

The effect of condensed tannins in *Lotus corniculatus* on plasma metabolism of methionine, cystine and inorganic sulphate by sheep

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Fresh *Lotus corniculatus* containing 27 g extractable condensed tannin (CT)/kg dry matter (DM) and 8 g bound CT/kg DM was fed at hourly intervals to sheep held in metabolism cages to study the effects of CT on nutrient digestion and on metabolism of methionine, cystine and inorganic sulphate in plasma. Polyethylene glycol (PEG) was continuously infused into the rumen of half the sheep to remove the effects of CT. Principal measurements in the two groups were plasma irreversible loss (IRL) rate and interconversions of methionine, cystine and inorganic sulphate using ³⁵S labelling. CT in *Lotus corniculatus* had no effects on the apparent digestion of cellulose and minerals, slightly depressed DM, organic matter and hemicellulose digestion and markedly reduced the apparent digestion of N ($P < 0.01$). The concentration of NH₃ and molar proportions of *iso*-butyric acid, *iso*-valeric acid and *n*-valeric acid in rumen fluid were markedly increased by the PEG infusion ($P < 0.01$), whereas total volatile fatty acid concentration and molar proportions of acetic acid, propionic acid and *n*-butyric acid were not affected. PEG infusion temporarily increased rumen protozoa numbers. CT greatly increased the IRL of plasma cystine (13.1 v. 7.0 μmol/min; $P < 0.05$) and reduced IRL of plasma inorganic sulphate (36.8 v. 48.1 μmol/min; $P < 0.01$) but had no effect on methionine IRL. CT increased transulphuration of methionine to cystine (4.37 v. 1.24 μmol/min; $P < 0.05$), increased cystine entering the plasma from whole-body protein turnover plus absorption from the small intestine (9.34 v. 5.75 μmol/min; $P < 0.05$) and increased cystine flux to body synthetic reactions (11.89 v. 5.41 μmol/min; $P < 0.05$). CT had no effect on the proportion of methionine total flux transferred to sulphate (0.05 v. 0.06; $P < 0.05$), reduced the proportion of methionine flux transferred to body synthetic reactions (0.68 v. 0.86) and markedly reduced the proportion of cystine flux transferred to sulphate (0.09 v. 0.27; $P < 0.01$). It was concluded that CT in *Lotus corniculatus* reduced rumen protein degradation and markedly increased utilization of plasma cystine for body synthetic reactions.

Condensed tannins: Digestibility: Sulphur-containing amino acids: Inorganic sulphate: Sheep

Condensed tannins (CT) are plant secondary compounds which can react with protein and carbohydrate by hydrogen bonding in a pH-dependent manner (McLeod, 1974). Nutritional effects of CT on the ruminant animal are dependent in part on their concentration in the plant. High levels of CT (50–100 g extractable CT/kg dry matter (DM)) have been demonstrated to depress voluntary feed intake (VFI) and reduce apparent digestibility of DM, organic matter (OM) and fibre (Donnelly & Anthony, 1969, 1970; Reed *et al.* 1982; Barry & Duncan, 1984; Barry & Manley, 1984; Chiquette *et al.* 1988; Pritchard *et al.* 1988). In particular, rumen protein and fibre digestion are depressed. As the CT–protein complex is stable and insoluble at pH 3.5–7.0 but dissociates and releases protein at pH < 3.5 (Jones & Mangan, 1977), low levels of CT (20–40 g extractable CT/kg DM) are believed to be beneficial in ruminant diets by reducing protein degradation in the

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rumen and increasing amino acid absorption from the small intestine, without depressing rumen fibre digestion and VFI (Barry *et al.* 1986; Waghorn *et al.* 1987a; Barry, 1989; Waghorn, 1990).

When *Lotus corniculatus* was fed to sheep, Waghorn *et al.* (1987b) found that apparent absorption of essential amino acids (EAA; excluding methionine, cystine and tryptophan) from the small intestine was increased 62% by the CT (22 g extractable/kg DM). Apparent absorption of non-essential amino acids (NEAA) was reduced 10%. More recently, McNabb *et al.* (1993) observed that in sheep fed on *Lotus pedunculatus* the presence of CT (55 g extractable/kg DM) prevented a net loss of methionine and cystine across the rumen, and increased apparent absorption of methionine from the small intestine by 27%. CT did not increase apparent absorption of cystine but it did increase both plasma irreversible loss (IRL) of cystine and the amount of cystine incorporated into body synthetic reactions, due to transulphuration of methionine to cystine. In the same study CT had no effect upon the apparent digestibility of methionine in the small intestine (0.78), but reduced the apparent digestibility of cystine in the small intestine from 0.53 to 0.42, showing that at this concentration CT may be affecting the hydrolysis of proteins in the small intestine and reducing the apparent absorption of some amino acids.

S-containing amino acids are of particular importance for sheep because both cysteine and methionine are precursors of cystine, which is a major component of wool protein (Reis, 1979). The present experiment was designed primarily to determine the effects of CT in *Lotus corniculatus* on plasma IRL of S-containing amino acids, together with transulphuration of methionine to cystine and oxidation to sulphate. The experimental protocol also provided an opportunity to examine effects of CT on rumen volatile fatty acids (VFA) concentrations and rumen protozoal numbers when *Lotus corniculatus* was fed.

