cambridge.org/dar

Research Article

Cite this article: Sousa RJM, Ribeiro SC, Baptista JAB and Silva CCG (2023). Evaluation of gamma-aminobutyric acid content in Portuguese cheeses with protected designation of origin status. *Journal of Dairy Research* **90**, 88–91. https://doi.org/10.1017/ S0022029923000043

Received: 31 March 2022 Revised: 5 September 2022 Accepted: 6 January 2023 First published online: 17 February 2023

Keywords:

Cheese; cow milk; GABA; PDO; sheep milk

Author for correspondence: Célia C. G. Silva, Email: celia.cg.silva@uac.pt

© The Author(s), 2023. Published by Cambridge University Press on behalf of Hannah Dairy Research Foundation





Evaluation of gamma-aminobutyric acid content in Portuguese cheeses with protected designation of origin status

Rodrigo J. M. Sousa, Susana C. Ribeiro, José A. B. Baptista and Célia C. G. Silva

IITAA-Institute of Agricultural and Environmental Research and Technology, University of the Azores, Angra do Heroísmo, Azores, Portugal

Abstract

Health-conscious consumers are increasingly paying attention to healthy diets and focusing on natural bioactive compounds in foods and their effects on mental health. This opens new opportunities for the study of artisanal cheeses as biofunctional foods. In the work described in this Research Communication, the gamma-aminobutyric acid (GABA) content of seven different Portuguese cheeses produced from unpasteurized cow, sheep, and goat milk and granted with protected designation of origin (PDO) status was analysed. The PDO cheeses made from cow milk analysed in this study were São Jorge (3, 4, 7, 12 and 24 months of maturation) and Pico cheeses. PDO cheeses made from sheep milk were Serra da Estrela, Serpa, Nisa and Azeitão. Cheeses made from sheep and goat milk included Beira Baixa yellow cheese. The GABA content in the Azorean PDO cheeses (made from cow milk) ranged from 1.23 to 2.64 g/kg of cheese. Higher variations in GABA content were observed in cheeses made from sheep and goat milk (0.73–2.31 g/kg). This study provides information on the GABA content in different Portuguese PDO cheeses and shows that hard or semi-hard ripened cheeses are a suitable matrix for GABA production by lactic acid bacteria.

The consumption of cheese is widespread throughout the world, as it is a convenient, ready-to-eat product and meets many nutritional needs due to its high content of various nutrients (Santiago-López *et al.*, 2018). Nowadays, consumers are increasingly aware that a healthy diet has potential benefits for human health. Biologically active compounds naturally present in cheese may provide these positive health effects in addition to their nutritional function (Sokovic Bajic *et al.*, 2019; Tofalo *et al.*, 2019).

Gamma-aminobutyric acid (GABA) is produced by glutamic acid decarboxylation by the enzyme L-glutamic acid decarboxylase. This enzyme is expressed by several types of microorganisms, including certain strains of lactic acid bacteria (LAB) associated with fermented dairy products such as yogurt and cheese. At the beginning of cheese production and during ripening, casein is the target of enzymes that cause proteolysis, releasing low molecular weight peptides and amino acids. These enzymes are present in milk (plasmin) and rennet (chymosin) or are released by the indigenous microbiota in raw milk. During this process, the release of a large amount of L-glutamate can be observed, as the native caseins of milk have a high content of this amino acid (Siragusa *et al.*, 2007; Tofalo *et al.*, 2019). Since it is a precursor of GABA, the accumulation of this amino acid leads to the synthesis of GABA in fermented foods. GABA is a neurotransmitter that plays an important role in the regulation of neuronal excitability as well as in the regulation of blood pressure (Inoue *et al.*, 2003; Siragusa *et al.*, 2007; Xu *et al.*, 2017; Tofalo *et al.*, 2019). Thus, GABA has the potential to serve as a bioactive ingredient in functional foods due to its antihypertensive, anti-anxiety and anti-stress properties (Linares *et al.*, 2016; Tofalo *et al.*, 2019).

