

Chronic ammonia exposure does not influence hepatic gene expression in growing pigs

Z. Cheng, E. A. O'Connor, Q. Jia, T. G. M. Demmers, C. M. Wathes and D. C. Wathes[†]

The Royal Veterinary College, Hawkshead Lane, Hatfield, Hertfordshire AL9 7 TA, UK

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Housed pigs are often exposed to elevated concentrations of atmospheric ammonia. This aerial pollutant is widely considered to be an environmental stressor that also predisposes to reduced growth rates and poor health, although evidence to support this view is limited. Hepatic gene expression is very responsive to stress and metabolic effects. Two batches of growing pigs were therefore exposed to a nominal concentration of atmospheric ammonia of either 5 ppm (low) or 20 ppm (high) from 4 weeks of age for 15 weeks. Growth rates were monitored. Samples of liver were taken after slaughter (at ~19 weeks of age). Samples from the second batch were analysed for global gene expression using 23 K Affymetrix GeneChip porcine genome arrays. Samples from both batches were subsequently tested for five candidate genes using quantitative real-time PCR (qPCR). The array analysis failed to detect any significant changes in hepatic gene expression following chronic exposure to atmospheric ammonia. Animals clustered into two main groups but this was not related to the experimental treatment. There was also no difference in growth rates between groups. The qPCR analyses validated the array results by showing similar fold changes in gene expression to the arrays. They revealed a significant batch effect in expression of lipin 1 (LPIN1), Chemokine (C-X-C motif) ligand 14 (CXCL14), serine dehydratase (SDS) and hepcidin antimicrobial peptide (HAMP). Only CXCL14, a chemotactic cytokine for monocytes, was significantly down-regulated in response to ammonia. As chronic exposure to atmospheric ammonia did not have a clear influence on hepatic gene expression, this finding implies that 20 ppm of atmospheric ammonia did not pose a significant material risk to the health or metabolism of housed pigs.

Keywords: pigs, ammonia, liver, gene expression, chronic exposure

Implications

Some earlier research suggests that exposing pigs to high concentrations of atmospheric ammonia (≥ 20 ppm) compromises their health. However, this speculation has been based upon extremely brief, unrealistic exposures. Recently, we kept pigs at either ~20 ppm ammonia or in 'fresh' air for 15 weeks (i.e. an entire production cycle) but did not find any evidence of a change in liver function. This implies that pigs can be kept at ~20 ppm ammonia without affecting either their health or metabolism.

Introduction

Housed pigs are routinely exposed to atmospheric ammonia at concentrations that some have long considered are hazardous to their health and/or productivity (e.g. Drummond *et al.*, 1980; Groot Koerkamp *et al.*, 1998). Furthermore,

ammonia is considered a risk factor for the development of lung disease in humans, in particular agricultural workers and cleaners (Omland, 2002; Anderson *et al.*, 2003; Vizcaya *et al.*, 2011). Although some studies have found effects of elevated ammonia on growth and feed conversion efficiency in pigs, the experimental concentrations used often exceeded those found in commercial piggeries, sometimes excessively so (up to 150 ppm Stombaugh *et al.*, 1969; Drummond *et al.*, 1980). With respect to health, experiments have often failed to show changes in lung tissue or growth when pigs were co-exposed to both ammonia and an infectious agent (aerosols of *Escherichia coli*, toxigenic *Pasteurella multocida* or natural exposure to mycoplasma and bacteria; Drummond *et al.*, 1978; Diekman *et al.*, 1993). The long-standing belief that exposure to ammonia is harmful was tested recently in a large experiment involving about 1000 weaner pigs; it showed that exposure for 5½ weeks post-weaning to concentrations of atmospheric ammonia up to ~40 ppm had no effect on either productivity (Wathes *et al.*, 2004) or respiratory disease (Done *et al.*, 2005), although weaner pigs were shown to find

[†] E-mail: dcwathes@rvc.ac.uk

concentrations >20 ppm aversive when given a choice of environments using preference testing (Jones *et al.*, 1999). The former findings are counter-intuitive and imply that there is little reason for farmers or others to worry about atmospheric ammonia in terms of pig production, other than any concerns about the pig's preferences for fresh air.