MATERIALS AND METHODS

The experiment involved fourteen castrated male Romney sheep aged 18 months which had been fitted previously with rumen cannulas (55 mm i.d.). They had been grazing out of doors and were brought inside (day 1) and held indoors in metabolism cages for 34 d. The sheep were introduced to *Lotus corniculatus* and allowed a 10-d adjustment period before sampling commenced.

Animals

The sheep weighed 48 (SE 1.5) kg at the beginning of the experiment. All were drenched with anthelmintic to control internal parasites (12 ml; Ivomec, Merck Sharp and Dohme (NZ) Ltd, Auckland, New Zealand) and treated for external parasites (10 ml; Wipeout, Coopers Animal Health, NZ Ltd, Upper Hutt, New Zealand) before the start of the experiment. The sheep had been held in metabolism cages previously. Feed intakes were recorded from day 4, and an intraruminal infusion of polyethylene glycol (PEG; molecular weight 3500) was given (50 g/d in 240 ml water) to six animals commencing on day 12 and continuing until day 34 (PEG group). The PEG binds with CT, preventing the CT from binding with protein (Jones & Mangan, 1977). The remaining eight animals did not receive an intraruminal infusion and acted as a control group.

Feed

The *Lotus corniculatus* was harvested daily at about 08.00 hours with a sickle-bar mower. It was vegetative throughout the trial and was fed hourly from overhead feeders. Feed was offered *ad lib.* for the first 7 d of the experiment and at about 90% of *ad lib.* thereafter. About 40% of the daily feed was placed on the feeders at 09.30 hours and the remainder was held at 4° until 17.00 hours and then placed on the feeders. Refusals were collected each

morning and daily DM intakes measured throughout the experimental period. Feed samples were collected at 2 d intervals and held separately at -20° for chemical analyses.

Rumen sampling protocols

Metal probes covered in a synthetic fibre (Estal-mono; Swiss Screens, Sydney, Australia; 88 μm pore size) were suspended in the rumen, enabling fluid to be collected by gentle suction (20 ml syringe). Rumen fluid was sampled between 13.00 and 15.00 hours commencing on day 11 of the experimental period (before PEG infusion) and on days 1, 2, 4, 6, 10, 16 and 22 after the PEG infusion started. The rumen fluid was used to determine rumen NH_3 and VFA concentrations and protozoal numbers.

The rumen fluid used for the determination of NH_3 concentration (20 ml) and VFA concentration (5 ml) was deproteinized immediately after sampling (5 ml 2.5 M- H_2SO_4 saturated with MgSO_4 for NH_3 samples, and 1 ml metaphosphoric acid-formic acid mixture for VFA samples). VFA protein precipitant comprised 375 g metaphosphoric acid and 500 ml formic acid, made up to 2 litres. The deproteinized NH_3 and VFA samples were then centrifuged at 1600 g and the supernatant fluid was stored at -20° . The rumen fluid used to determine protozoal numbers was preserved by addition of formal saline (9 g NaCl and 40 g HCHO/l) and stored at 4° .

Sulphur amino acid kinetics

Six control sheep and six PEG sheep had catheters installed in both jugular veins on day 15 of the experimental period in order to infuse [^{35}S]methionine (day 18), [^{35}S]cysteine (day 21) and $^{35}\text{SO}_4$ (day 24). Each isotope was infused continuously for 30 h. Blood was sampled from the opposite catheter before infusion (background) and after 24, 26, 28 and 30 h infusion to determine the specific radioactivity (SA) of isotope in plasma. The isotopes (Amersham, Australia Pty Ltd, Auckland, New Zealand) were prepared as follows: [^{35}S]methionine (203.5 MBq) was added to 3.3 litres sterile saline containing 0.25 mmol/l inert L-methionine (BDH, Poole, Dorset) as a carrier, [^{35}S]cysteine (199.8 MBq) was added to 3.21 sterile saline containing 0.24 mmol/l inert L-cysteine (BDH) as a carrier and [^{35}S]ammonium sulphate (388.5 MBq) was added to 3.31 sterile saline containing 0.31 mmol/l inert ammonium sulphate (BDH) as a carrier. The rates at which methionine, cysteine and inorganic sulphate were infused into each sheep were 0.4144, 0.4847 and 0.9250 MBq/h respectively.

During the methionine and cysteine infusions about 25 ml blood per sampling time was collected into syringes with Na-EDTA as an anticoagulant and placed on ice. The blood was centrifuged (3000 g, 15 min) to obtain plasma which was divided into two portions. One portion (10 ml) was treated with 15 μl 2-mercaptoethanol to reduce oxidation and centrifuged through a molecular filter (Centriprep-10 membrane filter tubes; molecular weight cut off 10000; Amicon, Beverly, MA, USA) at 1900 g to remove large proteins. The filtrate was frozen at -80° until analysed for methionine, cysteine and cystine. A second portion (2 ml) was deproteinized with 0.2 ml trichloroacetic acid (TCA) solution (500 g/l), allowed to stand at 0° for 15 min and was centrifuged at 3000 g for 20 min. The supernatant was retained at -80° for sulphate analysis. When $^{35}\text{SO}_4$ was infused, only 10 ml blood was collected and the plasma was deproteinized with TCA and held at -80° for analysis.