Portuguese cheeses possessing protected designation of origin (PDO) status have a long history of manufacture following traditional methods. They are produced from raw cow, sheep and/or goat milks without the addition of starters. These cheeses are typically named according to the region where they are manufactured, and all of them possess unique features associated with their artisanal modes of manufacture and ripening (Reis and Malcata, 2011). These cheeses are becoming increasingly popular with consumers due to their unique texture and flavour. Serra da Estrela is probably the best representative of PDO cheeses made from fresh raw sheep milk and uses a plant extract (*Cynara cardunculus*) as a coagulant. Other PDO cheeses made from raw milk from small ruminants (sheeps and/or goats) include those produced in the south of mainland Portugal, such as Serpa, Azeitão, Nisa and Beira Baixa. PDO cheeses in the Azores made from raw cow milk include Pico and São Jorge (S. Jorge) cheeses, produced on the Azorean islands of the same name. Few studies have reported the presence of GABA in cheese, and none have investigated the GABA content in Portuguese PDO cheeses. The quantification of GABA, derived from the metabolism of microorganisms present in cheese made from unpasteurized milk, may contribute to the valorization of artisanal cheese in the Azores region and mainland Portugal. Therefore, the objective of our study was to determine the GABA content in PDO cheeses produced from cow milk in the Autonomous Region of the Azores and from sheep and goat milk in mainland Portugal.

Materials and methods

Portuguese PDO cheeses were purchased at the local market and included the following varieties made from cow milk: Pico and S. Jorge (cheeses aged 3, 4, 7, 12 and 24 months) and these made from sheep milk or a mixture of sheep and goat milk: Azeitão, Nisa, Serpa, Serra da Estrela and yellow cheese from Beira Baixa. Each cheese (5 g) was grated and 15 ml of HCl (0.1 M) was added, followed by homogenization at 4000 RPM for 5 min, with an Ultra–Turrax homogenizer (Janke E Kungel IKA* – Labortechnik T50). After homogenization, the containers were sealed and immersed in a water bath (Büchi Waterbath B–480) at 50°C for 30 min. Then the samples were centrifuged (Hermle Z – 323, Mandel Scientific Company) at $3000 \times g$ for 15 min. The supernatant was collected and stored (0–5°C) until determination of GABA content.

GABA content in cheese extracts was determined by HPLC (VWR Hitachi, model Elite Lachrom, Japan) equipped with an automated injector, an RP-18 column (Purospher® STAR, 5 µm particle size), and a Diode Array Detector (DAD), according to the method described by Cunha et al. (2022). Samples (100 µl) of the HCl/cheese extracts (supernatants) were mixed with 900 µl of 20% trichloroacetic acid (TCA, Sigma, Darmstadt, Germany). The mixture was centrifuged at $4000 \times g$ for 10 min, and the supernatant was filtered using 0.45 µm filters (Whatman, Darmstadt, Germany). The filtrate was derivatized with a solution of o-phthaldialdehyde (Sigma). The reaction solution (OPA) was prepared on the same day using 10 mg of o-phthaldialdehyde reagent, 10 µl of 2-mercaptoethanol (99% extra pure, Acros organics, Geel, Belgium) and 2.5 ml of acetonitrile (Sigma), in a tube protected from light. For the derivatization reaction, 100 µl of sample/standard, 500 µl of borate buffer pH 10.4 (0.4 M boric acid) and 100 µl of OPA reagent were used. In tubes protected from light, the mixture was vortexed for 30 s and reacted at room temperature for 5 min. Each sample (20 μ l) was injected and monitored at 334 nm. Ammonium acetate buffer (0.02 M), pH 7.3 (solvent A) and acetonitrile (solvent B) were used as solvents. System elution consisted of the following program: 80% solvent A and 20% solvent B for 5 min; increase to 100% solvent B for 5 min; reduction to 20% solvent B and 80% solvent A for 2 min. The flow rate used was 0.6 ml/min and the column temperature was set at 25°C. All solvents were filtered through a 0.45 μ m filter and degassed in an ultrasound bath (Ultrasons Medi-II, Selecta Calibration curves were plotted using four concentrations of the GABA standard (150, 300, 450, and 600 mg/l).