Recently, we have studied the behavioural and physiological responses of growing pigs exposed continuously to atmospheric ammonia for 15 weeks (O'Connor *et al.*, 2010; Parker *et al.*, 2010). Here protracted, controlled exposure mimics environmental management in pig farming. Although there was evidence that the pigs found 20 ppm of ammonia exposure stressful (they exhibited less play behaviour and appeared to have down-regulated hypothalamic-pituitary axes), there were few effects on important production parameters such as growth and food conversion efficiency as well as general health scores. The higher concentration of ammonia used in this experiment (nominally 20 ppm) is commercially relevant as the current occupational exposure standard in humans is 8 h of exposure to 25 ppm ammonia in any 24 h period (Health and Safety Executive, 2011). This is often taken as a guideline for ammonia exposure of pigs and other housed livestock in the absence of evidence to the contrary: there is currently no legislation regarding maximum atmospheric ammonia in pig housing (Groot Koerkamp *et al.*, 1998).

The liver is an organ, which has a central role in the regulation of carbohydrate, protein and lipid metabolism, immune function, inflammation, hormone metabolism and removal of waste products from the blood (Nemeth *et al.*, 2009). Changes in circulating glucocorticoid levels and cytokines associated with stress and disease are known to influence hepatic function (Ingenbleek and Bernstein, 1999; Chida *et al.*, 2006; Marelli *et al.*, 2010). The liver therefore plays a sentinel role in detecting and responding to factors affecting normal homeostasis. The aim of this experiment was to determine any changes in hepatic gene expression in growing pigs, which were chronically exposed to atmospheric ammonia at a nominal concentration of 20 ppm.

Material and methods

Experimental design

This work was regulated under the United Kingdom Animals (Scientific Procedures) Act 1986. Full details of the experiment can be found in O'Connor *et al.* (2010). Briefly, two batches ($N_{\text{batch}} = 112$) of 4-week-old hybrid gilts (50% White synthetic Pietrain – 25% white Duroc, 12.5% Landrace, 12.5% Large White; PIC, Carlisle, Cumbria, UK) were obtained at weaning from the same commercial indoor pig farm. The first batch was obtained in May and the second in September 2008. On arrival, they were allocated randomly to groups of 14, kept in one of eight experimental rooms (~19.6 m²) in a 2³ fully factorial design and maintained under a combination of either low (control) or high ammonia (nominally 5 or 20 ppm of atmospheric ammonia; low or high mechanical noise (nominally <60 v. 80 dB(A)) and low or

high light intensity (nominally 40 v. 200 lux for 12 h). Only liver samples from the pigs in the control or single-treatment ammonia rooms were used for the current study, so they were also exposed to control levels of light intensity and noise, that is, nominally 200 lux and <60 dB(A). Exposure to the environmental stressors began at the start of the experiment (i.e. on arrival of the pigs at the experimental site) and continued to its conclusion 15 weeks later. Growth parameters were monitored by manually weighing all the pigs once per week using calibrated scales (Pharmweigh Junior, Bury St Edmunds, Suffolk, UK). Health was monitored weekly with respect to nasal and/or ocular discharge, respiratory difficulty, diarrhoea and lameness as described previously (O'Connor *et al.*, 2010).

The pigs were transported to a commercial abattoir at the end of the experiment for slaughter and removal of liver tissues. Small sections (~1 cm²) of each liver were dissected, wrapped in foil, frozen in liquid nitrogen, transferred to pre-labeled bags, transported back to the laboratory on dry ice and stored at –80°C before RNA extraction.