Digestibility

Following the isotope infusions, harnesses were put onto the sheep (day 26) enabling faeces collection bags to be attached for total faeces collection from day 27 to day 34. Faeces were collected, weighed and a sample (100 g/kg) was bulked over the 7 d collection period (at -20°) for analysis. Feed DM determinations were made in triplicate (95° for 24 h) and the

quantity of material refused was determined for each sheep. Samples of feeds and refusals were retained for analysis. Faeces DM was determined by drying at 95° for 48 h.

Analytical

Feed, feed refusal and faeces. All analyses of feeds, refusals and faeces were made with freeze-dried material. Carbohydrate fractions were determined by sequential detergent extraction (Van Soest, 1983) and total N by Kjeldahl digestion. Samples were heated at 550° for 16 h to determine ash content in the DM. CT were determined as extractable, protein-bound and fibre-bound fractions using the modified butanol-HCl procedure (Terrill *et al.* 1992*b*). Total S in feed DM was determined after oxidation by NaOBr (Tabatabai & Bremner, 1970) and feed samples for sulphate analysis were prepared by adding 1.0 g herbage to 50 ml 0.12 M-HCl, shaking overnight and making the extract up to 250 ml. The above prepared samples were analysed using an auto-analyser (Technicon Industrial Systems, Tarrytown, MD, USA) by the modified method of Johnson & Nishita (1952).

Rumen fluid. Rumen fluid samples were centrifuged (950 g, 15 min) and the supernatant was used for analysis. Rumen NH₃ was determined by distilling samples which had been made weakly alkaline with sodium tetraborate (pH 9.5) into boric acid (10 g/l) and titrating against 0.02 M-HCl using mixed bromocresol green-methyl red indicator (Kjeltec Auto 1030 Analyzer; Kjeltec, Höganäs, Sweden). Rumen VFA were determined by capillary gas chromatography (Carlo Erba GC-5380, Italy) using a 15 m × 0.53 mm i.d. FFAP (1.0 µm coat thickness column, Quadrex, Newhaven, CT, USA). The procedure used a split injection mode (approximately 2:1 split ratio) and a 5°/min temperature programme from 115 to 145°, with H₂ as carrier gas (pressure 0.5 kg/cm²), and a flame ionization detector. External standardization was used and the data were calculated using a Maxima 820 chromatography workstation (Waters Associates, Milford, MA, USA). Protozoa were counted in a 0.1 mm deep haemocytometer chamber, using light microscopy.

Plasma samples. All HPLC and radioactivity analyses for methionine, cysteine, cystine and inorganic sulphate commenced immediately after the animal work concluded. Measurement of SA from one control and one PEG sheep showed that SA of methionine, cystine and inorganic sulphate had reached plateau values after 24 h infusion. Therefore the plasma samples from 26, 28 and 30 h were pooled for each sheep. The concentrations and the radioactivities of methionine, cysteine, cystine and inorganic sulphate were then determined in the bulked samples.

The concentrations of methionine, cysteine and cystine were determined by HPLC (Waters Associates), using a reverse phase Pico.Tag column for free amino acids and using the Pico.Tag analytical method for physiological samples (Cohen *et al.* 1989). Deproteinized plasma (20 µl), derivatized with phenylisothiocyanate (PITC), was injected into the column. The amino acids were detected by the fluorescence of their PITC derivatives using a Waters (USA) 490E programmable multiwavelength detector. The data were calculated using a Maxima 820 chromatography workstation (Waters Associates).

The ³⁵S radioactivity in methionine, cysteine and cystine was determined in a 1 ml portion of deproteinized plasma, by HPLC with an ion-exchange column, using the method described by McNabb *et al.* (1993). Radioactivities were counted in a liquid scintillation counter (Wallac 1409, Pharmacia, Turku, Finland).

Inorganic sulphate in deproteinized plasma was separated from organic S with Dowex 1-X8 resin (Cl-form, 18-52 mesh size, 1.33 meq/ml binding capacity; BDH). Preparation of the resin and the separation procedure were the same as described by McNabb *et al.* (1993), except that 3 M-HCl instead of 1 M-HCl was used to elute inorganic sulphate off the resin. The radioactivity of inorganic sulphate was determined by adding 1 ml eluent to 10 ml high-efficiency phase combining system (PCS II) and 2.0 ml glacial acetic acid, and using a liquid scintillation counter (Wallac 1409). Inorganic sulphate concentration was

Table 1. *Chemical composition of the Lotus corniculatus (g/kg dry matter)*
(Mean values with their standard errors for twelve samples)

	Mean	SE
Organic matter	906	5.3
Total nitrogen	34.8	0.30
NDF	307	8.3
ADF	229	4.9
Hemicellulose	78	1.3
Cellulose	142	6.2
Lignin	80	8.1
Total sulphur	2.01	0.123
Sulphate sulphur	0.43	0.022
Extractable CT	27.1	1.69
Protein-bound CT	6.1	0.27
Fibre-bound CT	1.8	0.12
Total CT	35.0	1.78

NDF, neutral-detergent fibre; ADF, acid-detergent fibre; CT, condensed tannin.

determined by the automated method of Johnson & Nishita (1952), using an auto-analyser (Technicon Industrial Systems). All the standards used for auto-analysis were made up in 3 M-HCl which had been treated in an identical fashion to deproteinized plasma samples.

