

## Prevalence and growth kinetics of Shiga toxin-producing *Escherichia coli* (STEC) in bovine offal products in Japan

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### SUMMARY

Recent epidemiological data suggest a link between the consumption of bovine offal products and Shiga toxin-producing *Escherichia coli* (STEC) infection in Japan. This study thus examined the prevalence of STEC in various types of these foods. PCR screened 229 bovine offal products for the presence of Shiga toxin (*stx*) gene. Thirty-eight (16·6%) samples were *stx* positive, of which eight were positive for *rfbE*<sub>O157</sub> and three were positive for *wzy*<sub>O26</sub>. Four O157 and one O26 STEC isolates were finally obtained from small-intestine and omasum products. Notably, homogenates of bovine intestinal products significantly reduced the extent of growth of O157 in the enrichment process compared to homogenates of beef carcass. As co-incubation of O157 with background microbiota complex from bovine intestinal products in buffered peptone water, in the absence of meat samples, tended to reduce the extent of growth of O157, we reasoned that certain microbiota present in offal products played a role. In support of this, inoculation of generic *E. coli* from bovine intestinal products into the homogenates significantly reduced the extent of growth of O157 in the homogenates of bovine intestinal and loin-beef products, and this effect was markedly increased when these homogenates were heat-treated prior to inoculation. Together, this report provides first evidence of the prevalence of STEC in a variety of bovine offal products in Japan. The prevalence data herein may be useful for risk assessment of those products as a potential source of human STEC infection beyond the epidemiological background. The growth characteristic of STEC O157 in offal products also indicates the importance of being aware when to test these food products.

**Key words:** Background microbiota, bovine offal products, growth kinetics, Shiga toxin-producing *Escherichia coli* (STEC).

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## INTRODUCTION

Shiga toxin (Stx)-producing *Escherichia coli* (STEC) is a foodborne enteric pathogen that causes human illness ranging from mild diarrhoea to life-threatening haemolytic uraemic syndrome (HUS) [1]. Epidemiological studies have shown that STEC can potentially enter the human food chain from a number of animal sources; the most common sources are unpasteurized dairy products and meats cross-contaminated with faeces or intestinal contents after slaughter [2].

Because most causes of STEC outbreaks in the USA have been traced to beef containing the STEC serotype O157:H7 [3], epidemiological studies have focused on the prevalence of this serotype in beef and beef cattle. However, additional STEC serotypes, including serotypes O26, O91, O103, O111, O118, O145, and O166, have been isolated from beef and have caused human illnesses worldwide [4]. In Japan, the dominant serotype associated with human STEC infection is O157:H7, followed by O26:H11 and O111:H<sup>-</sup>, with these three accounting for more than 95% of human STEC infections [5]. Of the 373 STEC serotypes isolated from cattle faeces or hides, 65 were identical to those detected in HUS patients, and 62 are known to cause other human illnesses [6]; these data highlight the fact that cattle and bovine food products are important reservoirs for human STEC infection.

Notably, in some Asian countries, it is customary to eat bovine offal products including liver, heart, and intestines; some are consumed raw, as so-called *sashimi*. Recent epidemiological data have indicated that of the 28 foodborne STEC O157 outbreaks in which their causative foods were identified in Japan during 2007–2010, 11 (39%) outbreaks were caused by the consumption of these bovine offal products [7]. These cases suggest that such food products might be one of the major sources of foodborne STEC infection in Japan. Despite this understanding, contamination rates and doses of STEC in those food products along their supply chain remain unclear, although a small-scale detection study has been reported [8].

In the present study, we examined the prevalence of STEC (particularly serotypes O157, O26, and O111) in various types of bovine offal products that are widely distributed in Japan. Throughout the comparative assessment of the growth kinetics of STEC, we found that these food products significantly affected the growth of STEC during enrichment.

