

crease nosocomial infections in an intermediate care unit. *Crit Care Med* 2004;32(2):358–363.

9. Levchenko AI, Boscart VM, Fernie GR. The effect of automated monitoring and real-time prompting on nurses' hand hygiene performance. *Comput Inform Nurs* 2013;31(10):498–504.

Anatomic Sites of Colonization with Community-Associated Methicillin-Resistant *Staphylococcus aureus*

Methicillin-resistant *Staphylococcus aureus* (MRSA) has emerged over the past 20 years as a cause of infections in community populations, so-called community-associated MRSA (CA-MRSA). By pulsed-field gel electrophoresis (PFGE) subtyping, USA300 is the most common CA-MRSA strain in the United States. It has been suggested that the colonization dynamics for CA-MRSA may be different than those for traditional MRSA strains,¹ with extranasal colonization potentially playing a role in CA-MRSA transmission and infection.¹

Another distinguishing characteristic of USA300 MRSA strains is greater susceptibility to non- β -lactam antibiotics compared with healthcare-associated MRSA strains. However, multidrug-resistant (MDR) USA300 strains have been described, largely among patients infected with human immunodeficiency virus (HIV) and men who have sex with men (MSM).² Also of concern, resistance to common decolonizing agents, such as mupirocin² and chlorhexidine gluconate (CHG),³ has been reported in CA-MRSA.

The objectives of this study were to examine the phenotype of USA300 MRSA strains, the prevalence of the *qacA/B* gene in this population, and the anatomic sites of colonization by PFGE pattern.

We previously reported on the prevalence of nasal and extranasal CA-MRSA colonization among inpatients (374 HIV infected and 371 HIV negative) at Stroger Hospital of Cook County (CCH), the major safety net hospital in Chicago.⁴ As described elsewhere, nasal and extranasal (throat, axilla, inguinal, perirectal, and chronic wound, if present) surveillance swab specimens were collected from patients within 72 hours of admission from March 2011 to April 2012; cultures were processed with broth enrichment.⁴ Sex was recorded, and enrolled men were asked whether they identified themselves as MSM. Genotypic analysis with PFGE was performed on all identified MRSA isolates. Results were interpreted as described by McDougal et al.⁵

Confirmed MRSA isolates had antibiotic susceptibility determined (MicroScan Walkaway System, Siemens Healthcare Diagnostics). For USA300 MRSA strains, MDR was defined as resistance to 4 or more non- β -lactam antibiotic classes. High-level mupirocin resistance was assessed using disk diffusion.⁶

Carriage of *qacA* and *qacB* genes, which code for efflux

pumps associated with increased minimum inhibitory concentrations of CHG,⁷ was assessed using real-time polymerase chain reaction, as described previously.⁸

A χ^2 test was used to examine the association of PFGE patterns and colonization sites, with Fisher exact test used for small samples. SAS software (ver. 9.2; SAS Institute) was used for statistical analysis. The study was approved by the institutional review board of CCH and Rush University Medical Center.

We observed that following the nares, the perirectal area was the second most common site of colonization (58% of colonized individuals). Prevalence of extranasal and exclusive extranasal colonization was not significantly different between patients colonized with USA300 or non-USA300 strains (Table 1). However, the average number of sites colonized was significantly higher for USA300 versus non-USA300 strains (2.8 [standard deviation (SD), 1.51] and 2.2 [SD, 1.48], respectively; $P = .049$). Inguinal, perirectal, and concomitant inguinal and perirectal colonization were all significantly associated with colonization with the USA300 strain type in comparison to non-USA300 MRSA strains (Table 1). Inguinal or perirectal MRSA colonization was found more often in men (63/480; 13%)—MSM (odds ratio [OR], 2.2 [95% confidence interval (CI), 1.1–4.2]; $P = .02$) and heterosexual men (OR, 1.8 [95% CI, 1.02–3.2]; $P = .04$)—than in women (20/265; 8%; OR, 1.9 [95% CI, 1.1–3.1]; $P = .02$).

There were 5 individuals who had an MRSA infection at the time of enrollment, and they were all found to have colonization with MRSA. Four of these individuals had skin and soft tissue infections and were colonized with the USA300 strain type, and 1 individual had a bloodstream infection and was colonized with a non-USA300 strain type. Excluding chronic wound cultures, each of these individuals had 3–5 sites of MRSA colonization, suggesting a significant level of extranasal colonization and colonization burden for individuals infected with MRSA.

Of the colonized individuals, 3.4% carried high-level mupirocin-resistant strains (1 USA100, 2 USA500, 1 USA300). Of the individuals colonized with USA300 MRSA strains, 4 (5%) carried MDR strains. There were 117 MRSA isolates evaluated for the presence of the *qacA/B* genes; all were negative.

We examined colonization and molecular characteristics of CA-MRSA isolates collected from patients seeking care at the major safety net hospital in Chicago. We found that inguinal and perirectal colonization was more common with the USA300 strain type than with non-USA300 MRSA strains. In addition, highly antibiotic-resistant USA300 MRSA strains were rare, and none of the MRSA isolates collected over a 14-month study period were found to harbor the *qacA/B* genes.