³⁵S-labelled methionine, cysteine and ammonium sulphate infusates were added to deproteinized plasma in order to determine the recovery of ³⁵S-labelled methionine, cysteine and sulphate in each analytical method. Radioactivity of background samples was also determined in an identical fashion.

Calculation of the data and statistical analysis

Comparison between control and PEG treatments was by analysis of variance. Means are presented with the standard error of the mean (SEM). DM and OM digestibilities were calculated on a PEG-free basis.

Both concentrations and radioactivity of cysteine and cystine were combined and expressed as cystine. The calculation of SA, IRL and transfer quotient (TQ) used methods described by McNabb *et al.* (1993). Background values were deducted in determining the SA for all metabolites. All plasma metabolite concentrations and SA values were corrected for recoveries determined for each analytical method. The results of S amino acid kinetics are presented as a three-pool, compartmentalized model as described by McNabb *et al.* (1993) and the amounts of S from inorganic sulphate transferred to methionine and cystine were assumed to be zero. Total flux was calculated as the summation of all outflows from each pool, and individual outflows were then expressed as a proportion of the total flux. Oxidation of methionine and cystine was calculated as the proportion of total flux transferred to inorganic sulphate.

RESULTS

The DM content of *Lotus corniculatus* was 157 (SE 9.6) g/kg, and the chemical composition of the DM showed little variation throughout the experiment (Table 1).

Apparent digestibility

During the digestibility period the DM intake of the fresh *Lotus corniculatus* of the control sheep (985 (SE 84.5) g DM/d) and the sheep receiving PEG (982 (SE 90.7) g DM/d) was not significantly different ($P > 0.05$). The apparent digestibilities of DM (0.766 v. 0.786;

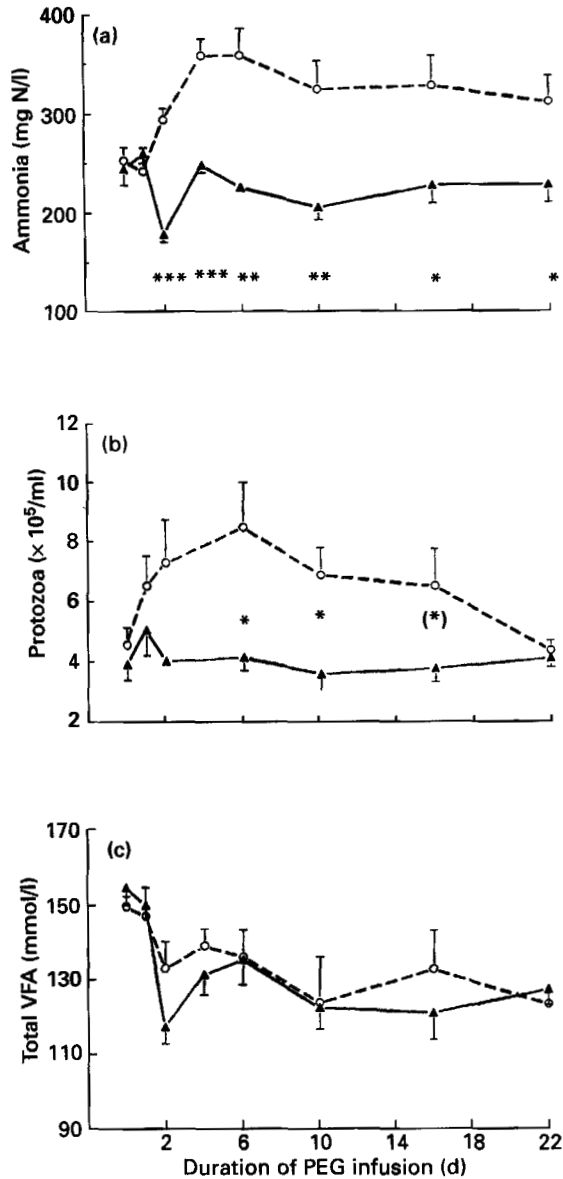


Fig. 1. The effect of intraruminal infusion of polyethylene glycol (PEG; molecular weight 3500) on (a) ammonia concentration (mg N/l), (b) protozoa numbers ($\times 10^5$ /ml) and (c) total volatile fatty acid (VFA) concentration (mmol/l) in the rumen fluid of sheep fed on fresh *Lotus corniculatus* at hourly intervals. Values are means for six sheep per group, with their standard errors represented by vertical bars. (\blacktriangle — \blacktriangle), Control sheep; (\circ — \circ), PEG-infused sheep. Mean values were significantly different between groups: (*) $P < 0.1$, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. For details of procedures, see pp. 924–926.