Table 1. Prevalence of stx and O157, O26, O111 serotype-specific genes in bovine internal organ foods

Name	No. tested	No. stx-positive (%)	No. positive		
			<i>rfbE</i> <sub>O157</sub>	<i>wzy</i> <sub>O26</sub>	<i>rfb</i> <sub>O111</sub>
Heart	6	2 (33.3)	—	—	—
Liver	36	5 (13.9)	1	—	—
Rumen	21	5 (23.8)	1	—	—
Reticulum	22	6 (27.3)	—	—	—
Omasum	38	4 (10.5)	2	1	—
Abomasum	24	2 (8.3)	—	—	—
Small intestine	54	10 (18.5)	3	1	—
Large intestine	22	4 (18.2)	1	1	—
Others*	6	0 (0.0)	—	—	—
Total	229	38 (16.6)	8	3	0

\* Others represent kidney ( $n=2$ ) and mixed intestinal products ( $n=4$ ).

Therefore, we investigated background microbiota from the offal products and their effects on the growth of STEC.

## MATERIALS AND METHODS

### Food samples

A total of 229 raw bovine offal products that are commercially distributed throughout Japan via the internet were purchased from July to November 2010 (Table 1). These samples were delivered to the laboratory by rapid, cold road transport and were processed on arrival. These samples included liver ( $n=36$ ), rumen ( $n=21$ ), reticulum ( $n=22$ ), omasum ( $n=38$ ), abomasum ( $n=24$ ), small intestine ( $n=54$ ), large intestine ( $n=22$ ), heart ( $n=6$ ), and others (two kidney and four mixed intestinal products).

### Culture enrichment and DNA extraction

A 25-g portion of each raw sample was diluted with 225 ml buffered peptone water (BPW) (pH 7.4) (Oxoid, UK) in a Stomacher bag (Eiken Kagaku, Japan), followed by homogenization for 1 min at 5.0 strokes/s using a Lab blender (IUL Instruments, Spain). After incubation at 42 °C for 20 h, a 1 ml aliquot of the enrichment broth was centrifuged at 21 500  $g$  for 5 min, and the cell pellets were re-suspended in 100  $\mu$ l PrepMan Ultra solution (Life Science Technology Inc., USA) to extract DNA according to the manufacturer's instructions.

### PCR detection for *stx* and serotype-specific genes

The DNA extract prepared as described above was subjected to PCR to detect the Shiga toxin (*stx*) gene as described previously [9]. The *stx*-positive samples were further subjected to PCR to detect the following genes: *rfbE*<sub>O157</sub> [10], *wzy*<sub>O26</sub> [11], and *rfb*<sub>O111</sub> [12], which can specifically detect serotypes O157, O26, and O111, respectively.

### Isolation and characterization of STEC from bovine offal products

Bacterial cultures that were PCR positive for any of the three serotype-specific genes were subjected to immunomagnetic separation using Dynabeads specific for O157 or O26 (DynaL, Norway) (according to the manufacturer's instructions) to isolate the target serotype bacteria. The following selective agar media were used: KBM STEC Chrom agar (Kojin Bio, Japan) and cefixime tellurite-sorbitol MacConkey (CT-SMAC) agar (Eiken Kagaku) for O157 and KBM STEC Chrom agar (Kojin Bio) and cefixime tellurite-rhamnose MacConkey (CT-RMAC) agar (Eiken Kagaku) for O26. The STEC isolates that were finally obtained were serotyped and genetically characterized for *stx*, *eaeA*, *EhlyA* and *uidA* as previously described [13]. *Stx* production was assayed with VTEC-RPLA (Denka Seiken, Japan).

### Construction of kanamycin (Km)-resistant STEC O157

The plasmid pWM1007, which contains *kan* (Km-resistant) loci [14], was introduced into STEC O157 strains 204, 466, and 470 by electroporation. Successful transformants (designated 204-Km, 466-Km, 470-Km, respectively) (Table 2) were selected on CT-SMAC agar containing 50 µg/ml Km (CT-SMAC-Km).

### Viable cell count of STEC O157 in bovine food homogenates

The STEC O157 strain 204-Km was grown in LB medium supplemented with 50 µg/ml Km (LB-Km) at 37 °C for 20 h. After being washed with sterile PBS twice, 100 µl each of the serially diluted suspension were separately used to inoculate 5-g portions of short-loin beef, liver, and small-intestine products in a Stomacher bag (Eiken Kagaku). Following suspension in 45 ml BPW, homogenization was performed

in a Lab blender (IUL Instruments) at 5.0 strokes/s for 1 min. The homogenates were then incubated at 42 °C for 20 h. The numbers of viable O157 cells were determined by plating the homogenates onto CT-SMAC-Km at 0 h and 20 h post-incubation. To exclude strain-specific growth characteristics, we separately inoculated three O157 strains (204-Km, 466-Km, 470-Km) in bovine small-intestine products and their growth through BPW enrichment was also measured by plate count.