We observed that males—both heterosexual males and MSM—had a higher prevalence of inguinal and perirectal MRSA colonization in comparison to females. Similarities observed in colonization patterns between MSM and heterosexual males suggest that perhaps social, hormonal, skin

TABLE 1. Association of Pattern of Anatomic Site of Colonization and Pulsed-Field Gel Electrophoresis Profile among Individuals Colonized with Community-Associated Methicillin-Resistant *Staphylococcus aureus* (CA-MRSA)

| Anatomic site of colonization | USA300 (<i>n</i> = 79) ^a | Non-USA300 (<i>n</i> = 36) | OR (95% CI) | <i>P</i> |
|-----------------------------------|--------------------------------------|-----------------------------|-----------------|----------|
| Anterior nares | 50 (63) | 21 (58) | ... | .61 |
| Throat | 35 (44) | 16 (44) | ... | .99 |
| Axilla | 31 (39) | 12 (33) | ... | .54 |
| Inguinal | 49 (62) | 15 (42) | 2.3 (1.02–5.11) | .04 |
| Perirectal | 52 (66) | 15 (42) | 2.7 (1.2–6.06) | .015 |
| Inguinal and perirectal region | 40 (51) | 9 (25) | 3.1 (1.28–7.37) | .01 |
| Wound | 6 (8) | 1 (3) | ... | .43 |
| Extranasal colonization | 74 (94) | 32 (89) | ... | .46 |
| Exclusive extranasal colonization | 29 (37) | 15 (42) | ... | .61 |
| Sites colonized, average (SD) | 2.8 (1.51) | 2.2 (1.48) | ... | .049 |

NOTE. Data are no. (%) of patients, unless otherwise indicated. Any extranasal colonization was defined as the presence of CA-MRSA extranasal colonization irrespective of anterior nares culture results. Exclusive extranasal colonization was defined as CA-MRSA colonization at extranasal sites and negative anterior nares cultures for CA-MRSA. CI, confidence interval; OR, odds ratio; SD, standard deviation.

^a Two individuals were colonized with both USA300 and non-USA300 strains and were excluded from the comparison of USA300 to non-USA300 strains. Therefore, the total number of patients used in the analysis was 115.

biology, or genetic differences between sexes play a role in colonization dynamics rather than sexual orientation.⁹

The absence of *qacA/B* genes among MRSA isolates in the population we studied is consistent with other reports in the United States.¹⁰ In contrast to reports in San Francisco,² MDR USA300 strains were relatively rare in our population, which comprised a diverse group of HIV-infected and HIV-negative individuals. However, continued surveillance of antibiotic resistance patterns is needed to understand the evolving epidemiology of USA300 strains as well as to inform empiric therapeutic decisions.

Our study is limited in that we did not assess frequency of mupirocin or CHG use in our population, although CCH does not have mupirocin on its antibiotic formulary and CHG bathing use has been limited to intensive care unit and pre-operative patients. In addition, we relied on self-report of MSM status, which could lead to recall bias. Finally, we performed PFGE on 1 MRSA colony morphotype per body site and therefore may have failed to detect coexistent minority subpopulations.

In summary, our study highlights that inguinal and perirectal colonization appears to be more frequent with the USA300 strain type and that sex may play a role in location of extranasal CA-MRSA colonization. Patients with clinical MRSA infections appeared to be those with more sites of MRSA colonization. Although mupirocin resistance and presence of *qacA/B* were uncommon, continued monitoring of MRSA prevalence and resistance is warranted.

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REFERENCES

- Miller LG, Diep BA. Clinical practice: colonization, fomites, and virulence: rethinking the pathogenesis of community-associated methicillin-resistant *Staphylococcus aureus* infection. *Clin Infect Dis* 2008;46(5):752–760.

2. Diep BA, Chambers HF, Graber CJ, et al. Emergence of multidrug-resistant, community-associated, methicillin-resistant *Staphylococcus aureus* clone USA300 in men who have sex with men. *Ann Intern Med* 2008;148(4):249–257.
3. Fritz SA, Hogan PG, Camins BC, et al. Mupirocin and chlorhexidine resistance in *Staphylococcus aureus* in patients with community-onset skin and soft tissue infections. *Antimicrob Agents Chemother* 2013;57(1):559–568.
4. Popovich KJ, Hota B, Aroutcheva A, et al. Community-associated methicillin-resistant *Staphylococcus aureus* colonization burden in HIV-infected patients. *Clin Infect Dis* 2013;56(8):1067–1074.
5. McDougal LK, Steward CD, Killgore GE, Chaitram JM, McAllister SK, Tenover FC. Pulsed-field gel electrophoresis typing of oxacillin-resistant *Staphylococcus aureus* isolates from the United States: establishing a national database. *J Clin Microbiol* 2003;41(11):5113–5120.
6. Clinical and Laboratory Standards Institute (CLSI). *Performance Standards for Antimicrobial Susceptibility Testing: 21st Informational Supplement*. Wayne, PA: CLSI, 2011. CLSI document M100-S21.
7. Horner C, Mawer D, Wilcox M. Reduced susceptibility to chlorhexidine in staphylococci: is it increasing and does it matter? *J Antimicrob Chemother* 2012;67(11):2547–2559.
8. Hayden MK, Lolans K, Li H, et al. Chlorhexidine gluconate (CHG) susceptibility of methicillin-resistant *Staphylococcus aureus* (MRSA) isolates from a multi-state study of adult intensive care unit (ICU) patients. In: *20th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America*. April 1–4, 2011; Dallas, TX. Abstract 369.
9. Zanger P, Nurjadi D, Gaile M, Gabrysch S, Kreamsner PG. Hormonal contraceptive use and persistent *Staphylococcus aureus* nasal carriage. *Clin Infect Dis* 2012;55(12):1625–1632.
10. McGann P, Kwak YI, Summers A, Cummings JF, Waterman PE, Lesho EP. Detection of qacA/B in clinical isolates of methicillin-resistant *Staphylococcus aureus* from a regional healthcare network in the eastern United States. *Infect Control Hosp Epidemiol* 2011;32(11):1116–1119.