RNA extraction

Total RNA was prepared from 100 mg of fragmented frozen liver tissue of randomly selected pigs from one control room and one ammoniated room in each batch (eight pigs per treatment in Batch 1, seven per treatment in Batch 2, total $n = 30$). Extraction of total RNA was carried out following an ARK Genomics Standard Operating Procedure (<http://www.ark-genomics.org/protocols> IGF100.00: Isolation of total RNA). A two-step extraction method was used, first with Trizol then with a RNeasy mini kit (QIAGEN 74104, Hilden, Germany) for further purification. RNA concentration and purity were determined using a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies Inc., Wilmington, DE, USA), where all samples had a 260/280 ratio of absorbance between 1.8 and 2.1. Samples from Batch 2 ($n = 14$) were used for microarray analysis followed by quantitative real-time PCR (qPCR) on selected genes. Samples from Batch 1 were only used for qPCR.

Microarray hybridisation

All total RNA samples were checked for the concentration, integrity and purity using an Agilent Bioanalyzer according to the manufacturer's instructions (Agilent Technologies Inc., Santa Clara, CA, USA). Microarray hybridisation and data acquisition were carried out by ARK-Genomics (Roslin Institute, University of Edinburgh) using 23 K Affymetrix GeneChip Porcine Genome Arrays based on established ARK-Genomics protocols (<http://www.ark-genomics.org/protocols>).

Microarray data analysis

Data were analysed using an S + ArrayAnalyzer 2.1 built in S-Plus Enterprise Developer 7.0 software package (Insightful Corp., Seattle, Washington, USA). Probe-level expression data generated by the scanner (.CEL files) were imported into the S + ArrayAnalyzer. Each probe set on the Affymetrix array contains 11 pairs of primers to target one gene: as an

initial quality control step any probes with readings for less than seven detected pairs were filtered out. Quality control diagnostics were performed using plots of MvA, Box, RNA degradation and principal component analysis for expression intensity, which confirmed the good quality of all slides. The probe pairs were summarised into a single value per gene using Robust Multichip Analysis and normalised with a median inter-quartile range method.

Principle component analysis of the array data was performed using the BioConductor 2.4 in R-Package. Differences of probes/genes expression between control and ammonia-treated groups were compared using an unpaired *t*-test with Benjamini–Hochberg adjustment for a false discovery rate (FDR). In addition, differences of the normalised expression values between treatment groups were tested using a Significant Analysis of Microarray package (SAM, Stanford University, USA) at a FDR rate of $\alpha=0.05$.

qPCR

Four of the most highly down-regulated genes were selected for follow-up studies by qPCR: lipin 1 (*LPIN1*), Chemokine (C-X-C motif) ligand 14 (*CXCL14*), serine dehydratase (*SDS*) and hepcidin antimicrobial peptide (*HAMP*) plus actin beta (*ACTB*; as a reference gene; see Table 1 for details). These genes were chosen based on the functional information available at the time for the pig genome, suggesting that they might play a role in metabolism and diseases affecting liver. All chemicals and reagents were purchased from Sigma-Aldrich Company Ltd. (Poole, Dorset, UK) or VWR International Ltd. (Poole, Dorset, UK) unless otherwise specified. Optimised qPCR assays were developed, as described previously (Fenwick *et al.*, 2008). Total RNA from each sample was treated for potential genomic DNA carryover in a single reaction in accordance with guidelines supplied by Promega (Promega Corporation, Madison, WI, USA). From this reaction, precisely 1 μg of DNase-treated RNA was reverse transcribed using random hexamer primers and processed accordingly (Reverse Transcription System Kit; Promega). A mastermix of reagents was prepared for the above reaction to minimise potential variation. A negative control included all reagents as above, minus the cDNA template. Gene symbols, sequence information, accession numbers and expected product lengths are provided in Supplementary Table S1.

Gene transcripts were quantified as described in detail previously (Fenwick *et al.*, 2008). Standards for qPCR were prepared from purified PCR products that were quantified by spectroscopy (NanoDrop) and diluted over at least eight orders of magnitude. Briefly, for each assay a mastermix was prepared that contained a final concentration of 10 μl Kapa qPCR SYBR Green Mix (Anachem Ltd, Luton, UK), 500 nM forward and reverse primers and nuclease-free water. Primer annealing and amplicon-specific melting temperatures were determined using the gradient function of CFX-96 Real-time PCR detection system (Bio-Rad Laboratories, Hercules, CA, USA). Equivalent amounts of sample cDNA were added to each reaction in duplicate. To minimise variation, all samples

