (FMC) in all three temperature regimes in four species of strongyles, but apparent interacting effects between temperature and moisture (visual assessment of their Figure 1) were not formally tested. Finally, a study of temperature (spring versus summer scenarios), drought (adequate water versus none), and exposure duration on the overall recovery of L3 larvae (proxy for survival) showed negative effects of temperature (on recovery from soil) and exposure duration (on recovery from grass), but possible interactions were not tested (Knapp-Lawitzke et al. 2016). Thus, more information on interacting factors might exist in the present literature, and this data could possibly be acquired to analyze interacting factors specifically.

2 What are the effects of temperature and moisture on the behaviour of free-living parasitic nematodes?

2.1 Temperature effects on parasitic nematode behaviour

The effect of temperature on parasite behaviour and movement (out of faeces and in soil) is not conclusive ($n_p = 6$, $n_r = 8$, Table S10) (Saunders *et al.* 2000a; Gyeltshen *et al.* 2022). For example, some laboratory studies showed migration declined over time (days) stored at a constant high temperature of 30°C (Hamilton *et al.* 2022) and an optimum temperature for L3 larvae migration out of faeces (Gyeltshen *et al.* 2022). However, temperature did not affect migration out of faeces in a greenhouse setting (Wang *et al.* 2018) and had no significant effect (but a positive trend) on vertical

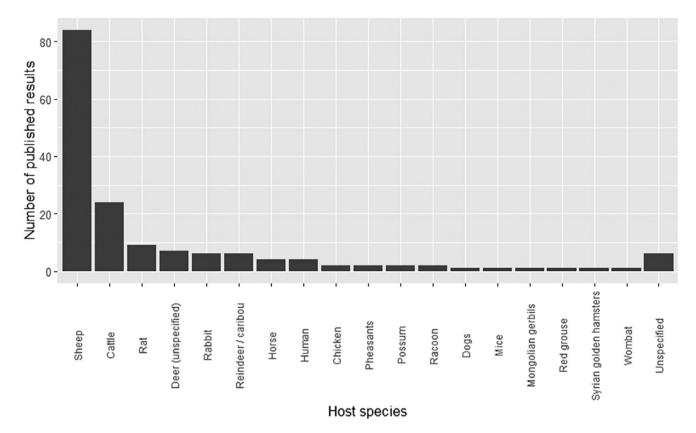


Figure 3. Bar graph of the host species that were used in the reviewed studies on the x-axis. On the y-axis, the number of published results on environmental effects on parasitic nematodes that were obtained from using the corresponding host species, ordered from most studied to least studied species.

migration of L3 larvae in soil (Knapp-Lawitzke *et al.* 2016). Moreover, the effect of temperature on parasite behaviour is complex, as temperature might interact with light and vegetation type (Saunders *et al.* 2000a). Studies under laboratory conditions conclusively indicated that temperature is important for thermokinetic behaviour and olfactory responses of skin-penetrating nematodes ($n_p = 10$, $n_r = 15$, Table S11) (Stankiewicz 1996; Dulovic *et al.* 2022; and for a more extensive review, see Bryant & Hallem 2018; Mendez *et al.* 2022; Takeishi 2022). We could not find field studies on the effect of temperature on parasitic nematode behaviour.

2.2 Precipitation and moisture effects on parasitic nematode behaviour

Studies under laboratory or greenhouse conditions $(n_p =$ 7, n_r = 19, Table S12) (Rossanigo & Gruner 1996; Gyeltshen et al. 2022) showed that water is required for infective larvae to migrate out of faeces (van Dijk & Morgan 2011; Wang et al. 2014), with both a minimum and optimum faecal water content (FWC) value for migration of infective larvae out of faeces (Gyeltshen et al. 2022). Similarly, there was increased migration under increasing simulated rainfall, again with a required minimum for migration to occur (Wang et al. 2022). In contrast to migration out of a faecal pat, it has been suggested that parasitic nematodes do not need water to move from the soil surface into the herbage (van Dijk & Morgan 2011). Simulated rainfall and evaporation also affected migration of L3 larvae to deeper soil layers. Recovery of L3 larvae in deeper soil layers increased with lower evaporation rates, with increasing simulated rainfall, and over time (O'Connor et al. 2008). In contrast, drought stress had no significant effect (but a negative trend) on L3 larvae migration from grass to soil (Knapp-Lawitzke *et al.* 2016). Moreover, it was suggested that migration can be mediated by individual differences in body length. Short L3 larvae (collected from dry faeces) migrated more slowly than long L3 larvae collected from wet faeces (Rossanigo & Gruner 1996). There is only one field study on the effect of rain and moisture on the behaviour of parasitic nematodes in Texas ($n_p = 1$, $n_r = 1$, Table S13) (Ogdee *et al.* 2016). This study indicated no (passive) movement of nematode eggs on dry soils and minor movement on wet soils (dependent on the soil type but remained within the top 10 cm in the soil column) over the course of two years (Ogdee *et al.* 2016).

2.3 Interacting temperature and moisture effects on parasitic nematode behaviour

None of the reviewed studies explicitly tested for an interaction effect of temperature and moisture on parasitic nematode behaviour. In the discussion we will elaborate on three studies (also described in paragraph 1.3) that included a combined effect of moisture and temperature and thus had a study design capable of testing for possible interacting effects, but which were not explicitly done (Table S10, S12; Rossanigo & Gruner 1996; Knapp-Lawitzke *et al.* 2016; Gyeltshen *et al.* 2022).