### Measurement, isolation, and characterization of background microbiota in bovine offal

A 5-g portion of small-intestine product and short-loin beef was homogenized (1 min, 5.0 strokes/s) in 45 ml BPW as described above, and the serial dilutions were immediately plated on Nutrient agar (NA) plates (Eiken Kagaku) to determine aerobic plate counts (APCs) by incubation at 37 °C for 22 h, or MRS agar (BD Biosciences, USA) to measure the number of lactic acid bacteria (LAB) by anaerobic incubation using GasPak (BD Biosciences) at 37 °C for 48 h as described previously [15]. The remainder of the homogenates were thereafter incubated at 42 °C for 20 h, and also subjected to the same measurement of bacterial counts. To rule out bacteria that predominantly grew in the homogenates after incubation, six representative colonies grown on the NA and MRS plates from the 20-h culture were separately chosen, and their 16S rRNA sequences were determined by cycle sequencing using the primers 5'-CAGGCCTAACACATGCAAGTC-3' and 5'-GGGGGGTGTACAAGGC-3', in an ABI3730xl DNA analyser (Life Science Technology) according to the manufacturer's instructions. In the case of complete sequence similarity in the isolates from NA plates, one representative isolate was biochemically characterized with the Api-20E kit (bioMérieux, France) and used in the following studies.

### Competitive growth assay

#### *Co-incubation with background microbiota complex*

A 5-g portion of short-loin beef or small-intestine samples was suspended in 45 ml BPW and vortexed for 60 s. Subsequently, the suspensions were filtered through a 20-µm-diameter filter (Millipore, USA) to remove the majority of the microbes from the food matrices. The filtrates were then centrifuged for

Table 2. *Bacterial strains used in the study*

Strain name	Taxonomy*	Source	<i>E. coli</i> serotype	Genotype				
				<i>stx1</i>	<i>stx2</i>	<i>eaeA</i>	<i>uidA</i>	<i>EhxA</i>
<b>STEC isolates</b>								
022-1	<i>E. coli</i>	Small intestine	O157:H7	–	+	+	+	+
066-1	<i>E. coli</i>	Small intestine	O157:H7	–	+	+	+	+
068-1	<i>E. coli</i>	Omasum	O26:H11	+	–	+	–	+
HG16	<i>E. coli</i>	Omasum	O157:H7	+	–	+	+	+
HG45	<i>E. coli</i>	Small intestine	O157:H21	–	–	+	+	+
<b>Background microbiota</b>								
SI-1	<i>E. coli</i> (AB609040)	Small intestine	OUT:H21	–	–	–	–	–
SI-2	<i>Aeromonas</i> sp. (AB609041)	Small intestine	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
SI-3	<i>E. coli</i> (AB609042)	Small intestine	OUT:HUT	–	–	–	–	–
SI-4	<i>Enterococcus</i> sp. (AB609043)	Small intestine	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
SI-7	<i>E. coli</i> (AB609044)	Small intestine	OUT:H16	–	–	–	–	–
LAB-1	<i>Lactobacillus</i> sp. (AB609049)	Small intestine	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
LAB-2	<i>Lactobacillus sakei</i> (AB609050)	Small intestine	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
LAB-3	<i>Lactobacillus</i> sp. (AB609051)	Small intestine	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
B-1	<i>Hafnia alvei</i> (AB609045)	Short-loin beef	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
B-4	<i>Enterobacter</i> sp. (AB609046)	Short-loin beef	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
B-6	<i>Yersinia enterocolitica</i> (AB609047)	Short-loin beef	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
B-9	<i>Hafnia alvei</i> (AB609048)	Short-loin beef	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.
<b>STEC O157 strains for spike experiments (harbouring plasmid pWM1007)</b>								
206-Km	<i>E. coli</i>	Human	O157:H7	–	+	+	+	+
466-Km	<i>E. coli</i>	Liver	O157:H7	+	+	+	+	+
470-Km	<i>E. coli</i>	Small intestine	O157:H7	–	+	+	+	+

n.t., Not tested.

