Fasciola hepatica: surfaces involved in movement of miracidia and cercariae

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Abstract

Rapid freezing and substitution with fixative prior to scanning electron microscopy was used to demonstrate the pattern of beat and recovery of the cilia of free swimming miracidia of Fasciola hepatica. There were stages of dexioplectic metachronal co-ordination and the power stroke was approximately 15° anticlockwise from the anterior-posterior axis. Around the circumference of the body of the miracidia there were approximately 12 metachronal waves of power and recovery. Free-swimming cercariae were recorded by time-lapse photography and, after conventional fixation, by scanning electron microscopy. Cercarial tail-beats were to the posterior of the body in the lateral plane at a rate of 8 Hz. The tail has paired lateral ridges positioned to act as leading edges. There is an array of 32 sensory papillae on the mid-ventral surface of the tail. The tegument of the most distal part of the tail is described: it is free of sensory endings and the surface shows a spiral pattern.

Introduction

Fasciola hepatica is a useful model for the understanding of digenean parasites (Bennett, 2001) and both F. hepatica and F. gigantica are infections of humans and animals (sheep and cattle) (Boray, 1985). Neuro-cytochemical studies of Fasciola have provided evidence of invertebrate-specific neurotransmitters (Brownlee et al., 1994; Brownlee & Fairweather, 1996) and as a result there is interest in the normal mechanisms of movement of these parasites. In all stages of the life cycle, e.g. in the newly excysted juvenile (Bennett & Threadgold, 1973) there is a basket of circular and longitudinal muscle which acts on the enclosed mesenchyme and organs that provide a hydrostatic skeleton.

Various aspects of the structure and movement of the free-living larval stages of F. hepatica have been described previously. Miracidia are propelled by cilia and swim in a spiralling movement in the direction of the apical papilla (Wilson, 1969; Wilson & Denison, 1970). The cilia of F. hepatica miracidia are found on 21 epidermal cells arranged in rows: 6 cells are in the most anterior row, with 6, 3, 4 and 2 respectively in subsequent rows (Mattes, 1949). The configuration of rootlets of the cilia (Wilson, 1969; Southgate, 1970) is comparable with that of other Platyhelminthes (Dorey, 1965) with main rootlets directed at right angles to the insertion of each cilium. Rootlets are arranged in a symmetrical circumferential pattern with spacing of approximately $2 \mu m$ (Wilson, 1969). In turbellarians, cilia beat synchronously in the same plane, with conspicuous metachronal wavefronts parallel to the meridians (Dorey, 1965). Knight-Jones (1954) described the beating of ciliated epithelia of Turbellaria such as Dugesia tigrina and Crenobia alpina as in antiplectic metachronism.

This study aims to demonstrate and interpret the beating of both the cilia of the miracidium and the tail of the cercaria of F. hepatica. The cercarial body and tail of F. hepatica are covered with a tegument, which is also found on other life-cycle stages of F. hepatica (Threadgold, 1963; Bennett, 1975a,b). While the surface morphology of the gymnocephalus cercaria of F. hepatica has been described from scanning electron microscopy (Køie et al., 1977) there have been no detailed descriptions of its movement. Those cercariae whose movement has been described include Parorchis acanthus (Rees, 1932), Himasthala secunda and Cryptocotyle lingua, (Chapman & Wilson, 1973; Chapman, 1974), and Transversotrema (Bundy, 1981). There are lateral fins on the tails of some cercariae, e.g. Clonorchis sinensis (Fujino et al., 1979).

The tail musculature of H. secunda and C. lingua has

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Fig. 1. Scanning electron micrographs of Fasciola hepatica miracidia (A and B) and cercariae (C, D and E). A. Miracidium fixed while swimming in direction of the arrow. B. High power region (from A) with cilia fixed in phases of dexioplectic metachronism. The direction of the power stroke of the metachronal wave is arrowed. C. Cercaria: the tail has paired lateral ridges and a region between the arrows with many sensory endings. D. Cercarial tail: (similar to region between arrows in C) with lateral ridges (R) and 8 μ m cilia on the ventral surface. E. The terminal portion of the cercarial tail with a spiral surface pattern to the tegument.

been examined by transmission electron microscopy (TEM) and is arranged in blocks, two being dorso-lateral and two ventro-lateral (Chapman, 1973). A similar arrangement of muscle blocks is to be found in the furcocercaria of Schistosoma mansoni (Nuttman, 1974). The rate of tail beating of cercariae of H. secunda, P. acanthus and S. mansoni has been estimated (Bundy, 1981) as 10.6 Hz, 2.9 Hz and 22.1 Hz respectively. Serotonin (5- HT) is an excitatory transmitter in the body wall musculature of F. hepatica (Tembe et al., 1993). In the tail of the cercaria of Sanguinicola inermis (McMichael-Phillips et al., 1996) cell bodies associated with the ventral and dorsal longitudinal nerve cords demonstrated either FMRFamide and S1 or 5-HT-like immunoreactivity. In the cercarial tail of S. inermis only 5-HT extended into the posterior tail furcae and there is other evidence that the serotinergic and peptidergic components of the nervous system are distinct. The unusual pattern of banding of the musculature, peculiar to the Platyhelminthes, is

thought to be a specific adaptation to rapid tail movement (Reger, 1976).