Discussion

In this review, we identified the knowledge gaps in research on abiotic effects on terrestrial parasitic nematodes in birds and mammals (Figure 4). While much is known about the separate effects of temperature and moisture on parasite life history (Figure 4, lines 1a, 1b), knowledge about the interactive effects

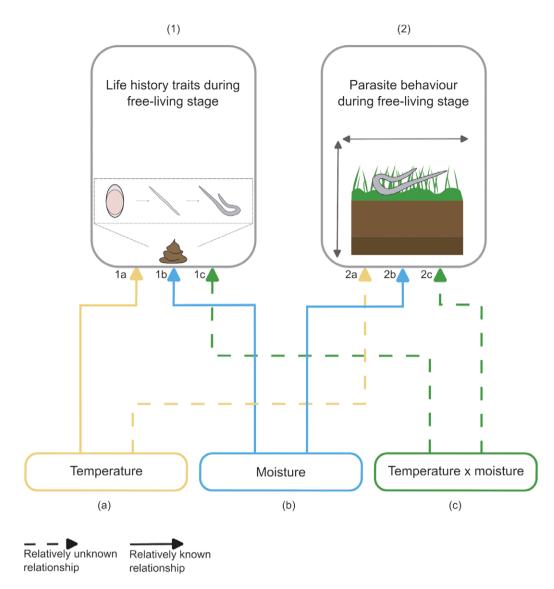


Figure 4. Visualization of the relatively known (solid arrows) and relatively unknown (dashed arrows) effects of temperature (yellow, a), moisture (blue, b), and temperature and moisture combined (green, c) on 1) life-history traits and 2) behaviour of parasitic nematodes of birds and mammals during the free-living stages.

of both temperature and moisture on parasitic nematode lifehistory traits is scarce (Figure 4, line 1c) even though this has long been recognised as important (see O' Connor et al. 2006). We highlight this as the first major knowledge gap, especially given recent climate change (IPCC 2021). Although positive moisture effects have been documented on parasite motility (Figure 4, line 2b), there is little known about temperature effects (Figure 4, line 2a) or the interaction of moisture and temperature (Figure 4, line 2c). This is the second major gap and is important because these interactions could influence transmission into the host (Silva et al. 2008). Also, we advocate for more experimental field studies that test the effects of multiple environmental factors on life-history traits and behaviour under more natural and realistic circumstances. Moreover, we encourage studies to expand the range of used hosts and parasitic nematodes. Future studies targeting these knowledge gaps should improve our understanding of how environmental change will affect parasitic nematodes and, consequently, influence parasite-host dynamics. Such research has broader implications for identifying infection risks,

which is highly relevant to livestock managers and wildlife conservationists, and for an improved understanding of ecosystem functioning in a rapidly warming climate.

1 Parasitic nematode life-history traits

Although temperature effects on hatching, development, and survival are generally well known (Figure 4, line 1a), we wish to encourage future research to focus on three issues. First, additional studies should calculate complete temperature dependency curves for all the different vital rates of a variety of species (see, for example, van Dijk & Morgan 2009). Insight into the full range of temperature dependency curves could be used for projecting parasite–host dynamics under changing climates with mathematical models (Fox *et al.* 2012; Molnár *et al.* 2017). Determining temperature dependency curves of different life-history rates will also allow the search for general patterns within and across species (Vineer *et al.* 2016; Phillips *et al.* 2022). Discovering general patterns (by using meta-analysis) would provide insight into the reliability

of translating findings to different study systems. Second, future studies should investigate how fluctuating temperatures, more representative of the increased variance observed under climate warming (IPCC 2021), affect life-history traits – therefore better representing natural circumstances. Third, studies should test which species require pre-hatch chilling and how a warming climate will affect and possibly reduce hatching rate of species that require chilling. For example, warming could delay accumulation of 'chilling units', which could delay hatching by one year (Oliver *et al.* 2016), with possible consequences for infectivity patterns. Thus, although temperature effects on life-history traits are generally well known, specific knowledge is still needed.

Most of the studies we reviewed that tested effects of moisture focused on the overall recovery of L3 larvae under different moisture conditions (Figure 4, line 1b). However, measuring total recovery of L3 larvae from faeces or soil is sometimes used as a proxy for developmental success (O'Connor et al. 2007) or survival (Knapp-Lawitzke et al. 2016), or is not specified (Hernandez et al. 2013). Nonetheless, true recovery is composed of hatching, development, larval survival, and possibly motility. This means that the effects of rain and moisture on the separate rates of hatching, development, and survival are still poorly known. To disentangle this, (laboratory) experiments are needed that sample frequently and differentiate between the different life stages and between dead and live larvae to separate survival and migration processes (for instance, by using extracting methods that are not based on motility and counting living and dead individuals). Studying the moisture response curve for separate instead of composite processes would also improve predictive models as advocated by Molnár and colleagues for thermal response curves (Molnár et al. 2017).

The effect of interacting environmental factors on parasitic nematodes is identified as an important knowledge gap because it is more representative of variation in the natural environment than the effect of the separate abiotic factors in isolation (Figure 4, line 1c). In addition, under the current climate change projections, more extreme weather events are expected (IPCC 2021), and therefore, experiments that test interacting effects of moisture and temperature across large gradients, even beyond the current norm, are increasingly valuable. Only two studies formally investigated the possible interaction between temperature and moisture and indicated an absence of interactive effect of temperature and moisture on parasite life history (Rossanigo & Gruner 1995; McCarthy et al. 2022). Among three other studies which focused on additive effects of abiotic factors on life-history traits without testing for possible interactions (e.g., Rossanigo & Gruner 1996; Knapp-Lawitzke et al. 2016; Gyeltshen et al. 2022), one presented a figure suggesting apparent interactions (Figure 1 in Rossanigo & Gruner 1996). When reanalysing the data provided in their Table 2, there is a statistically significant interaction between moisture and temperature on L3 larvae body size in all four species studied: the temperature regime leading to the largest larvae differed as faecal moisture content varied from 30 to 80%, reflecting that optimal development temperatures are modulated by moisture (Supplementary Materials – Appendix 2). Testing for interactions among multiple environmental factors (Rossanigo & Gruner 1995; Tarbiat et al. 2018; Shifaw et al. 2022) will advance our understanding of parasite-host interactions under increasingly variable conditions.