SEM 0.0079) and OM (0.774 v. 0.797; SEM 0.0084) were lower ($P = 0.084$ and 0.060 respectively) and total N (0.721 v. 0.795; SEM 0.0124) was significantly lower ($P < 0.01$) for control than for PEG sheep. Hemicellulose apparent digestibility (0.605 v. 0.666; SEM 0.0256) also tended to be lower in control sheep than in PEG sheep ($P = 0.118$). However, there were no differences between control and PEG sheep in the digestibility of cellulose (0.757 v. 0.763; SEM 0.0151) and minerals (0.692 v. 0.684; SEM 0.0097).

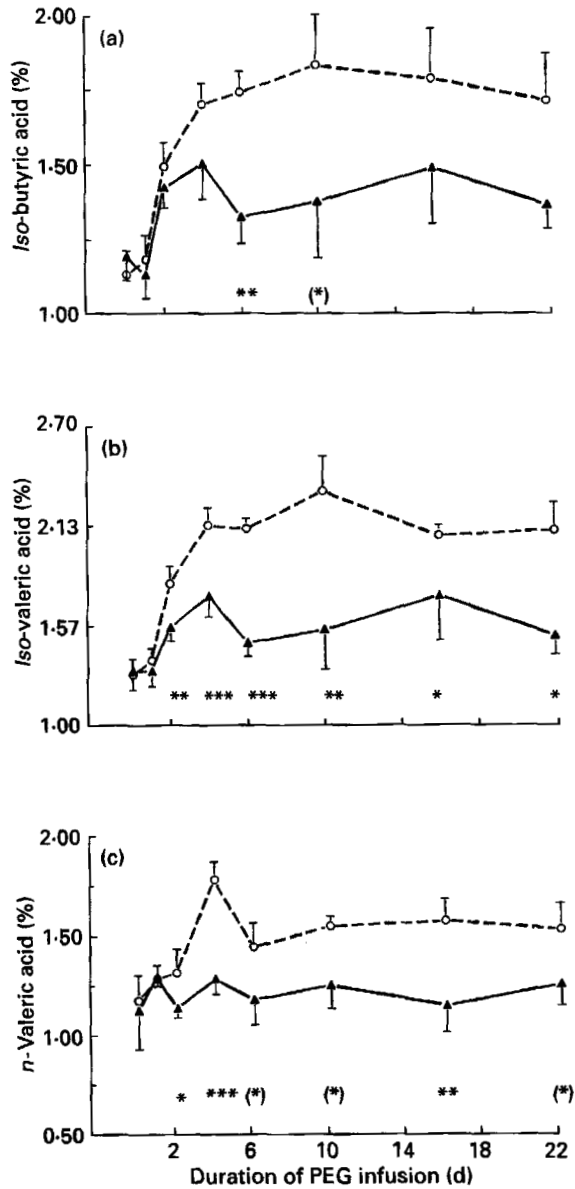


Fig. 2. The effect of intraruminal infusion of polyethylene glycol (PEG; molecular weight 3500) on the molar proportions (%) of (a) *iso*-butyric acid, (b) *iso*-valeric acid and (c) *n*-valeric acid in the rumen fluid of sheep fed on fresh *Lotus corniculatus* at hourly intervals. Values are means for six sheep per group, with their standard errors represented by vertical bars. (\blacktriangle - \blacktriangle), Control sheep, (\circ - \circ), PEG-infused sheep. Mean values were significantly different between groups: (*) $P < 0.1$, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. For details of procedures see pp. 924-926.

Rumen fluid

Ammonia concentration. There were no significant differences in the concentration of NH_3 in rumen fluid between control and PEG-infused sheep ($P > 0.05$; Fig. 1(a)) before commencement of PEG infusion or after 1 d intraruminal PEG infusion. However, from

Table 2. The concentration ($\mu\text{mol/l}$), irreversible loss rate ($\mu\text{mol/min}$), transfer quotients and the proportion of total flux flowing to various processes for cystine, methionine and inorganic sulphate in sheep fed on fresh *Lotus corniculatus*, with and without an intraruminal infusion of polyethylene glycol (PEG)†

(Mean values with their standard errors for six sheep in each group)

	Control sheep	PEG-infused sheep	SEM	Statistical significance
Concentration ($\mu\text{mol/l}$)				
Methionine	34.0	35.3	1.61	NS
Cystine	35.1	19.8	2.88	**
Sulphate	1012	1434	82.9	**
Transfer quotient				
Cystine from methionine	0.38	0.38	0.036	NS
Sulphate from methionine	0.01	0.01	0.001	NS
Sulphate from cystine	0.03	0.03	0.003	NS
Irreversible loss ($\mu\text{mol/min}$)				
Methionine	16.9	17.0	1.48	NS
Cystine	13.1	7.0	1.44	*
Sulphate	36.8	48.1	2.36	**
Proportion of flux transferred				
Methionine to cystine	0.27	0.08	0.039	**
Methionine to sulphate	0.05	0.06	0.004	NS
Methionine to P+M	0.68	0.86	0.040	*
Cystine to sulphate	0.09	0.27	0.043	**
Cystine to P+M	0.91	0.73	0.043	**

NS, not significant ($P > 0.05$); P+M, production and maintenance.

