

Sub-Ångström-resolution MicroED Using a Direct Detection Camera

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The recent development of microcrystal electron diffraction (MicroED) in electron cryo-microscopy (cryo-EM) has addressed many of the challenges of X-ray crystallography [1], filling a long-standing need in organic chemistry for rapid and accurate high-resolution structural characterization [2]. Like X-ray crystallography, this is also a diffraction technique requiring crystalline specimens, but because high energy electrons have a more favorable ratio of elastic to inelastic scattering [3], MicroED can acquire data using crystals that are orders of magnitude smaller than those needed for X-ray crystallography—even as small as 50 nm in length [4]. Additionally, the instrumentation required for MicroED is readily available, as many chemistry and biology labs already have access to cryo-capable transmission electron microscopes (TEM).

One of the critical hinderances to the broad adoption of this technique is its current requirement for a specialized TEM camera system [5]. Extending this technique to use the same direct detection camera systems already present in cryo-EM labs would significantly eliminate a large (and costly) barrier to adoption of this technique and allow it to reach its full potential for propelling widespread scientific progress. Additionally, the improved capabilities of direct detection cameras [6,7] compared to the scintillator-based cameras currently used for MicroED have the potential to significantly improve data quality, provided that they also have sufficient dynamic range to capture diffraction patterns. Direct detection cameras can deliver a much higher framerate, significantly improved single-electron signal-to-noise ratio, and significantly higher resolution than scintillator-based cameras.

We acquired continuous-rotation MicroED data using a DE-64 direct detection camera (Direct Electron LP, San Diego, CA) mounted on a Talos Arctica 200 keV TEM (Thermo Fisher, Waltham, MA). Small molecule biotin (Sigma-Aldrich, St Louis, MO) was used as a test sample in our experiment. This lyophilized powder was directly extracted from the commercial bottle, ground between two glass slides, and placed on a glow-discharged Quantifoil R1.2/1.3 Cu300 grid. The grid was then vitrified by manually plunging into liquid nitrogen.

Nano-sized crystals were searched using low-dose high-defocus diffraction mode to minimize radiation damage. For each crystal found, a low-dose trial shot (1 s exposure) was acquired with in-focus diffraction mode to examine whether the crystal diffracted to high-resolution. If so, a custom SerialEM [8] script was executed to record a continuous stream of frames on the DE-64 using in-focus diffraction patterns while the specimen stage was continuously rotated from -50° to $+50^\circ$ at $0.9^\circ/\text{s}$. The frame rate of the DE-64 was set to 20 frames per second (fps) and the total exposure time to 120 seconds. The exposure rate was $0.03 \text{ e}^-/\text{\AA}^2/\text{s}$, yielding a total accumulated dose of about $3.6 \text{ e}^-/\text{\AA}^2$. Indexing and integration was completed using XDS [9], followed by ab initio structure determination and structure refinement using SHELXT and SHELXL [10].

From a single acquisition of one biotin microcrystal, we obtained a 0.7 \AA -resolution map (Fig. 1) with exceptional goodness of fit (Fig. 2). All four types of atoms (C, O, N, S) were assigned correctly and hydrogen atoms were clearly positioned in the refined model as well. These results demonstrate that the

DE-64 direct detection camera may be used to rapidly generate atomic resolution models using MicroED [11].

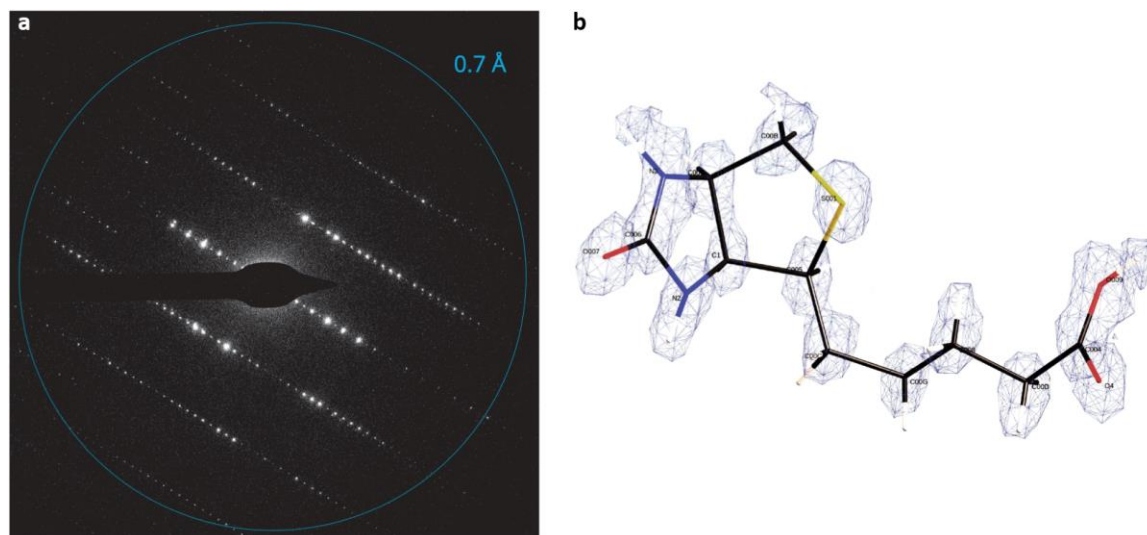


Figure 1. (a) A sum of 60 frames, representing about 3° of total rotation, from the movie collected on the DE-64 direct detection camera during continuous-rotation MicroED. Diffraction spots are clearly visible out to approximately 7 \AA resolution. (b) The resulting density map and atomic model after processing using XDS and SHELX.

Temperature (K)	100
Unit cell lengths (\AA)	5.440(2), 10.640(2), 21.710(4)
Angles ($^\circ$)	90.0(3), 90.0(3), 90.0(3)
Space group	$P2_12_12_1$
Reflections	5031(526)
Unique reflections	1439(162)
R_{obs}	21.5(31.5)
R_{meas}	25.1(37.1)
$CC_{1/2}$	96.5(82.0)
Resolution (\AA)	0.7
Completeness (%)	79.3(86.6)
Exposure ($e^-/\text{\AA}^2$)	~ 3.6
R	0.211
GooF	1.052

Figure 2. The final statistics for the model generated by our MicroED data set of biotin.

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