Rapidly moving cilia of protozoa have been best preserved for scanning electron microscopy (SEM) by a process of rapid freezing followed by fixation as thawing proceeds (substitution) (Barlow & Sleigh, 1978). It was proposed to use this method to determine the angle and type of metachronal beating of the cilia of F. hepatica miracidia.

Short sensory papillae $(1-3 \mu m)$ are found on the body of the cercaria of *F. hepatica* (Køie *et al.*, 1977) as well as on newly-excysted juveniles around the oral sucker, and scattered over the body surface between the spines (Bennett, 1975a). Relatively long $(8 \mu m)$ ciliary sensory endings are commonly present on the cercarial body and tails of many species of digeneans, e.g. Metagonimus (Fujino & Ishii, 1976). Sensory endings of this size have been described previously on the tail of F. hepatica cercariae (Køie et al., 1977) along with other prominent

Fig. 2. Phases of movement of Fasciola hepatica cercariae. Alternate time-lapse video-fields (intervals of 0.04 sec). One complete cycle = 6 video-fields = 0.12 sec. Scale bar = 50μ m. -, First video-field; - - - -, third video-field; ..., fifth videofield.

features: the dorso-lateral folds on either side of the tail. Additional aims of this study, therefore, were to relate the tail structure and position of sensory endings to the behaviour of the cercaria and movement of the tail as viewed by time lapse video-photography.

Materials and methods

Miracidia

Production

Eggs of F. hepatica from experimentally infected animals were supplied by the Veterinary Laboratories Agency (VLA), Weybridge, UK. They were embryonated at 22° C for 14 days (Al-Habbib, 1974) and miracidia were hatched from the eggs after exposure to light in filtered, oxygenated aquarium water.

Preparation for SEM

Several hundred, freshly hatched and free-swimming miracidia were frozen by plunging groups of specimens in $3 \mu l$ drops of water into liquid isopentane. Substitution was carried out over $30-40$ h with a solution comprising 40% ethylene glycol and 50% methanol saturated with $CaCl₂$ and $HgCl₂$, and 10% acrolein (Barlow & Sleigh, 1978). Critical point drying was carried out (Bartlett & Burstyn, 1975) in a specially constructed basket (Newell & Wrath, 1975). Specimens were sputtercoated with gold (Bennett, 1975a,b).

Scanning electron micrograph analysis

The number of metachronal waves per miracidium was estimated from photomicrographs of 50 freezesubstituted miracidia that had been fixed in the typical

Cercariae

Production

Miracidia of F. hepatica were used to infect Lymnaea truncatula at a rate of 5 per snail and after 40 days cercariae were stimulated to emerge by 'cold shock', by reducing the temperature of infected snails to 12°C for 30±45 min (Kendall & McCullough, 1951).

Preparation for SEM

Free-swimming cercariae were fixed with 3% glutaraldehyde in 0.1 ^M cacodylate buffer and, after dehydration and critical point drying, sputter-coated as above. Scanning electron microscopy was used to interpret the surface features and sensory structures of the tail.

Preparation for time-lapse photography

Cercariae were placed in a 4 mm horizontal cuvette under a phase contrast microscope and viewed at $\times 400$. Movement of cercariae was recorded by time lapse on VHS equipment, which allowed viewing of video-fields at intervals of 0.02 sec.

Results

Rapidly fixed F. hepatica miracidia had a body shape which was consistent with rapid swimming, i.e. there was a body extension of approximately $110 \mu m \times 35 \mu m$, (Erasmus, 1972). Cilia were arrested in typical positions of a dexioplectic metachronal beat with the orientation of the ciliary power stroke at 15° (\pm S.E. 5 $^{\circ}$) from the longitudinal body axis [\(fig. 1A\). T](#page-1-0)here were approximately 12 metachronal waves around the circumference of each miracidium, (six visible from each lateral view of a single miracidium). Some cilia were slightly damaged at their tips in the freeze-substitution processi[ng \(fig. 1B\).](#page-1-0)

Time-lapse images of F. hepatica cercarial swimming reveal that the cercarial tail beats broadly in the lateral plane with one cycle of beating per 0.12 sec (8 Hz) (fig. 2). When viewed from the dorsal or ventral aspect in real time the most posterior two-thirds of the tail provide an illusion of a figure of eight centred on an axis horizontally below the anterior-posterior line of the body and tail. As a result there is a side-to-side oscillation of the body of the cercaria centred on the ventral sucker in the middle of the body.