* $P < 0.05$, ** $P < 0.01$.

† For details of procedures, see pp. 924–927.

day 2 onwards, rumen NH_3 concentration was significantly lower in control sheep than in PEG-infused sheep. Rumen NH_3 concentrations after 4, 6, 10, 16 and 22 d PEG infusion were also significantly higher ($P < 0.01$) than those before PEG infusion commencing, whereas NH_3 concentration remained unchanged for control sheep during the experimental period ($P > 0.05$).

Rumen protozoa. Rumen protozoa numbers were not significantly different between control and PEG sheep before and on day 1 of the PEG infusion ($P > 0.05$), but protozoal numbers increased in PEG sheep relative to controls after 6 and 10 d ($P < 0.05$) and 16 d ($P = 0.064$) of PEG infusion (Fig. 1(b)). By day 22 the protozoal numbers had declined in PEG sheep and were not significantly different from those of control sheep ($P > 0.05$).

Rumen VFA. Total VFA concentrations (mmol/l rumen fluid) were similar for control (131.6 (SE 2.54) mmol/l) and PEG sheep (135.9 (SE 2.81) mmol/l) and no significant differences were detected at any sampling time (Fig. 1(c)); however, PEG infusion markedly changed molar proportions of some individual VFA (Fig. 2).

The molar proportions of acetic acid, propionic acid and *n*-butyric acid were not significantly different between control and PEG-infused sheep. Mean molar proportions of acetic acid, propionic acid and *n*-butyric acid over the entire experimental periods were 0.642 (SE 0.003), 0.220 (SE 0.004) and 0.098 (SE 0.003) for control sheep and 0.634 (SE 0.003), 0.214 (SE 0.004) and 0.104 (SE 0.002) for PEG-infused sheep. However, intraruminal infusion of PEG markedly increased the molar proportions of *iso*-butyric acid, *iso*-valeric

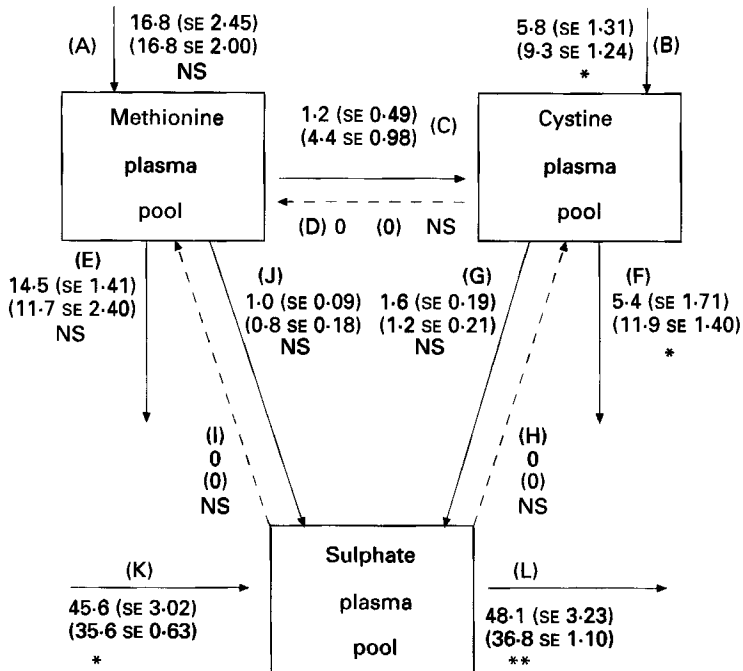


Fig. 3. A general three-pool, compartmentalized model for sulphur amino acid transactions in the posthepatic plasma of sheep fed on fresh *Lotus corniculatus*, with and without an intraruminal infusion of polyethylene glycol (PEG). Mean values with their standard errors are shown for six PEG-infused sheep, with the corresponding values for six control sheep given in parentheses. Mean values for PEG-infused sheep were significantly different from those for control sheep: * $P < 0.05$, ** $P < 0.01$. NS, not significant ($P > 0.05$). \downarrow , Rates of flow ($\mu\text{mol}/\text{min}$) in the model, which are often composite of several pathways of transfer. The pathways are: (A) methionine entering the plasma pool from whole-body protein turnover and absorption from the small intestine; (B) cystine entering the plasma pool from whole-body protein turnover and absorption from the small intestine; (C) transulphuration of methionine to cystine; (D) conversion of cystine to methionine, which does not occur in mammalian tissue; (E) methionine leaving the plasma and being utilized for productivity processes and maintenance; (F) cystine leaving the plasma pool and being utilized for productivity processes and maintenance; (G) cystine oxidized to sulphate (and carbon dioxide); (H) plasma sulphate re-assimilated as cystine. This cannot occur directly in mammalian tissue, but sulphate re-entering the rumen via saliva may be absorbed as cystine from microbial protein; (I) plasma sulphate re-assimilated as methionine. This cannot occur directly in mammalian tissue, but sulphate re-entering the rumen via saliva may be absorbed as methionine from microbial protein; (J) methionine oxidized to sulphate (and carbon dioxide); (K) sulphate entering the plasma, chiefly from oxidation of sulphide absorbed from the rumen, but also sulphate and oxidation of sulphide absorbed from the intestine; (L) sulphate leaving the plasma chiefly in urine, but also recycled directly to the intestines and rumen via saliva.