Statistical analysis

At least four samples were analysed for each cheese type. One-way analysis of variance (ANOVA) was applied to the results. When statistically significant differences (P < 0.05) were detected in the ANOVA, Tukey's test for multiple comparisons between cheese varieties was applied. Correlation analysis for maturation time of S. Jorge cheese and GABA content was performed by use of a bivariate (two-tailed significance) correlation test. Correlation was considered significant at P < 0.05. Statistical analysis was carried out with IBM SPSS Statistics, version 25 (SPSS Inc., Chicago, IL, USA).

Results

GABA concentration in PDO cheese made from cow milk varied from 1.23 to 2.26 g/kg cheese (Fig. 1). The lowest GABA level was found in Pico PDO cheese (1.23 \pm 0.24 g/kg), while the highest GABA concentration was observed in S. Jorge PDO cheese with 4 months maturation (2.64 \pm 0.32 g/kg). Since S. Jorge cheese is consumed with a wide range of ripening times (typically 3, 4, 7, 12, and 24 months), the variation in GABA content over the length of ripening time was also evaluated (Fig. 1). Despite the observed variation in the GABA content of S. Jorge cheese, no correlation (P > 0.05) was found with the ripening period. The highest GABA content was observed in 4-month-old cheeses (2.64 \pm 0.32 g/kg). However, a decrease in GABA content was observed in 7- and 12-month-old cheeses (2.07 \pm 0.63 g/kg and

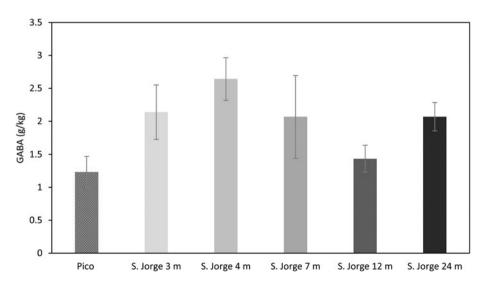


Fig. 1. Mean GABA concentration (g/kg cheese) in PDO cheeses made from raw cow milk: Pico cheese and S. Jorge cheeses with ripening times of 3, 4, 7, 12, and 24 months. The values are the average of four samples and the error bars show sem. No significant differences (P > 0.05) were found between cheese types.

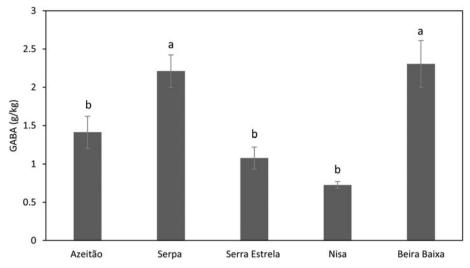


Fig. 2. Mean GABA concentration (g/kg cheese) in PDO cheeses made from raw sheep milk (Azeitão, Serpa, Serra da Estrela and Nisa) and in PDO cheese made from a mixture of goat and sheep milk (Beira Baixa). The values are the average of four samples and the error bars show SEM. Different lowercase letters indicate significant differences (P < 0.05) between cheese types.

 1.43 ± 0.20 g/kg, respectively), while GABA content increased in 24-month-old cheese (2.07 ± 0.22 g/kg). Cheeses made from sheep and goat milks are shown in Figure 2. The average GABA content in cheeses made from sheep milk was 1.36 ± 0.15 g/kg. High variability was observed in GABA values, which was significant (P < 0.05) and ranged from 0.73 to 2.31 g/kg. The lowest value was observed in Nisa cheese and the highest values were observed in Serpa cheese (made with sheep milk) and Beira Baixa spicy yellow cheese (made with a mixture of sheep and goat milk).