2 Parasitic nematode behaviour

Parasite motility influences contact rates between parasite and host, and hence potentially transmission rates. Also, parasite motility could modulate the effect of environmental change on life-history traits, by, for instance, the ability to 'escape' adverse environments. Nevertheless, there is limited knowledge on how temperature alone, or in combination with moisture, affect behaviour, especially the vertical migration of parasitic nematodes (Figure 4, lines 2a, c). Therefore, we advocate studies on temperature, moisture, and their combined effects on parasite behaviour and highlight two specific angles. First, how does temperature (Figure 4, line 2a) and soil moisture (Figure 4, line 2b), and their interaction (Figure 4, line 2c), affect vertical migration of parasitic nematodes in vegetation and in soil, especially under natural conditions? For instance, an entomopathogenic nematode (infecting insects) that poorly tolerates desiccation migrated towards the bottom soil layer with decreasing moisture content, while another species of nematode that is desiccation tolerant remained in the top layer (Salame & Glazer 2015). Whether parasitic nematodes of birds and mammals show similar migration behaviour is unknown. In addition, parasitic nematodes face a trade-off between avoiding adverse environmental conditions and opportunities enhancing the probability of being ingested by the host. For example, L3 larvae recovered from soil were considered fitter (measured with migration assay) than the individuals recovered from the grass (Knapp-Lawitzke et al. 2016). Migration behaviour might also vary between individuals with, for instance, longer nematodes (grown at optimal conditions), migrating more than shorter individuals and possibly increasing infection probability (Rossanigo & Gruner 1996). In addition, insight is needed on how parasitic nematodes are distributed among different vegetation types and possible interactions with abiotic factors (Saunders et al. 2000a). These insights could be provided by experiments that test vertical migration under different environmental factors on different vegetation types. Second, how do temperature and moisture levels affect the distance of horizontal dispersal (Figure 4, lines 2a, b, c), and do parasitic nematodes predominantly disperse actively or passively (for instance, rain splashes might be a major factor for translocation; Grønvold & Høgh-Schmidt 1989)? We advocate experiments that test dispersal range away from faeces under various environmental factors, including extreme temperature and moisture levels. Although rainfall is required for migration out of faeces (van Dijk & Morgan 2011; Wang et al. 2014), heavy precipitation could also have negative effects. For instance, irrigation of the pasture facilitated migration of larvae, but mortality was high, and fewer larvae were recovered from the herbage on irrigated plots compared to dry plots, possibly due to mortality or dispersal to wider areas (Young & Anderson 1981). Studying horizontal dispersal distances is especially relevant in relation to faecal avoidance by hosts (Hutchings et al. 1998; van der Wal et al. 2000). Understanding the factors that affect both vertical and horizontal movement and linking this to the contact place with the host will help in understanding transmission patterns and infection risk of the host. In addition, the availability of vegetation (in relation to environmental conditions) is potentially an important factor in understanding the link between contact rates with the host and transmission. For example, vegetation growth (due to high levels of rainfall) could lead to a dilution effect with a decreased density of nematodes in the environment and possibly result in fewer encounters with parasites by the host (Grenfell 1988; Grenfell 1992; Shearer & Ezenwa 2020).

3 Translation from laboratory to field conditions

We advocate more field studies, in addition to laboratory studies, because microclimate differences and soil structure might alter the relationships between environmental factors and parasites only documented under laboratory conditions. For example, solar radiation might cause mortality at temperatures lower than the threshold temperature determined in the laboratory (van Dijk et al. 2009), and snow layers could benefit parasite survival (Rossi et al. 2019). Moreover, air temperature might be an insufficient proxy for temperature conditions near or in soil (Lembrechts et al. 2022), especially during times with snow cover (Convey et al. 2015). Surface or soil temperature might be more important and could have different effects on life-history traits of nematodes (Jenkins et al. 2006; Kutz et al. 2014). Also, in the field, biotic interactions may affect parasite life history and behaviour in several ways. For example, the presence of earthworms can reduce the number of recovered larvae in the field (Waghorn et al. 2011). The presence of nemathophagous fungus was associated with a reduction in the number of infective parasite larvae (Kuzmina et al. 2006), but fungus can also play a role in and facilitate migration of parasitic larvae (McCarthy et al. 2022). Moreover, in some species, contact between bacteria and embryonated eggs is required for hatching within the host (Hayes et al. 2010). These biotic interactions add complexity beyond simple relationships between weather variables and parasite life-history traits and behaviour. Thus, field experiments and laboratory experiments are complementary. Lab experiments will continue to provide mechanistic links between the environment and parasite traits, but they need to be scrutinized in real-world settings to investigate if relationships hold. By doing so, we can more reliably draw conclusions on how changing climates affect parasites and hence their interaction with hosts.

4 Expanding the range of parasitic nematode and host species studies

In addition, we would like to stress that the majority of results described in the available studies use sheep and cattle as hosts (66%), and 32% of results describing environmental effects on parasitic nematodes are obtained from just the three nematode species: Teladorsagia circumcincta, Trichostrongylus colubriformis, and Haemonchus contortus (Figures 2 and 3). These species are the most abundant gastrointestinal nematodes in sheep and cause major production losses (O'Connor et al. 2006). However, we envisage that the nematode-host interaction could be different for livestock and free-living hosts. For example, wild animals generally live in lower population densities and do not receive (anthelmintic) treatments or supplemental food. Also, parasitic nematodes of livestock hosts might encounter different selection pressures due to rotational grazing or other treatments, which might result in differences in environmental effects on parasitic nematodes of wild compared to livestock animals. Because there is limited data on wildlife hosts and their parasitic nematodes, we encourage future studies on the parasitic nematodes of wild host species under natural conditions. Moreover, few of the reviewed studies involve the environmental effects on parasitic nematodes infecting humans. However, infection of soil-transmitted helminths in humans is a major cause of human disease (Brooker et al. 2006; Weaver et al. 2010), especially in places where sanitation facilities are limited but population densities are high (Brooker et al. 2006; Weaver et al. 2010). To limit infection by soil-transmitted helminths, insights are needed on how these parasites are affected by environmental changes (Weaver *et al.* 2010).