acid and *n*-valeric acid after 2 d PEG infusion until the conclusion of the experiments ($P < 0.05$; Fig. 2).

Sulphur amino acid kinetics

The feed intakes of the control and the PEG sheep were 989 (SE 71.4) and 983 (SE 87.6) g DM/d respectively and were not significantly different ($P > 0.05$) during the period when S amino acid kinetics were measured.

Methionine. The recovery of both methionine and [^{35}S]methionine infusate added to the deproteinized plasma by HPLC was 0.97 (SE 0.02). The plasma methionine concentration and IRL rates were not significantly different between control and PEG sheep (Table 2; Fig. 3). However, the proportion of methionine total flux transferred to production and maintenance function in control sheep (0.68) was lower than that in PEG sheep (0.86,

$P < 0.05$, Table 2), whilst the proportion of methionine flux transferred to cystine was much higher for control than for PEG sheep ($P < 0.01$). The proportion of methionine transferred to sulphate (i.e. oxidation) was very low (0.05 v. 0.06 ; $P > 0.05$) in control and PEG sheep respectively.

Cystine. The recovery of both cystine and [35 S]cysteine infusate, added to the deproteinized plasma, by HPLC was 0.96 (SE 0.03). Both plasma cystine concentration ($P < 0.01$) and IRL rate ($P < 0.05$) were higher in control sheep than in PEG sheep (Table 2). Cystine flux ($\mu\text{mol}/\text{min}$) into the plasma pool from both whole-body protein turnover plus absorption from small intestine and from the transulphuration of methionine were significantly higher in control sheep than in PEG sheep ($P < 0.05$; Fig. 3). The control sheep had a higher flux of cystine ($\mu\text{mol}/\text{min}$) to production and maintenance than the PEG sheep ($P < 0.05$) and a higher proportion of the cystine total flux transferred to production and maintenance than for PEG sheep ($P < 0.01$). However, the proportion of total cystine flux transferred to sulphate (i.e. oxidation) was less for control than for PEG sheep ($P < 0.01$).

Inorganic sulphate. The recovery from the separation procedure of both K_2SO_4 and [35 S]sulphate infusate added to the deproteinized plasma was 0.67 (SE 0.03). Control sheep had a lower plasma inorganic sulphate concentration and IRL rate than PEG sheep ($P < 0.01$; Table 2). This translated into less inorganic sulphate both entering the plasma pool from the digestive system and leaving via urinary excretion or recycling to the gut in control compared with PEG sheep (Fig. 3).

DISCUSSION

The most significant result of this study was the action of CT in increasing the amount of cystine leaving the plasma cystine pool to be used for body synthetic reactions (11.9 v. 5.4 $\mu\text{mol}/\text{min}$). This was due to increased entry flux (Fig. 3, B; 9.3 v. 5.8 $\mu\text{mol}/\text{min}$) and increased transulphuration of methionine to cystine (Fig. 3, C; 4.4 v. 1.2 $\mu\text{mol}/\text{min}$). McNabb *et al.* (1993) reported similar findings for the CT in *Lotus pedunculatus*, but cystine IRL rate determined in this experiment for control *Lotus corniculatus* (13.1 $\mu\text{mol}/\text{min}$) was lower than that for control *Lotus pedunculatus* (39.8 $\mu\text{mol}/\text{min}$). Lee *et al.* (1992) also found cysteine IRL in whole blood to be lower for sheep fed on *Lotus corniculatus* than *Lotus pedunculatus* (11 v. 24 $\mu\text{mol}/\text{min}$). This difference may be due to differences between the two *Lotus* species in CT reacting with plant protein and the different seasons when the two experiments were conducted.

Reactivity of CT with protein is probably a function of plant CT concentration, molecular weight and structure. Extractable and total (extractable + bound) CT were 27 and 35 g/kg DM for *Lotus corniculatus* (cv. Goldie) in the present study and 55 and 75 g/kg DM for *Lotus pedunculatus* (cv. Maku; Terrill *et al.* 1992*b*; Douglas *et al.* 1993; McNabb *et al.* 1993). In addition to the effects of increased concentration, reactivity of CT with protein increases with increasing degree of CT polymerization, and hence molecular weight (Horigome *et al.* 1988), and with increasing ratio of the CT constituent prodelphinidin (PD) to procyanidin (PC; Jones *et al.* 1976). Hence the higher PD:PC ratio and molecular weight of *Lotus pedunculatus* CT ($70:30$; 2900) than for *Lotus corniculatus* CT ($27:73$; 2000; Foo *et al.* 1982), together with the higher CT concentration, suggests that reactivity is greater for the CT in *Lotus pedunculatus* than in *Lotus corniculatus*.