All *E. coli* were genotyped for the presence (+/–) of *stx*, *eaeA*, *uidA*, and *EhxA* genes, and were O- and H-serotyped.

\* Taxonomy of the background microbiota were based on the 16S rRNA sequences (GenBank accession numbers given in parentheses). Km-resistant STEC O157 strains 204-Km, 466-Km, 470-Km were constructed by introducing the plasmid pWM1007.

15 min at 2150 g, and the bacterial pellets were carefully washed and resuspended in 45 ml fresh BPW. About  $1.5 \times 10^3$  cells of STEC O157 strain 204-Km were then used to inoculate background microbiota-containing BPW suspensions in a Stomacher bag and these cultures were then incubated at 42 °C for 20 h. After incubation, the growth of STEC O157 was determined by plate counts on CT-SMAC-Km.

#### *Co-incubation with representative microbiota in food homogenates*

A 5-g portion of short-loin beef or small-intestine products was added to 45 ml BPW, homogenized as described above in a Stomacher bag, followed by incubation for 30 min at 63 °C (heat+) to kill most bacteria, or kept at room temperature (heat–) not to kill bacteria (we confirmed that following the above

heating, both types of food homogenates contained  $< 10$  c.f.u./ml bacteria, enumerated by colony counts on NA agar). After cooling down to room temperature, the food homogenates were inoculated with  $8.6 \times 10^2$  cells of O157 strain 204-Km and co-cultured with approximately  $10^2$  c.f.u. orders of representative background microbiota from small-intestine products (SI-1, -2, -3, -4, and LAB-2) at  $42^\circ\text{C}$  for 20 h. The growth of O157 cells was determined by plate count on CT-SMAC-Km. The food homogenates spiked only with O157 culture (without microbiota) were used as controls.

### Nucleotide sequence data

The 16S rRNA sequences from the representative background microbiota were deposited in GenBank under accession numbers AB609039–AB609051.

### Statistical analysis

Data for the plate counts represent means  $\pm$  standard deviation from three sets in two independent experiments unless otherwise indicated. Statistical significance was calculated by Student's *t* test ( $P < 0.05$ ).

## RESULTS

### Prevalence of STEC-related genes in bovine offal products in Japan

From July to November 2010, 229 bovine offal products that were commercially distributed throughout Japan were subjected to testing for the prevalence of STEC. PCR screening detected the *stx* gene in 38 (16.6%) samples. These samples included heart (33.3%), liver (11.1%), rumen (23.8%), reticulum (27.3%), omasum (10.5%), abomasum (8.3%), small intestine (18.5%), large intestine (18.2%), and others (0%) (Table 1). Of these samples, eight were positive for *rfbE*<sub>O157</sub>, and three were positive for *wzy*<sub>O26</sub>. One omasum sample was positive for both *rfbE*<sub>O157</sub> and *wzy*<sub>O26</sub> (Table 1). The *rfbO*<sub>111</sub> gene was not detected in any of the samples tested.

### Isolation and characterization of STEC O157 and O26

Of the 10 samples that were positive for *rfbE*<sub>O157</sub> or/and *wzy*<sub>O26</sub>, immunomagnetic separation in combination with the selective media enabled us to isolate three O157:H7, one O157:H21, and one O26:H11

