Tiny Bubbles

Stephen W. Carmichael¹

Mayo Clinic

carmichael.stephen@mayo.edu

This is not an article about the song made famous by the late (great) Don Ho. This is about a breakthrough in the understanding of how micrometer-sized bubbles can be stabilized for long periods of time. This can influence the taste, smell, and consistency of consumer products including food and cosmetics.

In two-phase systems, which can include air (as bubbles) suspended within a liquid, the structures of the dispersed (bubbles) and continuous (liquid) phases play a critical role in determining the properties of the material. There is also the function of time in that the microstructure of the dispersed phase continuously evolves toward a state of lower energy by minimizing the surface area between the two phases (referred to as the interfacial area). In the long term, this time evolution diminishes the usefulness of two-phase systems. Emilie Dressaire, Rodney Bee, David Bell, Alex Lips, and Howard Stone have devised a way to stabilize a two-phase system for time periods of a year or longer.²

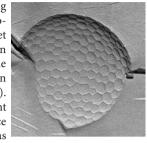
In gas-liquid two-phase systems, the air/liquid surface tension produces a pressure that drives the dissolution of gas into the liquid that leads to larger bubbles growing at the expense of smaller ones (a phenomenon known as Ostwald ripening). An important aspect of this is that it is the product of the surface tension and the curvature of the interface that produces the pressure difference between the bubble and the continuous phase. The time scale of this dissolution can be less than a second for micrometer-sized bubbles in pure water. The separation can be delayed by a few orders of magnitude by the addition of surfactants and other "tricks" that decrease the surface tension, but it is still difficult to keep small bubbles suspended in a liquid for more than a few months.

Dressaire et al. reported creating very stable gas dispersions obtained

using a standard multiphase mixing technique to trap air into surfactant shells within a viscous liquid. The surfactant solution was very viscous glucose syrup (approximattely 3 parts sugar to 1 part water) with sucrose stearate as the surfactant (2% by weight; mainly a mixture of mono- and diesters). The solution was aerated by vigorous mechanical mixing for 2 hours that "sheared" trapped bubbles within the viscous liquid to reduce the size of the bubbles.

The resulting foam was examined using several techniques and microscopes. These included freeze-fracture transmission electron

microscopy (TEM), cryogenic scanning electron microscopy (cyro-SEM), and cryo-TEM. Using these techniques, Dressaire et al. showed microbubbles (about 1 micron in diameter) and that every microbubble had a nearly hexagonal surface pattern on a nanometer scale (see cryo-SEM, right). They thought that using several different imaging methods was essential to convince themselves that what they were seeing was real rather than artifacts.



Dressaire et al. have demonstrated that it is possible to achieve stabilization of microbubbles in a viscous solution and that this system can be stabilized for more than a year. The regular surface patterning they found on the microbubbles is the thermodynamic signature of the formation of an elastic, condensed surfactant phase that also correlates with the extended stability of the system. The features of this system can be tuned on a nanometer scale by modifying the chemical composition of the interface. It will be interesting to see how this new system will be incorporated into consumer products in the future!

- 1. The author gratefully acknowledges Dr. Howard Stone for reviewing this article.
- Dressaire, E., R. Bee, D.C.Bell, A. Lips, and H.A. Stone, Interfacial polygonal nanopatterning of stable microbubbles, Science 320:1198-1201, 2008.