Discussion

Cheese made from raw milk has a high bacterial diversity that is not destroyed by pasteurization (Bottari *et al.*, 2018). Therefore, the microbial load of Portuguese traditional cheeses is high because they are not inoculated with commercial starter cultures and are made from raw milk (Reis and Malcata, 2011). The present study included traditional PDO cheeses produced either in the Azores from whole cow milk or in certain regions of mainland Portugal from sheep and goat milk.

The average GABA concentrations observed in this study (ranging from 0.73 to 2.64 g/kg) are higher than those found in other works. Siragusa *et al.* (2007) analysed the GABA concentration of 22 Italian cheeses and reported values ranging from 0.26 to 391 mg/kg of cheese. Similarly, Nomura *et al.* (1998) reported GABA levels ranging from 4.2 to 177 mg/kg in seven commercial cheeses (Camembert, Gouda, Blue, Cream, Cheddar, Edam, and Emmental). However, Lacroix *et al.* (2013) reported higher levels in four commercial French cheeses, with GABA content ranging from 0.3 to 3.2 g/kg cheese, so not dissimilar to our values. Also, high GABA levels, reaching maximum values of 10 g/kg, were reported in sheep milk cheeses from Sardinia (Manca *et al.*, 2015).

Compared with the cheeses made from cow milk, Pico cheese had the lowest GABA content, which could be related to its short ripening period, since it does not normally ripen for more than 22 d (Riquelme *et al.*, 2015). In contrast, the cheese S. Jorge had the highest GABA levels, since it has a longer ripening period, ranging from 3 to 24 months (Kongo *et al.*, 2009). The higher GABA levels in longer matured cheeses are most likely due to more extensive proteolysis in the cheeses. Milk caseins have a high content of glutamic acid, which can be released during

cheese ripening and converted to GABA by LAB with L-glutamic acid decarboxylase activity (Li et al., 2022). It has been reported that ripened hard and semi-hard cheeses provide an adequate matrix and ideal conditions for GABA production by LAB (Lacroix et al., 2013). The concentration of GABA in cheese is also related to the inhibition of GABA-degrading enzymes. GABA transaminase catalyses the reversible conversion of GABA to succinic semi-aldehyde, which is further metabolized to succinate (Carafa et al., 2019). In this way, the GABA content of aged cheese results from the balance between proteolysis and the release of glutamic acid, together with the action of L-glutamic acid decarboxylase, present in some LAB, and the degradation of GABA. This could explain the great variability of GABA content during the ripening of S. Jorge cheese. It is possible that the decrease in GABA content in cheeses older than 4 months is due to the action of GABA transaminases present in some LAB. In contrast, GABA levels increase again in cheeses with 24 months of maturation. Because different batches were analysed for each ripening time, it is not possible to conclude that the observed differences are due to an effective pattern of variation in GABA content during ripening of this cheese. It is possible that these differences are due to variations in the microbiota in each cheese/batch, which is to be expected in cheeses made from raw milk. To confirm this observation, the same batch would need to be analysed throughout the ripening period. However, several authors also reported similar variations in GABA content during the ripening period (Siragusa *et al.*, 2007; Manca et al., 2015).

In reference to Portuguese PDO cheese, made from raw sheep milk, the average GABA concentration $(1.36 \pm 0.15 \text{ g/kg})$ is slightly higher than the values reported by Diana *et al.* (2014) in two Spanish sheep milk cheeses (0.98 g/kg), but more similar to the GABA content found by Renes *et al.* (2019) in sheep milk cheeses (1.3 g/kg) with longer maturation time (240 d of ripening). According to Tofalo *et al.* (2019), the quality of sheep milk and the type of rennet used in the production of cheese seem to have some influence on the GABA content of these cheeses. Some of the Portuguese cheeses made from sheep milk, such as Serra da Estrela cheese, are curdled with an aqueous extract of wild thistle (*Cynara cardunculus*). In Serra da Estrela cheese, it was previously found that GABA content increases with ripening time, reaching 55 and 131 mg/100 g DM (about 0.36 and 0.85 g/kg of cheese) after 90 and 150 d, respectively