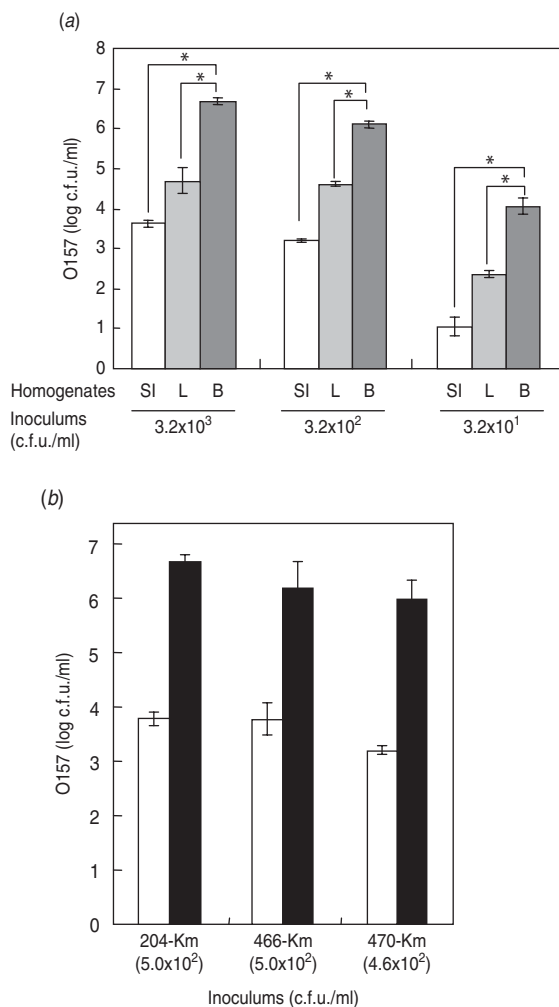
STEC from small-intestine and omasum products (Table 2). All STEC strains except the strain HG45 (serotype O157:H21) produced cytotoxic Shiga toxins, and their *stx* genotypes consisted of *stx1* alone ( $n=2$ ), and *stx2* alone ( $n=2$ ) (Table 2). They all possessed the *EhxA* gene, and the *uidA* gene was detected in all O157 isolates (Table 2).

### Type of food matrix that affects the growth kinetics of STEC O157

Having little information on the enrichment efficiency of the bovine offal products for STEC bacteria, we experimentally used the Km-resistant STEC O157 strain 204-Km to inoculate samples of short-loin beef, liver, and small intestine, followed by homogenization. After incubation of the homogenates, the growth of O157 cells was determined. The short-loin beef homogenates allowed the STEC bacteria to grow to a concentration ranging between  $1.1 \times 10^7$  and  $2.8 \times 10^7$  c.f.u./ml when inoculated with  $2.5 \times 10^3$  c.f.u. of STEC O157 strains (Fig. 1*a*). However, the inoculums were not significantly increased in the homogenates of small-intestine products, exhibiting only a 3.2- to 4.8-fold increase ( $8.1 \times 10^3$  to  $1.2 \times 10^4$  c.f.u./g) (Fig. 1*a*). The growth of O157 cells in the liver sample homogenates was intermediate;  $5.9 \times 10^4$  to  $3.2 \times 10^5$  c.f.u./g of STEC O157 were recovered under the same conditions (Fig. 1*a*). The reduced extent of STEC O157's growth in the homogenates of bovine small-intestine products was confirmed not to be a strain-dependent characteristic because three different O157 strains exhibited similar trends towards O157 growth (Fig. 1*b*). Taken together, the data demonstrated that homogenates of bovine digestive-tract products significantly reduced the extent of STEC O157 growth in the enrichment process compared to homogenates of loin beef and bovine liver products.

### Background microbiota in bovine intestinal products plays a role in affecting STEC O157 growth in BPW

Due to the significant effect of the homogenates of small-intestine products on STEC O157 growth, we assumed that background microbiota in the homogenates might play a role in reducing the extent of growth of STEC bacteria in BPW. To examine this issue, we first isolated the background microbiota complexes from short-loin beef or bovine small-intestine products, using filtration-based isolation of the background microbiota; this enabled us to collect



