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Relationships between multiple welfare indicators measured in individual chickens across different time periods and environments

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Abstract

The assessment of animal welfare requires the collection of multiple indicators of welfare but quantification of their associations in different contexts is lacking. Previous studies have examined correlations between a few indicators, but not relationships between many different indicators, or between indicators taken from the same individuals in more than one environment. We housed 60 hens for six sequential 35-day phases in different pen environments. During each phase, a series of behavioural and physiological measures was taken for every bird: body and plumage condition, surface body temperature, behaviours observed in the home pens and during test periods, tonic immobility, physiological blood profiles, and faecal sample composition. Most variation in nearly all measures was not explained by either individual bird or grouping effects but varied across phases within the birds. Acknowledging this, we examined correlations between all parameters at the phase within-bird level, selecting a conservative P-value. A consistent set of correlations showed that a slow approach response and alert behaviour in the novel object test was associated with higher bodyweight, lower body temperature and lower acute phase protein, heterophil:lymphocyte ratio and blood glucose level. A cluster analysis confirmed these correlations. Other important parameters known to be linked to the hens' environmental preference (eg comfort behaviour) were independent of the set described above. We conclude that statistical techniques can reveal patterns of independence and redundancy in the collection of behavioural and physiological measures of welfare.

Keywords: animal welfare, cluster analysis, laying hen, multi-level modelling, stress, welfare indicator

Introduction

Animal welfare science is increasingly seen as a multidisciplinary exercise. No one marker can indicate good or poor welfare (Mason & Mendl 1993) and welfare assessments are more likely to incorporate multiple markers than in the past. For example, information on physiology was obtained for only 2% of 300 laying hen flocks included in the Laywel database in 2005 (http://www.laywel.eu), whereas recent experimental studies have generally taken multiple measures of welfare, including organ weights, white blood cell ratios, plasma, faecal or egg corticosterone levels, tests of immune function and observations of behaviour (Nicol et al 2006; Barnett et al 2009; Singh et al 2009; Tactacan et al 2009; Thogerson et al 2009). The collection of multiple measures for monitoring hen welfare on farms has also become accepted practice (eg Welfare Quality, www.welfarequality.net).

If different aspects of welfare could be separated into independent, non-overlapping components (eg injury, stress response, resting comfort), and each component unambiguously measured, then just one measure would be needed to represent each component. The relative importance of the components could be weighted, and the measures combined to draw overall conclusions. In reality, though, welfare is not a combination of separate non-overlapping components. Different aspects are inter-related in complex ways (eg injury will affect resting comfort), and the underlying biological systems are hugely complex. As a precaution, scientists take many measures of each aspect of welfare (eg Rodenburg *et al* 2008).

This is a pragmatic approach, but one that risks potentially expensive and time-consuming redundancy in data collection (Richard et al 2007). In addition, combining non-independent measures to draw overall conclusions is difficult, especially when they do not co-vary in a consistent way. For example, Nicol et al (2006) found that hens housed in single-tier aviaries at low stocking densities had higher mortality and worse plumage condition than hens housed at higher densities, but lower percent liver weights, indicating relatively lower stress levels (eg Thaxton & Puvadolpirod 2000). It is difficult to interpret such findings without knowing whether the lack of co-variance occurs at a flock or an individual level. At a flock level, severe feather pecking could result in mortality and feather damage to a proportion of the flock, but in relatively low stress conditions for the surviving perpetrators. However, if individual

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birds had both poor plumage condition and low liver weights this would be more difficult to explain.

Decisions about which measures to take would be helped by prior statistical information on the relationships between different welfare indicators in different contexts. Currently, very little such information is available for hens. Many studies report informally that a particular treatment, situation or genetic breed difference influences two or more indicators in the same way (eg Cheng & Jefferson 2008; Campo & Prieto 2009; Kjaer 2009; Sherwin *et al* 2010) and, occasionally, the relative time-courses of different welfare indicators in response to a given situation are examined (eg Dawkins *et al* 2004). However, in these studies, as in most others, different measures were taken on different birds, or information from groups of birds was pooled before analysis.

Individual-level explanations require data for both variables to be obtained from the same birds. Studies that have done so have focused mostly on traits correlated with feather pecking (Vestergaard *et al* 1993; Cloutier & Newberry 2002; Albentosa *et al* 2003) and fear (Ghareeb *et al* 2008) and rarely examined a wide range of indicators (though see Webster & Hurnik 1991). Recent work showing that hens fall into different 'types' based on their different patterns of response to behavioural tests (Ghareeb *et al* 2008) or environmental conditions (Nicol *et al* 2009), provides a further reason to ensure that relationships between welfare indicators are examined at a within-bird level. Decisions to streamline data collection should be taken only if the measures are strongly correlated for individual birds.

In a previous paper, we investigated the relationship between welfare indicators and environmental preference in laying hens (Nicol et al 2009). The aim of this further analysis was to use our extensive dataset to investigate relationships between the many measures recorded on each bird during the six housing phases of the experiment, ie at a phase within-bird level. We first examined the amount of variation in each indicator that occurred at the various levels in the dataset (phases nested within individual birds nested within groups). Indicators could be primarily socially shared traits, fixed individual traits, or flexible responses that moderate with different environments. The relative variances estimate the likelihood of these three scenarios for each indicator. This is important information to be used in deciding whether future studies need to collect data at an individual or flock level. We then examined correlations between pairs of selected indicators. Finally, we examined how the different indicators were related more globally, using cluster analyses.