(Tavaria *et al.*, 2003). In the present study, the GABA levels of Serra da Estrela cheese were slightly higher than the values observed in 150-day-old cheese. However, Tavaria *et al.* (2003) reported that cheese made from refrigerated milk had higher levels of GABA, which was due to the growth of psychrotrophic bacteria known to favour the decarboxylation of glutamate to GABA. In addition, several authors reported that cheese made from sheep and goat milk had the highest GABA levels and had a microbiota with a higher GABA-producing capacity than cheese made from cow milk (Siragusa *et al.*, 2007; Diana *et al.*, 2014). However, the present study showed that raw cow milk cheese may contain high levels of GABA. In addition, several LAB strains previously isolated from raw cow milk (Pico and S. Jorge PDO cheeses) were found to produce high levels of GABA (Ribeiro *et al.*, 2018; Cunha *et al.*, 2022).

Based on the range of GABA content in cheeses of 73–264 mg/ 100 g determined in this work, a daily portion of 30 g of cheese provides approximately 22–79 mg of GABA. Thus, all PDO cheeses analysed had a GABA content greater than 16 mg per serving, a value that was reported as the minimum value associated with health promotion in the study by Renes *et al.* (2019). The same authors reported that consumption of a serving of cheese with a GABA content of 16 mg was effective in lowering blood pressure in individuals with mild hypertension. In the present study, all PDO cheeses analysed exceeded this value and had the potential to lower blood pressure.

In conclusion, a large variability in total GABA content was found between the different Portuguese PDO cheeses. However, regardless of the origin of the milk (cow, sheep or goat milk), the cheeses investigated in this study had high levels of GABA, above the minimum amount indicated for a health-promoting effect, so that they can be proposed as functional foods.

Acknowledgements. This work was supported by the Fundação para a Ciência e Tecnologia (FCT) – Project UID/CVT/00153/2019. S.C. Ribeiro is thankful to the Fundo Regional da Ciência e Tecnologia (FRCT) for financial support – grant M3.1.a/F/017/2018.

References

- Bottari B, Levante A, Neviani E and Gatti M (2018) How the fsheepst become the greatest. L. casei's impact on long ripened cheeses. *Frontiers in Microbiology* 9, 2866.
- Carafa I, Stocco G, Nardin T, Larcher R, Bittante G, Tuohy K and Franciosi E (2019) Production of naturally γ-aminobutyric acid-enriched cheese using the dairy strains Streptococcus thermophilus 84C and Lactobacillus brevis DSM 32386. Frontiers in Microbiology 10, 93.
- Cunha DS, Coelho MC, Ribeiro SC and Silva CC (2022) Application of Enterococcus malodoratus SJC25 for the manufacture of whey-based beverage naturally enriched with GABA. Foods (Basel, Switzerland) 11(3), 447.
- Diana M, Rafecas M, Arco C and Quílez J (2014) Free amino acid profile of Spanish artisanal cheeses: importance of gamma-aminobutyric acid (GABA) and ornithine content. *Journal of Food Composition and Analysis* 35(2), 94–100.
- Inoue K, Shirai T, Ochiai H, Kasao M, Hayakawa K, Kimura M and Sansawa H (2003) Blood-pressure-lowering effect of a novel fermented

milk containing γ-aminobutyric acid (GABA) in mild hypertensives. *European Journal of Clinical Nutrition* **57**(3), 490–495.