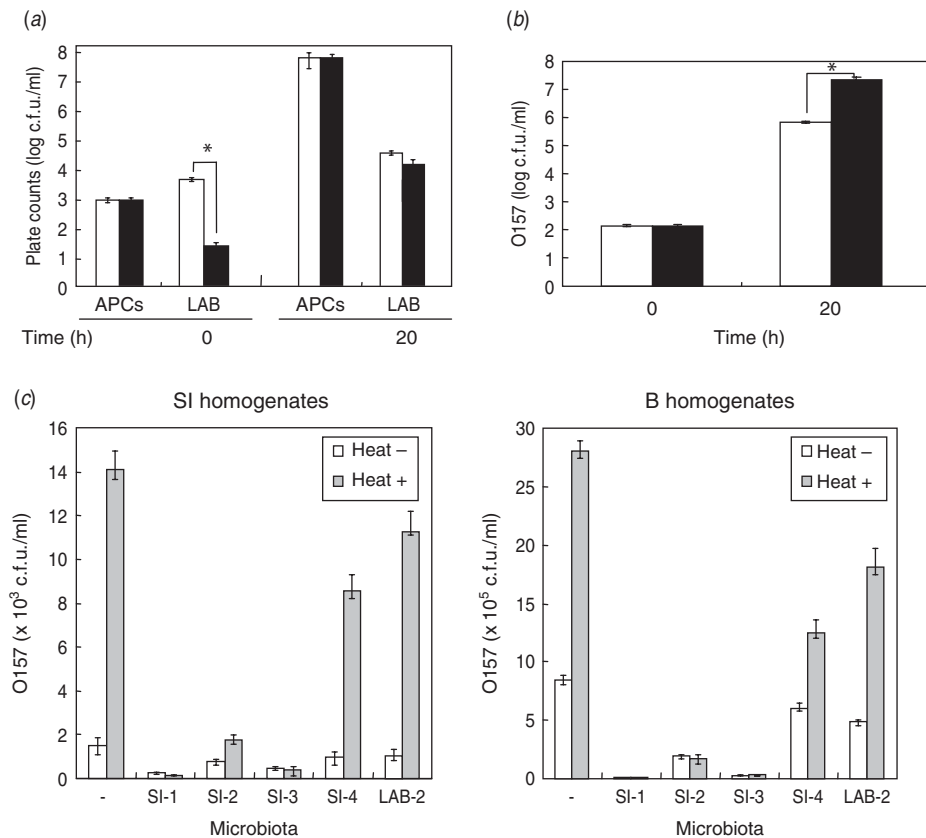
**Fig. 1.** Food matrices of bovine digestive-tract products significantly affect the growth of STEC O157 during the enrichment process. (a) Different numbers of STEC O157 strain 204-Km ( $3.2 \times 10^3$ ,  $3.2 \times 10^2$ ,  $3.2 \times 10^1$  c.f.u.) were inoculated into 5-g portions of bovine products (SI, small intestine; L, liver; B, short-loin beef) and homogenized in BPW. After incubation, the STEC O157 cells were counted on CT-SMAC-Km. (b) Three STEC O157 strains (204-Km, 466-Km, 470-Km) were used to inoculate bovine small-intestine ( $\square$ ) and short-loin beef ( $\blacksquare$ ) products and homogenized in BPW. In panels (a) and (b), means represent viable numbers of O157 (c.f.u./ml), and error bars represent standard deviation. Asterisks indicate statistical significance ( $P < 0.05$ ).

approximately  $4.8 \times 10^3$  or  $5.8 \times 10^3$  cells of the microbiota from 5-g short-loin beef and small-intestine products, respectively, which corresponded to 40% and 44.6%, respectively, of the APCs that were counted by direct plating of the homogenates ( $1.2 \times 10^4$  or  $1.3 \times 10^4$  cells, respectively) (Fig. 2a). Through the co-incubation with loin beef-originated

background microbiota complex in the absence of food homogenates, the O157 strain 204-Km grew to a concentration ranging from  $2.8 \times 10^7$  to  $4.5 \times 10^7$  c.f.u./ml BPW medium (Fig. 2b). In comparison, co-incubation with background microbiota from small-intestine products allowed STEC cells to grow only to  $7.7 \times 10^5$  c.f.u./ml, which was about 30-fold less than from co-incubation with short-loin beef-derived background microbiota (Fig. 2b). These data thus suggested that the background microbiota complex in small-intestine products played a role in affecting the growth of STEC O157 in BPW.

### Bovine intestinal products contain different composition of background microbiota than short-loin beef products

Given the different effects of background microbiota from the loin-beef and bovine small-intestine products on the growth of O157 under the experimental co-culture conditions, we next examined the population and composition of APCs, and LAB in the homogenates of bovine short-loin beef and small-intestine products before and after incubation. Between the two types of the homogenates, APCs were not significantly different throughout the enrichment ( $1.2 \times 10^3$  and  $1.3 \times 10^3$  c.f.u./ml at pre-incubation, and  $7.8 \times 10^7$  and  $7.1 \times 10^7$  c.f.u./ml at 20 h post-incubation, respectively) (Fig. 2b). Moreover, the homogenates of small-intestine products contained more LAB compared to the short-loin beef sample homogenates at pre-incubation ( $6.8 \times 10^3$  and  $4.2 \times 10^1$  c.f.u./ml,  $P < 0.05$ ), whereas both were increased to almost equal levels after incubation (Fig. 2b). Through genetic and biochemical characterization, we subsequently identified representative bacterial colonies predominantly grown after incubation; the microbiota in small-intestine products consisted of *E. coli*, *Aeromonas* spp., and *Enterococcus faecium* (Table 2). In comparison, no *E. coli* were isolated from the short-loin beef homogenate sample, which instead contained *Hafnia alvei*, *Enterobacter* spp., and *Yersinia enterocolitica* (Table 2). In addition, representative LAB isolates from the homogenates of small-intestine products were genetically identified as *Lactobacillus* spp. and *L. sakei* (Table 2). These findings indicated that microbiota that grew predominantly in homogenates of bovine small-intestine products were compositionally different from those in the homogenates of short-loin beef.