5 Link to life stages within the host and parasite host dynamics

The scope of this review was to identify the knowledge gaps in research about the abiotic factors impacting hatching, development, survival, and behaviour of direct life-cycle parasitic nematodes in their free-living stages. However, we have not considered how environmental factors experienced in these free-living stages might affect parasites once they are within the host - for example, when they need to exsheath, develop, survive, and reproduce (Figure 1) (Kutz et al. 2009; Morley & Lewis 2014; Cizauskas et al. 2017). Also, environmental factors can affect parasitic nematodes during the stages within the host through effects on homeostasis of the host, for instance (see for a review, but not specifically on parasitic nematodes, Morley & Lewis 2014). It is also suggested that rainfall could affect parasite infection levels through changes in host susceptibility, with low rainfall levels (compared to high rainfall levels) increasing susceptibility of the host to parasite infection (Shearer & Ezenwa 2020). This could possibly be explained by reduced food availability and quality under low rainfall conditions, and hence changes and reduction in body condition of the host, but this mechanism needs to be tested (Shearer & Ezenwa 2020). Also, we need to consider the abilities for adaptation to changing conditions by the host and by the parasites (reviewed by Aleuy & Kutz 2020) to get a full comprehension of the environmental effects on parasitic nematode.

6 Implications

In this review, we have focused attention on understudied abiotic impacts that could improve our insights into how environmental factors affect parasitic nematodes in their free-living stages, and hence the likely impact of climate change. Addressing these knowledge gaps will also have implications for identifying infection risks, of high relevance to livestock managers and wildlife conservationists, as well as improve our understanding of ecosystem functioning under a rapidly warming climate. First, experiments on the temperature and moisture (and their interaction) dependency curves of parasitic lifehistory traits and behaviour could better guide livestock managers about parasite dynamics and epidemiology (considered as an important knowledge gap according to the research community in livestock helminthology; Morgan et al. 2019) and infection risks under changing climates. For instance, improved knowledge on how abiotic factors influence parasite life history and behaviour can help improve models that project infection pressure under various climate change scenarios (Rose et al. 2016). Moreover, data from these experiments could refine infection risk maps and calendars estimating when and where infection risks are high (Navarre 2020; McFarland et al. 2022) and provide insights about pasture management strategies (Morgan et al. 2019, Navarre 2020), such as fencing off risk areas and drainage (Beltrame et al. 2021). For instance, experiments on the effect of temperature and moisture (and their interaction) on the vertical and horizontal distribution of parasitic nematodes could provide information on infection hotspots in the herbage and might guide management of forage heights (Navarre 2020). Second, knowledge (obtained by additional experiments on temperature and moisture effects on parasites) could provide insights benefiting wildlife conservation, such as disease risk frameworks and optimal timing of intervention, particularly in areas with a co-existence of livestock and

wildlife (Khanyari et al. 2022). Moreover, the presence of parasites might be, in some cases, beneficial to population health (Thompson et al. 2010). For instance, parasites could possibly act on delayed density dependent mechanisms, which could dampen host population fluctuations and hinder population crashes resulting from rapid increases. Thus, understanding environmental effects on parasites and if and how parasite species should be conserved (Thomas et al. 2005) will have indirect benefits for wildlife and ecosystem conservation. Third, knowledge about environmental effects on parasitic nematodes (based on experiments that test the effect of a broad range of moisture and temperature regimes on life-history traits and behaviour of a wide range of parasites) might improve the understanding of ecosystem functioning. Parasites play a role in ecosystem engineering processes, either by modifying hosts that are ecosystem engineers or as engineers themselves (Thomas et al. 1999). They can also impact intraspecific species interaction (Hudson et al. 2006) such as predator-prey interactions (Hudson et al. 1992; Hatcher et al. 2006) and hence energy flow through the system (Hudson et al. 2006). Understanding which functions in the ecosystem will be lost if a parasite species disappears might result in the realization that parasite species need to be conserved in order to maintain an ecosystem and its functions (Thomas et al. 2005). Parasites could also cause cascading effects on other (trophic) species in the ecosystem (Thomas et al. 2005). For instance, it was suggested that lethal and sub-lethal effects of a parasitic nematode on an herbivorous host could result in trophic cascades (in terms of lower herbivore host biomass and increased producer biomass) (Koltz et al. 2022). Thus, changes in the environmental effects on parasitic nematodes might have larger consequences on the ecosystem than just on the parasite and host. In conclusion, addressing and hence filling the knowledge gaps about abiotic impacts on parasitic nematodes could have broad implications for livestock managers and wildlife conservationists and improve our fundamental understanding of parasitic nematodes and ecosystem functioning under a rapidly warming climate.