INDEX OF ARTICLES

Tiny Bubbles
Stephen W. Carmichael, Mayo Clinic
New Large Area Silicon Drift Detectors - Fast Analysis
without Compromise6
Clair Collins, Neil Rowlands, Peter Statham, and James Holland
Oxford Instruments, High Wycombe, Bucks, England
Microscopy Today New Publication Directions12
Ron Anderson and Charles Lyman,*Microscopy Today, Largo, FL and *Lehigh University Bethlehem, PA
Manufacturer Training of Electron Microscopy and
Analysis Techniques18
Neil Rowlands, Oxford Instruments, Concord, MA
Remote Microscopy for Education and Outreach22
S. Seraphin, S. Hernandez, G. Chandler, D. Bentley*, K. Dorame, M. Sellers, ** Univ. of Arizona, Tucson, AZ, * ** N. Arizona Univ., Flagstaff, AZ
The Electron Microscopy Database: an Online Resource
for Teaching and Learning Quantitative Transmission
Electron Microscopy26
Paul M. Voyles, Department of Materials Science and Engineer- ing, University of Wisconsin, Madison, Madison, WI
SEM Short Courses for Industry: the Lehigh Microscopy
School as an example28
Charles E. Lyman, Department of Materials Science and Engineering, Lehigh University, Bethlehem, PA
Direct Visualisation, Sizing and Counting of Virus and
3
Bob Carr, and Duncan Griffiths,* NanoSight Ltd., Salisbury, UK, *NanoSight USA, Costa Mesa, CA

Single-Molecule DNA Stretching Using Optical Tweezers4	2
Joost van Mameren, Anna Wozniak, and Sid Ragona,* JPK	
Instruments, Berlin, Germany, *Ragona Scientific, Pittsford, N	Y
Event Streamed Spectrum Imaging using Programmed	
Beam Acquisition in Biological Microprobe Analysis 4	4
P. Ingram, * S. D. Davilla, ** & A. LeFurgey*, *Duke Univ. and Veteran	ıs
Affairs Med. Ctr, Durham, NC, **4pi Analysis Inc., Durham, NC	
RGB-Splitting and Multi-Shot Techniques in Digital	
Photomicrography–Utilization of Astronomic RGB-Filters	
n True Color Imaging 4	
Jörg Piper, Clinic "Meduna," Bad Bertrich, Germany	
Preventing the Sale of Fraudulent Gemstones using Non-	
Destructive X-Ray Fluoresence Spectroscopy5	2
Mary S. Goldman, Dan L. Davis, Robert H. Clifford, Shimadzu Scientific Instruments Inc., Columbia, MD	ı

This image is a thin (30 micrometer) section of a lunar breccia recovered by Edwin Aldrin and Neil Armstrong in the Sea of Tranquility. The image was recorded in polarized light on a Nikon OptiPhot-Pol microscope and has a magnification of ~400x. Cover courtesy of Molecular Expressions, (microscopy.fsu.edu).

COMING EVENTS

2009

✓ NANOAFRICA 2009

February 1-4, 2009 Pretoria, South Africa www.nanoafrica.co.za

✓ The 4th International workshop on Piezoresponse Force Micros. February 2009, Aviero, Portugal

ftp.ua.pt/incoming/4th_PFM_workshop/4thWorkshop_Aveiro.pdf

✓ Principles and Practice of Light Microscopy: A Training Course March 1-7, 2009. Bangalore, India www.ncbs.res.in/events/microscopy.html

Workshop on FRET Microscopy March 3-7, 2009, Charlottsville, VA, www.kcci.virginia.edu/workshop/workshop2009/index.php

✓ PITTCON 2009

March 8-13, 2009, Chicago, IL www.pittcon.org

 ✓ Focus On Microscopy 2009
 April 5-8, 2009, Krakow, Poland www.FocusOnMicroscopy.org

 EELS and EFTEM Analysis Course April 6-9, 2009, Pleasanton, CA www.gatan.com

√ 2009 MRS Spring meeting

April 13-17, 2009, San Francisco, CA www.mrs.org
See especially Symposium JJ on Nanoscale Electromechanics and Piezoresponse, www.mrs.org/s_mrs/sec.

asp?CID=14465&DID=211517 Force Microscopy

American Soc. for Biochemistry and Molecular Biology

April 18-22, 2009, New Orleans, I.A.