- Kongo JM, Gomes AM, Malcata FX and McSweeney P (2009) Microbiological, biochemical and compositional changes during ripening of São Jorge – a raw milk cheese from the Azores (Portugal). Food Chemistry 112(1), 131–138.
- Lacroix N, St-Gelais D, Champagne C and Vuillemard J (2013) Gamma-aminobutyric acid-producing abilities of lactococcal strains isolated from old-style cheese starters. *Dairy Science & Technology* 93(3), 315–327.
- Li Y, Chen G, Ge F, Dang T, Ren Y, Zeng B and Li W (2022) Characterization and mutagenesis of a novel *Mycobacterium smegmatis*derived glutamate decarboxylase active at neutral pH. World Journal of *Microbiology and Biotechnology* 38, 75.
- Linares DM, O'Callaghan TF, O'Connor PM, Ross RP and Stanton C (2016) Streptococcus thermophilus APC151 strain is suitable for the manufacture of naturally GABA-enriched bioactive yogurt. Frontiers in Microbiology 7, 1876.
- Manca G, Porcu A, Ru A, Salaris M, Franco MA and De Santis E P (2015) Comparison of γ -aminobutyric acid and biogenic amine content of different types of sheep milk cheese produced in Sardinia, Italy. *Italian Journal of Food Safety* **4**(2), 123–128.
- Nomura M, Kimoto H, Someya Y, Furukawa S and Suzuki I (1998) Production of γ -aminobutyric acid by cheese starters during cheese ripening. *Journal of Dairy Science* **81**(6), 1486–1491.
- Reis PJ and Malcata FX (2011) Current state of Portuguese dairy products from ovine and caprine milks. *Small Ruminant Research* **101**(1–3), 122–133.
- **Renes E, Ladero V, Tornadijo M and Fresno JM** (2019) Production of sheep milk cheese with high γ-aminobutyric acid and ornithine concentration and with reduced biogenic amines level using autochthonous lactic acid bacteria strains. *Food Microbiology* **78**, 1–10.
- Ribeiro SC, Domingos-Lopes MFP, Stanton C, Ross RP and Silva CCG (2018) Production of γ-aminobutyric acid (GABA) by Lactobacillus otakiensis and other Lactobacillus sp. isolated from traditional Pico cheese. International Journal of Dairy Technology 71, 1012–1017.
- Riquelme C, Câmara S, Maria de Lurdes N, Vinuesa P, da Silva CCG, Malcata FX and Rego OA (2015) Characterization of the bacterial biodiversity in Pico cheese (an artisanal Azorean food). *International Journal of Food Microbiology* 192, 86–94.
- Santiago-López L, Aguilar-Toalá JE, Hernández-Mendoza A, Vallejo-Cordoba B, Liceaga AM and González-Córdova AF (2018) Invited review: bioactive compounds produced during cheese ripening and health effects associated with aged cheese consumption. *Journal of Dairy Science* 101(5), 3742–3757.
- Siragusa S, De Angelis M, Di Cagno R, Rizzello CG, Coda R and Gobbetti M (2007) Synthesis of γ-aminobutyric acid by lactic acid bacteria isolated from a variety of Italian cheeses. *Applied and Environmental Microbiology* **73**(22), 7283–7290.
- Sokovic Bajic S, Djokic J, Dinic M, Veljovic K, Golic N, Mihajlovic S and Tolinacki M (2019) GABA-producing natural dairy isolate from artisanal zlatar cheese attenuates gut inflammation and strengthens gut epithelial barrier in vitro. *Frontiers in Microbiology* 10, 527.
- Tavaria FK, Franco I, Carballo FJ and Malcata FX (2003) Amino acid and soluble nitrogen evolution throughout ripening of Serra da Estrela cheese. *International Dairy Journal* 13(7), 537–545.
- **Tofalo R, Perpetuini G, Battistelli N, Pepe A, Ianni A, Martino G and Suzzi G** (2019) Accumulation γ-aminobutyric acid and biogenic amines in a traditional raw milk sheep cheese. *Foods (Basel, Switzerland)* **8**(9), 401.
- Xu N, Wei L and Liu J (2017) Biotechnological advances and perspectives of gamma-aminobutyric acid production. World Journal of Microbiology and Biotechnology 33(3), 1–11.