included in each analysis were derived from the same reverse transcription (RT) batch, prepared under the same conditions and were analysed on a single plate. Thermal cycling conditions applied to each assay consisted of an initial *Taq* activation step at 95°C for 15 min followed by 38 cycles of denaturation (95°C), annealing (range 50.0°C to 64.2°C, see Supplementary Table S1), extension (72°C) and an amplicon-specific fluorescence acquisition reading (range 74°C to 84°C). A melting curve analysis was performed for each amplicon between 50°C and 95°C and as such any smaller non-specific products such as dimers were melted (if present) before fluorescence acquisition. All qPCR results were recorded with the Bio-Rad CFX Manager Software (V1.6, Bio-Rad Laboratories). For comparison of expression data, absolute values were derived from standard curves generated from purified cDNAs identical to amplified products and expressed as fg/ μg reverse-transcribed RNA.

Data analysis

Data from the qPCR analysis were expressed as the mean and root mean square error in fg/1 μg RNA and subjected to ANOVA using SPSS V19 software package (Chicago, IL, USA), including treatment (Control or High Ammonia) and Batch (1 or 2) as factors. Fold changes between groups were calculated as Ammonia/Control when the value of the ammonia group was greater than that of control group and as $-\text{Control/Ammonia}$ when the value of ammonia group was less than that of control group using the values without logarithmic transform. Differences were considered significant at $P<0.05$.

Results

Array analysis

Out of 23 K probes/genes included on the array, around 15 K currently have gene symbol annotations. Overall, only 21 of these genes were down-regulated >1.5 fold and nine were up-regulated >1.5 fold in the 20 ppm ammonia-treated group in comparison with the controls. The fold changes for the top 8 genes in each direction are shown in Table 1. Samples from all 14 pigs were analysed using principal component analysis. This identified two groups of animals, but pigs from the control and high ammonia treatments were equally divided between the two groups (Figure 1). Differences of probes/genes expression between control and ammonia-treated groups were next compared using standard array analysis methods: (i) an unpaired *t*-test with Benjamini–Hochberg adjustment for FDR; and (ii) normalised expression values between treatment groups were tested using SAM at an FDR rate of $\alpha=0.05$. Neither method detected significant differential expression of hepatic genes between treatment groups.

qPCR

Five genes were selected for follow-up studies by qPCR to validate the array results: *ACTB* (reference gene), *LPIN1*,

Table 1 Top eight hepatic genes up- or down-regulated after 15 weeks exposure of growing pigs to atmospheric ammonia (either <5 ppm (control) or ~20 ppm) according to fold changes in expression

Gene symbol	Gene name	Fold change	Brief description of function
Down-regulated			
<i>HAMP</i> ^a	Hepcidin antimicrobial peptide	-3.24	Iron homoeostasis, antimicrobial activity
<i>CYP3A22</i>	Cytochrome P450, subfamily IIIA, polypeptide 22	-2.06	Metabolism of steroids and drugs
<i>SDS</i> ^a	Serine dehydratase	-2.0	Metabolism including conversion of L-serine to pyruvate and ammonia
<i>LPIN1</i> ^a	Lipin 1	-1.94	Triglyceride synthesis
<i>LOC100513945</i>	Enoyl-CoA hydratase domain-containing protein 3, mitochondrial-like	-1.89	Not known
<i>CXCL14</i> ^a	Chemokine (C-X-C motif) ligand 14	-1.82	Cytokine, monocyte chemotaxis
<i>DDO</i>	D-aspartate oxidase	-1.75	Oxidative deamination of D-aspartate and N-methyl-D-aspartate
<i>IGFBP1</i>	Insulin-like growth factor binding protein 1	-1.66	Modulation of IGF bioactivity
Up-regulated			
<i>LOC100153368</i>	Similar to elongation of very long fatty acids-like 2	+3.05	Fatty acid synthesis
<i>F13A1</i>	F13A1 coagulation factor XIII, A1 polypeptide	+2.29	Blood clotting
<i>ZNF133</i>	Zinc finger protein 133	+1.84	Transcription factor
<i>LOC100515931</i>	Protein phosphatase 1 regulatory subunit 3B-like	+1.72	Putative role in glycogen synthesis
<i>NPG1</i>	Protegrin 1	+1.68	Antimicrobial
<i>CLDN1</i>	Claudin 1	+1.59	Component of tight junctions
<i>PROM1</i>	Prominin 1	+1.54	Pentaspans transmembrane glycoprotein
<i>ALAS1</i>	Aminolevulinate, delta-, synthase 1	+1.54	Haeme biosynthesis