Wool growth rate has been shown to be seasonal in Romney sheep in New Zealand, with highest rates in spring and summer and lowest rates in winter (Bigham *et al.* 1978). The experiment undertaken by McNabb *et al.* (1993) was carried out in spring, whereas the present study was conducted in late autumn, when wool-growth rate would be declining.

Because of its high S content, which is mainly in cystine form (Reis, 1965*a, b*; Hogan, 1975), rapid wool growth will create a high demand for cystine. Therefore, compared with the results of McNabb *et al.* (1993), the lower flow of cystine to body synthetic reactions in the present study may also be due to lower potential wool-growth rates and hence lower cystine requirement. The increased cystine flux to body synthetic reactions due to CT could result in increased wool growth.

Results from a grazing experiment using the same sward and twice daily oral administration of PEG showed that action of CT in *Lotus corniculatus* increased wool growth by 12% ($P < 0.05$; Y. Wang, G. B. Douglas, G. C. Waghorn, T. N. Barry, A. G. Foot and R. W. Purchas, unpublished results). Increased wool growth due to CT has also been reported in sheep grazing sulla (*Hedysarum coronarium*; Terrill *et al.* 1992*a*).

The best indication of methionine and cystine oxidation in plasma is considered to be the proportion of total flux transferred to inorganic sulphate. CT markedly reduced the oxidation for cystine but not methionine, which was already low. The low cystine oxidation rate (0.09) and high proportion used for production and maintenance function (0.91) in sheep fed on *Lotus corniculatus* with CT operating suggests that cystine availability may limit wool growth in sheep selected for high wool production.

The effects of CT are normally assessed through administration of PEG to bind and inactivate CT, and a comparison is made between control and PEG sheep (Barry *et al.* 1986; Waghorn *et al.* 1987*a, b*; Barry, 1989; Terrill *et al.* 1992*a*; McNabb *et al.* 1993). The effects of PEG are assumed to be specific for CT and to affect only protein digestion, without affecting digestion of other nutrients, but this has not been fully investigated. The present experiments showed that PEG supplementation increased protein breakdown in the rumen, as indicated by the increased NH_3 concentration. Minor VFA are produced from deamination of specific amino acids, with *iso*-butyric acid from valine, *iso*-valeric acid from leucine and *n*-valeric acid from arginine, ornithine, proline, σ -aminovaleric acid or lysine (El-Shazly, 1952; Van Soest, 1983), of which valine, leucine, arginine and lysine are EAA. Hence the reduced *iso*-butyric acid, *iso*-valeric acid and *n*-valeric acid in control compared with PEG sheep also suggests that CT reduced deamination of these EAA. However, the similarities of total VFA concentration and major VFA resulting from carbohydrate digestion (acetic acid, propionic acid and *n*-butyric acid) in control and PEG-infused sheep suggest that 35 g total CT/kg in the forage DM does not affect carbohydrate digestion in the rumen.

The present results for rumen protozoa populations are opposite to the results of Terrill *et al.* (1992*a*), who found CT in sulla (29 g extractable CT and 47 g total CT/kg DM) increased rumen protozoa counts. They explained the effect of CT in increasing protozoa numbers by the requirement of protozoa for insoluble protein and soluble carbohydrates. The rapid increase in protozoal numbers following PEG administration in the present experiment suggests inhibition of protozoa by CT. However, the response was transitory and a component may have been due to effects of PEG *per se*. A long-term study is needed to evaluate the effect of CT in *Lotus corniculatus* on rumen protozoal populations.

Tannins have been reported to inhibit digestive enzyme activities (Horigome *et al.* 1988; Ahmed *et al.* 1991; Longstaff & McNabb, 1991*a, b*; Yuste *et al.* 1992) and to increase endogenous N excretion in single-stomached animals (Price & Butler, 1980; Cousins *et al.* 1981). The rapidly increasing pH along the small intestine may enable CT-protein complexes to be re-formed to protect protein against being hydrolysed by endogenous enzymes, or the dissociated CT may bind to the gut epithelium and affect amino acid absorption. All these could reduce the apparent digestibility of N in the post-ruminal digestive tract and may explain the effects of CT in reducing apparent N digestibility in the present study. Research in this area should be conducted to quantify the effects of low

dietary CT levels on post-ruminal protein digestion, and especially on endogenous N excretion.

The results of the present study show that the lower CT concentration in *Lotus corniculatus* compared with *Lotus pedunculatus* (McNabb *et al.* 1993) still effectively increased the amount of cystine flowing to body synthetic reactions, but was probably close to the CT range for depressing rumen fibre digestion. Hence, *Lotus* species with lower contents of CT should be evaluated in order to define the optimum concentration of CT in this species for increasing the absorption of EAA, increasing the flow of S-containing amino acids to body synthetic reactions and increasing the efficiency of livestock production.

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