**Fig. 2.** Background microbiota affects the growth of STEC O157 during enrichment. (a) Aerobic plate counts (APCs) and lactic acid bacteria (LAB) counts in homogenates of small-intestine (□) and short-loin beef (■) products were counted before (0 h) and after (20 h) incubation. (b) Background microbiota complex was isolated from small-intestine (□) or short-loin beef products (■), then co-incubated with O157 strain 204-Km in BPW in the absence of food homogenates. At 0 h and 20 h post-incubation, numbers of STEC O157 were counted. Asterisks in section panels (a) and (b) indicate statistical significance ( $P < 0.05$ ). (c) Homogenates of small-intestine products (left panel) or short-loin beef (right panel) were treated with heat (heat+) for 30 min or untreated (heat-). These food sample homogenates were then inoculated with O157 strain 204-Km and representative microbiota from small-intestine products (SI-1, -2, -3, -4, and LAB-2). After incubation, the growth of O157 was determined. Food sample homogenates spiked only with O157 (with no spiked microbiota) were used as control (-). Error bars in panels (a)–(c) indicate standard deviation.

**Generic *E. coli* reduces the extent of STEC O157 growth in BPW homogenates**

Having established that the background microbiota complex from bovine small-intestine products significantly reduced the extent of growth of O157 in BPW in the absence of food sample homogenates, and in which different composition of microbiota were predominantly grown compared to the homogenates of short-loin beef, we next examined the effect of representative microbiota isolates on STEC O157 growth by co-incubation with the food homogenates. Prior to incubation, the homogenates of small-intestine and short-loin beef products were treated with or without heating (63 °C, 30 min). These homogenates were then spiked with STEC O157 and equal orders of microbiota isolates from small-intestine products,

following by incubation for 20 h. In all homogenates, the heat treatment increased the growth of O157 compared to unheated homogenates (Fig. 2c); in the absence of spiked microbiota, the heat treatment increased the number of O157 by 9.2-fold in small-intestine homogenates and by 3.3-fold in loin-beef homogenates, respectively (Fig. 2c). Of homogenates spiked with microbiota, those with generic *E. coli* microbiota (SI-1, SI-3) allowed less growth of O157 cells in small-intestine and loin-beef homogenates (Fig. 2c). Spiking with LAB (LAB-2) did not produce any marked influence on the growth of O157 in either type of homogenate, compared to generic *E. coli* (Fig. 2c). Taken together, we were able to demonstrate that generic *E. coli* was one of the predominant microbiota in reducing the extent of STEC O157 growth in the homogenates during the enrichment process.

## DISCUSSION

The types of food associated with STEC outbreaks and the geographical distribution of cases differ between countries. These differences have originated from local food preferences, culinary customs, and patterns of food distribution. Recent epidemiological data in Japan have revealed that consuming bovine offal products seems to be linked to the occurrence of STEC infection [7]. Therefore, for the first time, this study examined the prevalence of STEC in various types of bovine offal products.

The efficacy of a detection method depends on the choice of the base medium, selective agents, and their concentrations. The interactions between these factors are also expected to affect the sensitivity of the detection method, especially when the test sample contains a small number of STEC cells. After slaughter, bovine offal products are often treated with chlorine, and it is likely that such sanitizing damages or kills many of the STEC bacteria [16]. Thus, it is conceivable that a portion of STEC in the test samples might have been damaged by chlorination. A comparative evaluation study on the recovery medium for STEC O157, O26, and O111 showed that BPW could efficiently recover damaged O157 bacteria culture, while tryptone soya and EC broths, and supplementation with vancomycin and novobiocin prevented bacterial growth [17, 18]. Further, BPW without additives was superior to BPW with additives for the isolation of STEC O157 by immunomagnetic separation from bovine faecal samples [19]. Therefore, we selected BPW without supplements as the enrichment media.