^aUsed in follow-up study by qPCR.

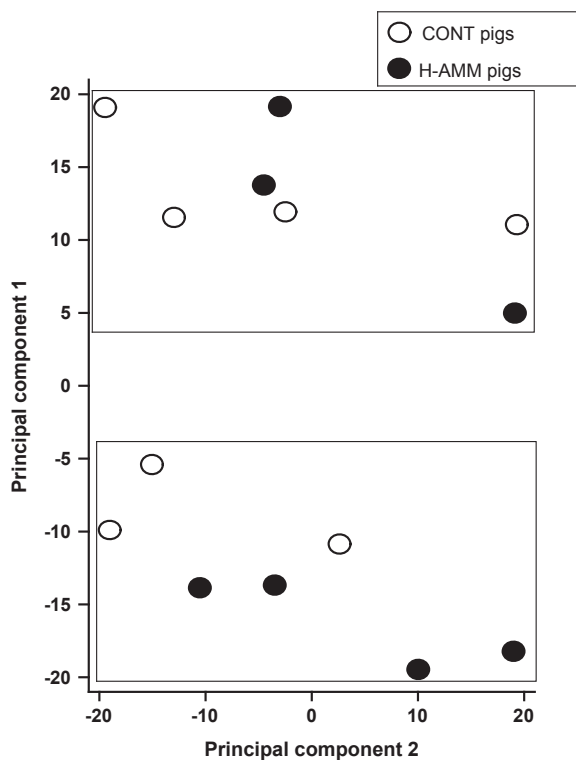


Figure 1 Principal component analysis of hepatic samples from growing pigs exposed to either nominally 5 ppm (CONT) or 20 ppm ammonia (H-AMM), $n = 7$ pigs per treatment. Affymetrix GeneChip porcine arrays. Samples were analysed by 23 K Affymetrix porcine arrays. Two clusters were identified but they did not relate to the treatment.

CXCL14, *SDS* and *HAMP*. The samples from pigs in Batch 2 from the array analysis were repeated and eight additional pigs from each of the control and ammonia-treated groups in Batch 1 were also included. Results are summarised in Table 2. This showed good agreement in the fold changes in expression detected by array and qPCR analysis in Batch 2 but these fold change results were not consistent with those found using qPCR in Batch 1. ANOVA indicated that batch had a significant influence on expression of all of these genes at $P \leq 0.01$ but only one (*CXCL14*) was significantly influenced by ammonia treatment ($P = 0.002$). The reference gene *ACTB* was not altered in response to treatment. As a further check on the data, the expression of the four test genes was calculated as the ratio of expression to *ACTB*. Using this approach, there were no significant differences in expression according to treatment (P all > 0.1).

Weight and growth rates

Data on weights of animals and growth rates have previously been reported for the complete experiment covering 224 pigs (O'Connor *et al.*, 2010). In summary, the mean weight of the pigs increased from 7.8 ± 0.1 kg in 4-week-old pigs at the start of the experiment to 81.9 ± 0.7 kg in the 19-week-old pigs at the end of the experiment. There was a significant difference between batches in the mean weight over the course of the experiment: Batch 1, 36.0 ± 0.2 kg; Batch 2 39.3 ± 0.2 kg, ($P < 0.01$). When batch was controlled for there was, however, no significant effect of any of the

Table 2 Hepatic gene expression measured by qPCR in the growing pig after 15 weeks exposure to atmospheric ammonia (either <5 ppm (control) or ~20 ppm of atmospheric ammonia)