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References

- Aleuy OA, Hoberg EP, Paquette C, Ruckstuhl KE, and Kutz S (2019) Adaptations and phenotypic plasticity in developmental traits of Marshallagia marshalli. International Journal for Parasitology 49, 789–796.
- Aleuy OA and Kutz S (2020) Adaptations, life-history traits and ecological mechanisms of parasites to survive extremes and environmental unpredictability in the face of climate change. *International Journal for Parasitology: Parasites and Wildlife* 12, 308–317.
- Altizer S, Dobson A, Hosseini P, Hudson P, and Pascual M (2006) Seasonality and the dynamics of infectious diseases. *Ecology Letters* 9, 467–484.
- Beltrame L, Rose Vineer H, Walker JG, Morgan, ER, Vickerman P, and Wagener T (2021) Discovering environmental management opportunities

for infectious disease control. *Scientific Reports* **11**. https://doi.org/10.1038/ s41598-021-85250-1

- Brooker S, Clements AC, and Bundy DA (2006) Global epidemiology, ecology and control of soil-transmitted helminth infections. *Advances in parasitology* 62, 221–261.
- Bryant AS and Hallem EA (2018) Temperature-dependent behaviors of parasitic helminths. *Neuroscience Letters* 687, 290–303.
- Charlier J, Höglund J, von Samson-Himmelstjerna G, Dorny P, and Vercruysse J (2009) Gastrointestinal nematode infections in adult dairy cattle: impact on production, diagnosis and control. *Veterinary Parasitology* 164, 70–79.
- Chaudary FR, Qayyum M, and Miller JE (2008) Development and survival of *Haemonchus contortus* infective larvae derived from sheep faeces under subtropical conditions in the Potohar region of Pakistan. *Tropical Animal Health and Production* 40, 85–92.
- Chung JA and Boeri F (2012) Nematodes: morphology, functions and management strategies. New York: Nova Science Publishers, Incorporated.
- **Ciordia H and Bizell WE** (1963) The effects of various constant temperatures on the development of the free living- stages of some nematode parasites of cattle. *The Journal of Parasitology* **49**, 60–63.
- Cizauskas CA, Carlson CJ, Burgio R, Clements CF, Dougherty ER, Harris NC, and Phillips AJ (2017) Parasite vulnerability to climate change: an evidence-based functional trait approach. *Royal Society Open Science* 4. https://doi.org/10.1098/rsos.160535
- Convey P, Abbandonato H, Bergan F, Beumer LT, Biersma EM, Bråthen VS, D'Imperio L, Jensen CK, Nilsen S, Paquin K, Stenkewitz U, Svoen ME, Winkler J, Müller E, and Coulson SJ (2015) Survival of rapidly fluctuating natural low winter temperatures by High Arctic soil invertebrates. *Journal of Thermal Biology* 54, 111–117.
- **Crofton HD** (1965) Ecology and biological plasticity of sheep nematodes. I. The effect of temperature on the hatching of eggs of some nematode parasites of sheep. *Cornell Veterinarian* **55**, 242–250.
- **Dobson A, Molnár PK, and Kutz S** (2015) Climate change and Arctic parasites. *Trends in Parasitology* **31**, 181–188.
- Dulovic A, Norman M, Harbecke D, and Streit A (2022) Chemotactic and temperature-dependent responses of the *Strongyloidoidea* superfamily of nematodes. *Parasitology* 149, 116–123.
- Fox NJ, Marion G, Davidson RS, White PCL, and Hutchings MR (2012) Livestock helminths in a changing climate: approaches and restrictions to meaningful predictions. *Animals* 2, 93–107.
- Gardner MP, Gems D, and Viney ME (2004) Aging in a very short-lived nematode. Experimental Gerontology 39, 1267–1276.
- Grenfell BT (1988) Gastrointestinal nematode parasites and the stability and productivity of intensive ruminant grazing systems. Philosophical Transactions of the Royal Society of London. B, Biological Sciences 321, 541–563.
- Grenfell BT (1992) Parasitism and the dynamics of ungulate grazing systems. The American Naturalist 139, 907–929.
- **Grønvold J and Høgh-Schmidt K** (1989) Factors influencing rain splash dispersal of infective larvae of *Ostertagia ostertagi (Trichostrongylidae)* from cow pats to the surroundings. *Veterinary Parasitology*, 57–70.
- Gyeltshen T, Kahn LP, and Laurenson YCSM (2022) Ecology of the free-living stages of *Trichostrongylid* parasites of sheep. *Veterinary Parasitology* 303. https://doi.org/10.1016/j.vetpar.2022.109683
- Halvorsen O (2012) Reindeer parasites, weather and warming of the Arctic. *Polar Biology* **35**, 1749–1752.
- Hamilton KM, Waghorn TS, de Waal T, Keane OM, Green P, and Leathwick DM (2022) In vitro evaluation of fitness parameters for isolates of *Telador-sagia circumcincta* resistant and susceptible to multiple anthelmintic classes. *Veterinary Parasitology* **310**. https://doi.org/10.1016/j.vetpar.2022.109791
- Hatcher MJ, Dick JTA, and Dunn AM (2006) How parasites affect interactions between competitors and predators. *Ecology Letters* 9, 1253–1271.
- Hayes KS, Bancroft AJ, Goldrick M, Portsmouth C, Roberts IS, and Grencis RK (2010) Exploitation of the intestinal microflora by the parasitic nematode *Trichuris muris. Science* **328**, 1391–1394.
- Hernandez AD, Poole A, and Cattadori IM (2013) Climate changes influence free-living stages of soil-transmitted parasites of European rabbits. *Global Change Biology* 19, 1028–1042.