Many studies have focused the prevalence of STEC O157 in cattle faeces, hides, trim, or ground beef because of their epidemiological importance to public health [6]. In this study, the *stx* gene was detected from a variety of bovine offal products. However, it is difficult to conclude that these rates reflect the contamination rates of STEC in these products because of the different numbers tested. Conversely, successful isolation of STEC O157 and/or O26 only from digestive-tract products including small-intestine and omasum (Table 2) suggested that these products might be contaminated with high populations of STEC pathogens, compared to the others. Most O157 STEC strains relating to human infection possess H7 flagellin or no flagellin (H<sup>-</sup>), while the serotype O157:H21 isolated from bovine small-intestine products in this study appears to be rare and

the association with human infection is unknown, because no report exists for the isolation of this serotype from beef cattle and their products [6].

We repeatedly subcultured the samples that were positive for *stx* and either *rfbE*<sub>O157</sub> or *wzy*<sub>O26</sub>, in different enrichment media for the isolation of STEC. Nevertheless, some of the STEC were not isolated. Possible explanations for this result include the following: the STEC had already died, the STEC were minimally damaged during storage and transport of these products, or the growth of STEC was competitively inhibited by certain microbes present in the food samples during the processes of storage and enrichment.

Our focus on the growth kinetics of STEC in the food matrix during the process of enrichment thereby revealed that bovine small-intestine homogenates greatly reduced the extent of STEC O157 growth compared to short-loin beef homogenates. Consequently, we are able to demonstrate that in a variety of background microbiota, generic *E. coli* predominantly played a role in reducing the extent of STEC growth in the homogenates of bovine offal products. Although we also found higher levels of LAB in small-intestine products compared to that in short-loin beef, co-incubation with LAB weakly affected STEC O157 growth relative to generic *E. coli* and others in the food sample homogenates (Fig. 2c). In addition, the numbers of LAB after incubation were not significantly different between the two types of food products and both reached only around 10<sup>4</sup> c.f.u./ml (Fig. 2b), further supporting our notion that LAB might not be the predominant microbiota to affect the growth of STEC in the enrichment process. Even with heat pretreatment to kill most microbiota, O157 still grew more in homogenates of short-loin beef than in homogenates of bovine intestinal products (Fig. 2c). This suggests the additional factors in the food matrices also influence the growth of O157. Although the bovine intestinal products contained greater amounts of fat content than the loin beef products (data not shown), a recent work by Bosilevac *et al.* reported that increased fat content did not interfere with recovery of STEC O157 from beef carcasses [20]. Further exploration for such factors would be valuable to improve the isolation method of STEC from bovine offal products.

The population of STEC may also be affected by the processing and storage conditions of the food products. A previous study reported that feedlot cattle are infected with O157 at a level of about log



1·17 MPN/g, and some cattle exhibit a ~1000-fold increase at that stage of slaughter (log 5·66 MPN/g) [21]. The samples tested in this study were further multi-processed with washing, cutting, packaging, storage and transportation at 4 °C before being subjected to examination. The data provided here are only for the final products, and therefore the compositional changes of STEC and microbiota in these foods during each process needs to be clarified in our future work. A global metabolomic approach will be valuable to gain comprehensive information on the composition and functional roles of background microbiota in food products.

In conclusion, we demonstrated the prevalence of STEC in various types of bovine offal products for the first time; the prevalence data (i.e. contamination rates with the isolate's virulence characteristics) herein may be useful for the risk assessment of those products as a potential source of human infection. Because a low dose of this pathogen can cause a relatively high attack rate, even survival without growth in bovine digestive-tract products may pose a public health risk if the food is insufficiently heat-treated or consumed raw. Other data such as 'annual consumption volumes/rates' for these foods, would further improve our understanding of the epidemiological importance. The characteristic growth kinetics of STEC O157 shown in this study also indicates the importance of being aware when to test these foods.

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## DECLARATION OF INTEREST

None.

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