	Batch	Gene ^a				
		<i>ACTB</i>	<i>LPIN1</i>	<i>CXCL14</i>	<i>SDS</i>	<i>HAMP</i>
Control ^b	1	490	24	3.9	2.3	38
High ammonia ^b	1	501	21	2.6	2.5	37
RMSE	1	348.4	11.4	2.3	1.8	32.1
Fold change qPCR	1	1.02	-1.14	-1.51	1.11	1.02
Control ^b	2	1650	126	9.4	24	315
High ammonia ^b	2	1220	59	4.5	13	152
RMSE	2	548.3	68.3	2.7	13.8	235.8
Fold change qPCR	2	-1.35	-2.14	-2.09	-1.79	-2.06
Treatment <i>P</i>	1 + 2	0.22	0.054	0.002	0.16	0.18
Batch <i>P</i>	1 + 2	<0.001	<0.001	<0.001	<0.001	0.003
Treatment × Batch <i>P</i>	1 + 2	0.19	0.076	0.057	0.138	0.186
Fold change array	2	-1.4	-1.94	-1.82	-2.03	-3.24

RMSE = root mean square error; qPCR = quantitative real-time PCR.

Gene expression was analysed using both qPCR and gene expression array.

^aGenes analysed were: *ACTB* = actin beta; *LPIN1* = lipin 1; *CXCL14* = Chemokine (C-X-C motif) ligand 14; *SDS* = serine dehydratase; *HAMP* = hepcidin antimicrobial peptide.

^bValues shown are the means of eight pigs per group in Batch 1 and seven pigs per group in Batch 2 expressed in fg/μg RNA × 10⁶.

Table 3 Comparison of weights and growth rates between growing pigs kept for 15 weeks in either control (<5 ppm) or high ammonia (~20 ppm) atmosphere

Treatment	<i>n</i>	Initial weight (kg) at 4 weeks of age	Final weight (kg) at 19 weeks of age	Growth rate (kg/day)
Control	28	8.0 ± 0.2	80.6 ± 1.1	0.8 ± 0.1
Ammonia	28	8.0 ± 0.2	80.8 ± 1.1	0.8 ± 0.1
RMSE		1.6	8.1	0.3
<i>P</i> -value		0.89	0.88	0.89

RMSE = root mean square error.

Values are expressed as mean ± s.e.m.

treatments on the mean weight or growth trajectories of the pigs. The average data for the animals in the control and ammonia-treated groups are summarised in Table 3.

Discussion

The results of this study showed little or no significant hepatic response of growing pigs after 15 weeks of exposure to 20 ppm of atmospheric ammonia. Using standard analytical methods for gene expression arrays, there were no significant changes in expression. This was surprising for two reasons. First, array studies are able to detect small changes in gene expression across the whole genome. The experiment's design meant that it was able to detect a fold change of 1.25 between two samples, at $\alpha = 0.05$ and a power >0.8 in a test of the experimental hypothesis with an unpaired *t*-test and making the usual assumptions about heteroscedasticity, random allocation of experimental subjects, etc.

Our results showed an extremely good relationship between the fold changes in gene expression measured by the arrays and qPCR. This provides further confidence in the validity of the results.

Second, the liver is a central organ, which responds to stress, metabolic changes and disease, all of which might reasonably have been expected to change in response to the ammonia treatment (Berczi, 1998; Chida *et al.*, 2006). The biological functions of the liver include detoxification of harmful substances in the blood, glycogen storage, protein synthesis, bile secretion and hormone synthesis (Nemeth *et al.*, 2009). In the context of this experiment, it is unlikely that uptake of the atmospheric ammonia would have been sufficient to produce any measurable change in plasma ammonium levels. Instead it might have caused lung disease (Done *et al.*, 2005) or caused the animals stress through living in an environment which they could find aversive (Jones *et al.*, 1999). Using array analysis, we have previously detected many significant changes in hepatic gene expression in response to negative energy balance in cows (McCarthy *et al.*, 2010) and transport stress in broiler chickens (Sherlock *et al.*, 2012). Similarly, lesions of the foot and hock induced by ammonia treatment of the litter resulted in differential expression of 417 hepatic genes in broiler chickens (Sherlock *et al.*, 2012). Others have successfully used expression profiling to monitor responses to calorie restriction in prepubertal pigs (Lkhagvadorj *et al.*, 2010). Therefore, the lack of differentially expressed genes in liver of pigs exposed to ammonia suggests that a myriad of metabolic processes were apparently unaffected by this treatment.