- Hoar B (2012) Ecology and transmission dynamics of Ostertagia gruehneri in Barrenground Caribou. Calgary, AB: University of Calgary.
- Holand H, Jensen H, Kvalnes T, Tufto J, Pärn H, Sæther BE, and Ringsby TH (2019) Parasite prevalence increases with temperature in an avian metapopulation in northern Norway. *Parasitology* 146, 1030–1035.
- Hudson PJ, Cattadori IM, Boag B, and Dobson AP (2006) Climate disruption and parasite–host dynamics: patterns and processes associated with warming and the frequency of extreme climatic events. *Journal of Helminthology* 80, 175–182.
- Hudson PJ, Dobson AP, and Newborn D (1992) Do parasites make prey vulnerable to predation? Red grouse and parasites. *Journal of Animal Ecology* 61, 681–692.
- Hutchings MR, Kyriazakis I, Anderson DH, Gordon IJ, and Coop RL (1998) Behavioural strategies used by parasitized and non-parasitized sheep to avoid ingestion of gastro-intestinal nematodes associated with faeces. *Animal Science* **67**, 97–106.
- IPCC. (2021) Summary for policymakers. In Masson-Delmotte VP, Zhai P, Pirani A, Connors BZSL, Péan C, Berger S, Caud N, Chen Y, Goldfarb L, Gomis MI, Huang M, Leitzell K, Lonnoy E, Matthews JBR, Maycock TK, Waterfield T, Yelekçi O, Yu R (Eds), Climate change 2021: The physical science basis. Contribution of working group I to the sixth assessment report of the intergovernmental panel on climate change.
- Jenkins EJ, Veitch AM, Kutz SJ, Hoberg EP, and Polley L (2006) Climate change and the epidemiology of *protostrongylid* nematodes in northern ecosystems: *Parelaphostrongylus odocoilei* and *Protostrongylus stilesi* in Dall's sheep (*Ovis d. dalli*). *Parasitology* 132, 387–401.
- Khadijah S, Kahn LP, Walkden-Brown SW, Bailey JN, and Bowers SF (2013) Soil moisture influences the development of *Haemonchus contortus* and *Trichostrongylus colubriformis* to third stage larvae. *Veterinary Parasitology* 196, 161–171.
- Khadijah S, Kahn LP, Walkden-Brown SW, Bailey JN, and Bowers SF (2013a) Effect of simulated rainfall timing on faecal moisture and development of *Haemonchus contortus* and *Trichostrongylus colubriformis* eggs to infective larvae. Veterinary Parasitology 192, 199–210.
- Khadijah S, Kahn LP, Walkden-Brown SW, Bailey JN, and Bowers SF (2013b) Soil moisture modulates the effects of the timing and amount of rainfall on faecal moisture and development of *Haemonchus contortus* and *Trichostrongylus colubriformis* to infective third stage larvae. Veterinary Parasitology 196, 347–357.
- Khadijah S, Kahn LP, Walkden-Brown SW, Bailey JN, and Bowers SF (2013c) Translation of *Haemonchus contortus* and *Trichostrongylus colubriformis* from egg to establishment in grazing sheep is unaffected by rainfall timing, rainfall amount and herbage height under conditions of high soil moisture in the Northern Tablelands of NSW. Veterinary Parasitology 197, 204–211.
- Khanyari M, Milner-Gulland EJ, Oyanedel R, Vineer HR, Singh NJ, Robinson S, Salemgareyev A, and Morgan ER (2022) Investigating parasite dynamics of migratory ungulates for sustaining healthy populations: application to critically endangered saiga antelopes Saiga tatarica. Biological Conservation 266. https://doi.org/10.1016/j.biocon.2022.109465
- Knapp-Lawitzke F, von Samson-Himmelstjerna G, and Demeler J (2016) Elevated temperatures and long drought periods have a negative impact on survival and fitness of *strongylid* third stage larvae. *International Journal for Parasitology* 46, 229–237.
- Koltz AM, Civitello DJ, Becker DJ, Deem SL, Classen AT, Barton B, ... and Ezenwa VO (2022) Sublethal effects of parasitism on ruminants can have cascading consequences for ecosystems. *Proceedings of the National Academy* of Sciences 119, e2117381119.
- Kutz SJ, Hoberg EP, Molnár PK, Dobson A, and Verocai GG (2014) A walk on the tundra: Host-parasite interactions in an extreme environment. *International Journal for Parasitology: Parasites and Wildlife* 3, 198–208.
- Kutz SJ, Jenkins EJ, Veitch AM, Ducrocq J, Polley L, Elkin B, and Lair S (2009) The Arctic as a model for anticipating, preventing, and mitigating climate change impacts on host-parasite interactions. *Veterinary Parasitology* 163, 217–228.
- Kuzmina TA, Kuzmin YI, and Kharchenko VA (2006) Field study on the survival, migration and overwintering of infective larvae of horse strongyles on pasture in central Ukraine. *Veterinary Parasitology* 141, 264–272.