April 18-22, 2009, New Orleans, LA www.asbmb.org

Microanalysis of Particles

April 200-23, 2009, Westmont, IL

www.microbeamanalysis.org/meetings/topical/Particles2009/index.htm

Lehigh Microscopy School (Multiple Courses) June 1-13, 2009, Bethlehem, PA www.lehigh.edu/microscopy/

✓ Frontiers in Polymer Science June 7-9, 2009, Mainz, Germany www.frontiersinpolymerscience.com

14th Short Course on 3D Microscopy of Living Cells June 13-25, 2009, Vancouver, BC, Canada www.3dcourse.ubc.ca/

Microscopy and Microanalysis 2009 July 26-30, 2009, Richmond, VA www.msa.microscopy.org

✓ Neuroscience 2009 October 17-21, 2009, Chicago, IL www.sfn.org

2010

✓ Microscopy and Microanalysis 2010 August 1-5, 2010, Portland, OR

2011

Microscopy and Microanalysis 2011
 August 7-11, 2011, Nashville, TN

2012

✓ Microscopy and Microanalysis 2012 July 29-August 2, Phoenix, AZ

Please check the "Calendar of Meetings and Courses" in the MSA journal "Microscopy and Microanalysis" for more details and a much larger listing of meetings and courses.

MICROSCOPY TODAY

The objective of this publication is to provide material of interest and value to working microscopists!

The publication is owned by the Microscopy Society of America (MSA) and is produced six times each year in odd months, alternating with MSA's peer-reviewed, scientific journal *Microscopy and Microanalysis*. We greatly appreciate article and material contributions from our readers—"users" as well as manufacturers/suppliers. The only criterion is that the subject matter be of interest to a reasonable number of working microscopists. *Microscopy Today* has authors from many disparate fields in both biological and materials sciences, each field with its own standards. Therefore *MT* does not have a rigid set of style instructions and encourages authors to use their own style, asking only that the writing be clear, informative, and accurate. Length: typical article length is 1,500 to 2,000 words plus images, longer articles will be considered. Short notes are encouraged for our Microscopy 101 section. See our "Instructions to Authors" document on our website.

MICROSCOPY TODAY

ISSN 1551-9295

Charles E. Lyman, Editor-in-Chief

charles.lyman@lehigh.edu

Ron Anderson, Technical Editor microscopytoday@tampabay.rr.com Phil Oshel, Technical Editor

oshel1pe@cmich.edu

Thomas E. Phillips, Contributing Editor PhillipsT@missouri.edu

Dale Anderson, Art Director microscopytoday@tampabay.rr.com Renée Stratmoen, Advertising Director

oshel1pe@cmich.edu
Regular Mail to:

Microscopy Today, P.O. Box 247, Largo, FL 33779

Telephones:

1-(727)507-7101 • Fax: (727)507-7102 • Cell: (727) 631-1022

e-Mail:

microscopytoday@tampabay.rr.com

Website:

http://www.microscopy-today.com

Colophon: Microscopy Today is created using components of Adobe Creative Suite CS4®

Total Circulation: 16,409

Disclaimer: By submitting a manuscript to *Microscopy Today*, the author warrants that the article is original (or that the author has the right to use any material copyrighted by others). The use of trade names, trademarks, *etc.*, does not imply that these names lack protection by relevant laws and regulations. *Microscopy Today*, the Microscopy Society of America, and any other societies stated, cannot be held responsible for opinions, errors, or for any consequences arising from the use of information contained in *Microscopy Today*. The appearance of advertising in *Microscopy Today* does not constitute an endorsement or approval by the Microscopy Society of America of the quality or value of the products advertised or any of the claims, data, conclusions, recommendations, procedures, results or any information found in the advertisements. While the contents of this magazine are believed to be accurate at press time, neither the Microscopy Society of America, the editors, nor the authors can accept legal responsibility for errors or omissions.

© Copyright, 2009, The Microscopy Society of America. All rights reserved.

See surface detail at the nanoscale like never before Magellan™ XHR SEM

Discover a world never seen before

- Subnanometer resolution from 30 kV down to 1 kV
- Surface details at low voltages you cannot see any other way
- Fast, accurate, precise analysis at higher voltages with high, stable current
- No compromises in sample size, shape, composition and preparation
- Simple and easy to use

This series of images from FEI's Magellan XHR SEM shows nanotubes imaged at 200 V, from 10 k to 600 k times magnification. Note the contrast and surface detail that would not be possible on other SEMs. Images courtesy of Prof. Raynald Gauvin and Camille Probst, McGill University.