The pigs used in this experiment were part of a larger study into the effects of three environmental stressors (ammonia, noise and low light). This found no significant

difference in the overall activity levels or growth rates of pigs housed in either high-ammoniated or low-ammoniated environments, although, in common with the results on gene expression reported here, there was a significant batch effect (O'Connor *et al.*, 2010; Parker *et al.*, 2010). In particular, the mean weight of the pigs over the course of the experiment was higher in Batch 2, although both sets of animals came from the same source and genetic stock and were housed in the same building. After controlling for the batch effect, the pigs in the ammoniated rooms had slightly lower salivary cortisol, larger adrenal cortices and performed less play in week 3 of the experiment, although there were no differences in play behaviour in weeks 8 or 14. As liver samples were only collected at the end of the experiment, it remains possible that changes in hepatic gene expression may have occurred earlier but that the pigs adapted to cope with this environment over time. However, we were interested in chronic not acute changes and from both a commercial and welfare perspective the net effect, integrated over an animal's lifetime, may be more important.

We selected four genes in addition to the reference gene *ACTB* for further analysis, which showed slightly lower hepatic expression in the ammonia-treated pigs: *SDS*, *LPIN1*, *HAMP* and *CXCL14*. Although their changes in expression were not statistically significant, the functions of the proteins derived from these genes suggested that they might plausibly have been influenced by the treatment. Serine dehydratase is one of three enzymes involved in metabolising serine and glycine. The protein is found mainly in liver where L-serine dehydratase converts L-serine to pyruvate and ammonia (Ogawa, 2000). Lipin 1 is involved in adipose tissue development and triglyceride metabolism (Reue and Brindley, 2008). In humans, *LPIN1* is a candidate gene for lipodystrophy, a disease that is characterised by loss of body fat, fatty liver, hypertriglyceridemia and insulin resistance. *HAMP* is produced by the liver and plays an important role in the maintenance of iron homeostasis. The gene is up-regulated during inflammation, potentially resulting in anaemia (Deicher and Hörl, 2006). Chemokine (C-X-C motif) ligand 14 is an immunoregulatory cytokine, which displays chemotactic activity for monocytes (Shurin *et al.*, 2005).

Out of these genes, only *CXCL14* showed a significant effect of ammonia treatment across both batches of pigs ($P < 0.001$), although there was also a trend ($P < 0.054$) for *LPIN1*. There has been limited previous work investigating hepatic *CXCL14* expression using murine models. It is up-regulated in response to hepatic injury (De Minicis *et al.*, 2007; Li *et al.*, 2011) and schistosome infection (Burke *et al.*, 2010) and is thought to attract monocytes, promote fat deposition, inhibit hepatocyte proliferation and contribute to the formation of hepatic granulomas and fibrosis. Hepatic down-regulation, as seen here in response to atmospheric ammonia, would presumably decrease such responses, although the relevance of this observation is uncertain without further investigation.

Overall, our results did not support the hypothesis that chronic exposure to ammonia would be reflected in changes

in hepatic gene expression. This suggests that growing pigs can adapt to this level of environmental exposure without affecting their production efficiency or health. Such levels of ammonia do, however, raise cause for concern with regard to welfare as pigs are known to find them aversive (Jones *et al.*, 1999).

Conclusions

Despite traditional beliefs about the putative effects of exposure to atmospheric ammonia on the livestock performance, this experiment has not provided any scientific evidence that chronic exposure to a nominal concentration of 20 ppm affects hepatic gene expression in growing pigs. Other studies of respiratory disease have also failed to find an effect.

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Supplementary material

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S1751731113002127>

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