- Lafferty KD, Dobson AP, and Kuris AM (2006) Parasites dominate food web links. Proceedings of the National Academy of Sciences of the United States of America 103, 11211–11216.
- Langrová I, Jankovská I, Borovský M, and Fiala T (2003). Effect of climatic influences on the migrations of infective larvae of *Cyathostominae*. Veterinarni Medicina 48, 18–24.
- Lembrechts JJ, Hoogen J, Aalto J, Ashcroft MB, de Frenne P, Kemppinen J, Kopecký M, Luoto M, Maclean IMD, Crowther TW, Bailey JJ, Haesen S, Klinges DH, Niittynen P, Scheffers BR, van Meerbeek K, Aartsma P, Abdalaze O, Abedi M, ... Lenoir J (2022) Global maps of soil temperature. *Global Change Biology* 28, 3110–3144.
- May K, Raue K, Blazejak K, Jordan D, and Strube C (2022) Pasture rewetting in the context of nature conservation shows no long-term impact on endoparasite infections in sheep and cattle. *Parasites and Vectors* **15**. https://doi. org/10.1186/s13071-022-05155-4
- McCarthy C, Vineer HR, Morgan ER, and van Dijk J (2022) Predicting the unpredictable? A climate-based model of the timing of peak pasture infectivity for *Dictyocaulus viviparus*. Veterinary Parasitology **309**. https://doi. org/10.1016/j.vetpar.2022.109770
- McFarland C, Rose Vineer H, Chesney L, Henry N, Brown C, Airs P, Nicholson C, Scollan N, Lively F, Kyriazakis I, and Morgan ER (2022) Tracking gastrointestinal nematode risk on cattle farms through pasture contamination mapping. *International Journal for Parasitology* 52, 691–703.
- Mehlhorn H (2016) Worms (Helminths). pp. 251–498 in Mehlhorn H (Eds), Animal parasites. Cham: Springer International Publishing.
- Melville LA, van Dijk J, Mitchell S, Innocent G, and Bartley DJ (2020) Variation in hatching responses of *Nematodirus battus* eggs to temperature experiences. *Parasites & Vectors* 13. https://doi.org/10.1186/s13071-020-04368-9
- Mendez P, Walsh B, and Hallem EA (2022) Using newly optimized genetic tools to probe Strongyloides sensory behaviors. Molecular and Biochemical Parasitology 250. https://doi.org/10.1016/j.molbiopara.2022.111491
- Molnár PK, Dobson AP, and Kutz SJ (2013) Gimme shelter the relative sensitivity of parasitic nematodes with direct and indirect life cycles to climate change. *Global Change Biology* 19, 3291–3305.
- Molnár PK, Kutz SJ, Hoar BM, and Dobson AP (2013) Metabolic approaches to understanding climate change impacts on seasonal host-macroparasite dynamics. *Ecology Letters* 16, 9–21.
- Molnár PK, Sckrabulis JP, Altman KA, and Raffel TR (2017) Thermal performance curves and the metabolic theory of ecology – a practical guide to models and experiments for parasitologists. *The Journal of Parasitology* 103, 423–439.
- Morgan ER, Aziz NAA, Blanchard A, Charlier J, Charvet C, Claerebout E, Geldhof P, Greer AW, Hertzberg H, Hodgkinson J, Höglund J, Hoste H, Kaplan RM, Martínez-Valladares M, Mitchell S, Ploeger HW, Rinaldi L, von Samson-Himmelstjerna G, Sotiraki S, ... Vercruysse J (2019) 100 questions in livestock helminthology research. *Trends in Parasitology* 35, 52–71.
- Morley NJ and Lewis JW (2014) Temperature stress and parasitism of endothermic hosts under climate change. *Trends in Parasitology* 30, 221–227.
- Navarre CB (2020) Epidemiology and control of gastrointestinal nematodes of cattle in southern climates. Veterinary Clinics of North America – Food Animal Practice 36, 45–57.
- O' Connor LJ, Walkden-Brown SW, and Kahn LP (2006) Ecology of the freeliving stages of major *trichostrongylid* parasites of sheep. *Veterinary Parasitology* 142, 1–15.
- O'Connor LJ, Kahn LP, and Walkden-Brown SW (2007) Moisture requirements for the free-living development of *Haemonchus contortus*: quantitative and temporal effects under conditions of low evaporation. *Veterinary Parasitology* **150**, 128–138.
- O'Connor LJ, Kahn LP, and Walkden-Brown SW (2008) Interaction between the effects of evaporation rate and amount of simulated rainfall on development of the free-living stages of *Haemonchus contortus*. *Veterinary Parasitology* **155**, 223–234.
- Ogdee JL, Henke SE, Wester DB, and Fedynich AM (2016) Permeability and viability of *Baylisascaris procyonis* eggs in Southern Texas soils. *Journal of Parasitology* **102**, 608–612.

- Oliver AB, Pomroy WE, Ganesh S, and Leathwick DM (2016) Chilling requirements for hatching of a New Zealand isolate of *Nematodirus filicollis*. *Veterinary Parasitology* **226**, 17–21.
- Peacock SJ, Kutz SJ, Hoar BM, and Molnár PK (2022) Behaviour is more important than thermal performance for an Arctic host-parasite system under climate change. *Royal Society Open Science* 9. https://doi. org/10.1098/rsos.220060
- Phillips JA, Vargas Soto JS, Pawar S, Koprivnikar J, Benesh D, and Molnár PK (2022) The effects of phylogeny, habitat, and host characteristics on the thermal sensitivity of helminth development. *Proceedings of the Royal Society* B 289. https://doi.org/10.1098/rspb.2021.1878
- Rose H, Caminade C, Bolajoko MB, Phelan P, van Dijk J, Baylis M, Williams D, and Morgan ER (2016) Climate-driven changes to the spatio-temporal distribution of the parasitic nematode, *Haemonchus contortus*, in sheep in Europe. *Global Change Biology* 22, 1271–1285.
- **Rossanigo CE and Gruner L** (1995) Moisture and temperature requirements in faeces for the development of free-living stages of gastrointestinal nematodes of sheep, cattle and deer. *Journal of Helminthology* **69**, 357–362.
- **Rossanigo CE and Gruner L** (1996) The length of *strongylid* nematode infective larvae as a reflection of developmental conditions in faeces and consequences on their viability. *Parasitology Research* **82**, 304–311.
- Rossi L, Interisano M, Deksne G, and Pozio E (2019) The subnivium, a haven for *Trichinella* larvae in host carcasses. *International Journal for Parasitology: Parasites and Wildlife* 8, 229–233.
- Salame L and Glazer I (2015) Stress avoidance: vertical movement of entomopathogenic nematodes in response to soil moisture gradient. *Phytoparasitica* 43, 647–655.
- Santos MC, Silva BF, and Amarante AFT (2012) Environmental factors influencing the transmission of *Haemonchus contortus*. *Veterinary Parasitology* 188, 277–284.
- Saunders LM, Tompkins DM, and Hudson PJ (2000a) Spatial aggregation and temporal migration of free-living stages of the parasitic nematode *Trichos*trongylus tenuis. Functional Ecology 14, 468–473.
- Saunders LM, Tompkins DM, and Hudson PJ (2000b) The role of oxygen availability in the embryonation of *Heterakis gallinarum* eggs. *International Journal for Parasitology* 30, 1481–1485.
- Shearer CL and Ezenwa VO (2020) Rainfall as a driver of seasonality in parasitism. International Journal for Parasitology: Parasites and Wildlife 12, 8–12.
- Shifaw A, Feyera T, Elliott T, Sharpe B, Ruhnke I, and Walkden-Brown SW (2022) Method optimisation for prolonged laboratory storage of Ascaridia galli eggs. Veterinary Parasitology 309. https://doi.org/10.1016/j.vetpar.2022.109758
- Silva BF, Amarante MRV, Kadri SM, Carrijo-Mauad JR, and Amarante AFT (2008) Vertical migration of *Haemonchus contortus* third stage larvae on *Brachiaria decumbens* grass. *Veterinary Parasitology* **158**, 85–92.
- Stankiewicz M (1996) Observations on the biology of free-living stages of Parastrongyloides trichosuri [Nematoda, Rhabditoidea]. Acta Parasitologica 1, 38–42.
- Stromberg BE (1997) Environmental factors influencing transmission. Veterinary Parasitology 72, 247–264.
- Takeishi A (2022) Environmental-temperature and internal-state dependent thermotaxis plasticity of nematodes. Current Opinion in Neurobiology 74. https://doi.org/10.1016/j.conb.2022.102541
- Tarbiat B, Rahimian S, Jansson DS, Halvarsson P, and Höglund J (2018) Developmental capacity of Ascaridia galli eggs is preserved after anaerobic storage in faeces. Veterinary Parasitology 255, 38–42.
- Thomas F, Bonsall MB, and Dobson AP (2005) Parasitism, biodiversity, and conservation. pp. 124–139 in Thomas F, Renaud F, and Guegan J (Eds), *Parasitism and ecosystems*. Oxford: OUP Oxford.