Standard fixation for SEM resulted in tails fixed in a range of configurations similar to those exhibited by time-lapse photography described above (fig. 2). The tail is approximately $500 \mu m$ in length and is covered with an invaginated tegument and both anterior-posterior and transverse corrugatio[ns \(fig. 1E\). A](#page-1-0) pair of prominent lateral ridges or fins extended along either side of the tail [\(fig. 1C and E\).](#page-1-0) A consistent feature of all cercarial tails was the spiral pattern of the tegument on the most distal portion $(35-40 \mu m)$ [\(fig. 1D\).](#page-1-0) Ciliated sensory endings were arrayed only on the mid-ventral surface of the tail in 16 pairs, 13 pairs with long (approximately $8 \mu m$) cilia, the most distal three pairs having shorter $(1-2 \mu m)$ cilia.

Discussion

The spiral swimming pattern of the miracidia of F. hepatica (Wilson & Denison, 1970) is explained by the images of angled beating of the cilia recorded here. Miracidia fixed by rapid freeze-substitution successfully preserved cilia in positions of power stroke and sideways recovery typical of dexioplectic co-ordination (Knight-Jones, 1954). This pattern of beating has not previously been described in the Platyhelminthes where antiplectic beating was ascribed to the Turbellaria (Knight-Jones, 1954). The ciliary power stroke is at approximately 15° to the main axis of the bod[y \(fig. 1A\).](#page-1-0) Fasciola hepatica, therefore, swims by power strokes of its cilia directed approximately along the sinistral ciliary rows. The sinistral line of cilia of a platyhelminth (Dorey, 1965) is at approximately 30° to the anterior-posterior line. A fully extended miracidium of *F. hepatica*, in swimming posture, is approximately $170 \mu m \times 20 \mu m$ (Erasmus, 1972). This is $1.5-2.0\times$ its relaxed length. Extension and reduction of body length by adjustments of the hydrostatic skeleton may effect both a change of angle of the sinistral line of ciliary rootlets and the direction of the power stroke as indicated by local body shape. The more pronounced spiralling seen in ageing miracidia which are shorter and wider is consistent with this idea. Corkscrewing, as imparted by the rifling of gun-barrels, is associated with stability in flight and the 15° angled beat of the cilia demonstrated here may have been selected for to this end.

In many other species the beating of the cercarial tail is across or over the body. In the case of F. hepatica, however, the beating is at a distance from the body with the effective strokes being in the lateral plane. Dorsolateral folds or ridges are present on each side of the tail of F. hepatica cercariae (Køie et al., 1977) and are in a similar position to the more prominent fins seen on the tail of \tilde{C} . sinensis (Fujino et al., 1979). These structures were consistently found in this study but were not sufficiently large to be described as fins and are therefore referred to here as lateral ridges. Since the beating of the tail is in the direction of these ridges, they may provide an effective leading edge. No sensory papillae are present in the most posterior region of the tail where the tegument is spirally twisted. There is no explanation for this spiralled tail-end in the literature, leaving one to speculate for example that it may play a part in emergence of the cercaria from the redia and snail host.

The beating of the tail of *F. hepatica* cercariae differs from that of other cercariae recorded to date. In P. acanthus, for example, the tail and body counter-balance each other while beating as a single unit in one plane (Rees, 1932). It also differs somewhat from the tailbeating of H. secunda and C. lingua, which both coil the tail behind the body in the cycle of beating. There is, however, some similarity to C. lingua, which with its longer tail tends towards a figure of eight pattern of beating (Chapman & Wilson, 1973). There are other similarities with the beating of the furcate cercarial tail of T. patialense with respect to the beat and recovery strokes and approximate orientation to the body (Bundy, 1981). In F. hepatica the beating of the tail is lateral, being in the same plane as the dorso-ventrally flattened body.

Additionally, the oscillation of the body is such that there is minimal movement of the ventral sucker. Both of these features may be useful adaptations to attachment to a flat substrate to encyst. Bulbous sensory endings with enclosed cilia located around the ventral sucker in the cercaria/infective stage (Bennett, 1975a) are well placed to detect this contact.

The large $8 \mu m$ uniciliate papillae of *F. hepatica* are found in similar positions to those on the tail of S. mansoni cercariae (Gordon et al., 1934; Sakamoto & Ishii, 1978). Ventral papillae on the tail may provide feedback about the movement of the cercarial tail in the lateral plane or, on contact, inhibit tail beating on location of a substrate for encystment.

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