- Thomas F, Poulin R, de Meeüs T, Guégan JF, and Renaud F (1999) Parasites and ecosystem engineering: what roles could they play? *Oikos* 84, 167–171.
- Thompson RCA, Lymbery AJ, and Smith A (2010) Parasites, emerging disease and wildlife conservation. *International Journal for Parasitology* 40, 1163–1170.
- Tompkins DM, Dobson AP, Arneberg P, Begon ME, Cattadori IM, Greenman JV, Heesterbeek JAP, Hudson PJ, Newborn D, Pugliese A, Rizzoli AP, Rosa R, Rosso F, and Wilson K (2002) Parasites and host population dynamics. pp. 45–62 in Hudson PJ, Rizzoli A, Grenfell BT, Heesterbeek H, and Dobson AP (Eds), *The ecology of wildlife diseases*. Oxford: Oxford University Press.
- van der Wal R, Irvine J, Stien A, Shepherd N, and Albon SD (2000) Faecal avoidance and the risk of infection by nematodes in a natural population of reindeer. *Oecologia* **124**, 19–25.
- van Dijk J, de Louw MDE, Kalis LPA, and Morgan ER (2009) Ultraviolet light increases mortality of nematode larvae and can explain patterns of larval availability at pasture. *International Journal for Parasitology* 39, 1151–1156.
- van Dijk J and Morgan ER (2008) The influence of temperature on the development, hatching and survival of *Nematodirus battus* larvae. *Parasit*ology 135, 269–283.
- van Dijk J and Morgan ER (2009) Hatching behaviour of Nematodirus filicollis in a flock co-infected with Nematodirus battus. Parasitology 136, 805–811.
- van Dijk J and Morgan ER (2011) The influence of water on the migration of infective trichostrongyloid larvae onto grass. Parasitology 138, 780–788.
- van Dijk J and Morgan ER (2012) The influence of water and humidity on the hatching of Nematodirus battus eggs. Journal of Helminthology 86, 287–292.
- van Dijk J, Sargison ND, Kenyon F, and Skuce PJ (2010) Climate change and infectious disease: helminthological challenges to farmed ruminants in temperate regions. *Animal* 4, 377–392.
- Vineer HR, Steiner J, Knapp-Lawitzke F, Bull K, Fernex EVS, Bosco A, Hertzberg H, Demeler J, Rinaldi L, Morrison AA, Skuce P, Bartley DJ, and Morgan ER (2016) Implications of between-isolate variation for climate change impact modelling of *Haemonchus contortus* populations. *Veterinary Parasitology* 229, 144–149.
- Viney ME (1996) Developmental switching in the parasitic nematode Strongyloides ratti. Biological Sciences 263, 201–208.
- Waghorn TS, Reynecke DP, Oliver AMB, Miller CM, Vlassoff A, Koolaard JP, and Leathwick DM (2011) Dynamics of the free-living stages of sheep intestinal parasites on pasture in the North Island of New Zealand. 1. Patterns of seasonal development. New Zealand Veterinary Journal 59, 279–286.
- Wang T, van Wyk JA, Morrison A, and Morgan ER (2014) Moisture requirements for the migration of *Haemonchus contortus* third stage larvae out of faeces. *Veterinary Parasitology* 204, 258–264.
- Wang T, Vineer HR, Morrison A, van Wyk JA, Bolajoko MB, Bartley DJ, and Morgan ER (2018) Microclimate has a greater influence than macroclimate on the availability of infective *Haemonchus contortus* larvae on herbage in a warmed temperate environment. Agriculture, Ecosystems and Environment 265, 31–36.
- Wang T, Vineer HR, Redman E, Morosetti A, Chen R, McFarland C, Colwell DD, Morgan ER, and Gilleard JS (2022) An improved model for the population dynamics of cattle gastrointestinal nematodes on pasture: parameterisation and field validation for Ostertagia ostertagi and Cooperia oncophora in northern temperate zones. Veterinary Parasitology 310. https://doi.org/10.1016/j.vetpar.2022.109777
- Weaver HJ, Hawdon JM, and Hoberg EP (2010) Soil-transmitted helminthiases: implications of climate change and human behavior. *Trends in parasitology* 26, 574–581.
- Young RR and Anderson N (1981) The ecology of the free-living stages of Ostertagia ostertagi in a winter rainfall region. Australian Journal of Agricultural Research 32, 371–388.