Materials and methods

Subjects and housing

Sixty, medium Hyline laying hens were obtained at 18 weeks of age and housed in 24 pens $(0.96 \times 1.2 \times 2.0 \text{ m};$ width \times depth \times height) either in groups (12 groups of 4 birds) or individually (12 birds) with *ad libitum* feed and

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water. Individual birds could see other birds only when they were feeding at the externally mounted feed trough. Room temperature was held at between 18 and 22°C, and the birds were on a schedule of 12 h light:12 h dark. The pens could be converted into three different environments by altering the floor type and resources available: Wire floor (W), Shavings floor (Sh), and Peat, Perch and Nest-box (PPN), with additional peat and wood shavings on top of a solid wooden base floor, two perches of diameter 0.035 m fitted across the width of the pen at heights of 0.25 and 0.45 m from floor level and a nest-box. The hens lived in each environment for a 35-day period, before moving to a new environment established in a pen on the opposite side of a corridor. In total, each of the three environments was experienced twice in six sequential phases counterbalanced for order between groups. All work was conducted under UK Home Office licence.

Welfare indicator sampling

During each 35-day housing phase, physical examination of the birds was conducted on days 24 and 32. Faecal samples were collected in the afternoons; one pooled sample was collected over days 29 to 31 from each bird. Blood samples were taken in the mornings from half the birds on day 29 and the other half on day 30. Direct behavioural observations were taken in the mornings on day 16 and 31, tonic immobility (TI) tests were given on the afternoons of days 16 and 30, novel object tests and resource competition tests were conducted throughout day 23. Video recordings were made on days 5, 12, 19, 26 and 33, each for 15 min starting at 0900h and 15 min starting at 1615h. Further details of the techniques used are given in Nicol *et al* (2009).

Physical examination of birds (days 24 and 32 of each phase)

Each hen was weighed, then gently held by an assistant whilst integument and plumage condition was assessed. The condition of the skin and plumage of the head, neck, breast, back, tail, wings and vent were scored on ordinal scales 0-5 (no damage to extensive or severe damage). Ordinal scales were also used to record comb elevation 0-2 (upright, partially floppy, completely floppy), comb colour 0-2 (red, medium, pale), comb size 0-3 (0 < 3.5 cm²; 1 = 3.5 to 5.5 cm², 2 = 5.5-7.5 cm²; 3 > 7.5 cm²), foot cleanliness 0–5 (clean to very dirty). Claw length was measured in mm. The temperature of the head, comb, eye and foot was recorded at a distance of 1 m from the bird, within 1 min of removal from the home pen to a holding pen at the same ambient temperature, using a thermal camera (ThermaCAM, FLIR systems, http://www.flir.com/Thermography/eurasia/en/). Foot lesions, crouched posture, lack of alertness, dull eyes and plumage damage (except damage to the breast region) occurred rarely or never and were not analysed further. Interobserver reliability was checked at intervals throughout the experiment. Differences between the two time points were minimal for the physical indicators variables, and for the statistical analyses reported here, average values from the two measurements collected in each phase were used.

Direct behavioural observations (days 16 and 31 of each phase) An observation hatch (27.5×50 cm; width × height) at the front of each pen was opened to allow the two observers to see the birds. After 1 min, each bird was observed continuously for 5 min using the ethogram described in Nicol *et al* (2009), and the frequency and duration of all behaviour patterns recorded during morning observation sessions. Pens and hens within pens were observed in a pre-determined random order.

Video behavioural observations (days 5, 19 and 33 of each phase)

Hens were filmed with a camera mounted centrally at the top of each pen. Each hen was observed using focal animal sampling for a continuous 15-min period, taken between 1615 and 1630h. Behavioural analysis of the footage was conducted for each individual bird using Observer Video-Pro 5.0 software (Noldus Information Technology, Wageningen, The Netherlands) by the observers who had taken direct observations, and by two additional observers who were trained in the same way. We collected a total of 270 min behavioural data for 30 hens and, due to a technical failure on day 33 of phase 1, 255 min behavioural data for the other 30 hens. For statistical analysis, we therefore averaged data from the available observations obtained in each phase, and report these as average duration per 15-min period.

Tonic immobility (days 16 and 30 of each phase)

Tonic immobility is usually measured after birds have been exposed to a potentially fear-inducing stimulus. Since we wanted to assess basal fear levels in birds that were well habituated to handling, we had to employ a longer restraint time of 30 s compared with the usual 15 s. Each bird was restrained gently on its side (a procedure we have used before [Cashman *et al* 1989]) with light pressure applied to the body and neck region. The operator's hands were then removed. Three attempts were made to induce TI for every bird. The number of attempts that resulted in a state of TI of more than 15 s was recorded. Birds were allowed to stay in TI for a maximum of 300 s on any occasion, after which they were gently righted. TI durations were markedly skewed with mostly very short durations, so only the number of inductions was used as a measure for further analysis.

Blood sampling and analysis (am on days 29-30 of each phase)

A trained operator removed no more than 4 ml of blood from the wing vein, within 1 min of handling. Heterophil and lymphocyte counts were taken immediately to assess H:L ratio. Samples (0.5 ml) of whole blood were assayed immediately for glucose and lactate (mmol L⁻¹) using the inhouse biochemistry laboratory. The remaining 3.5 ml sample was stored on ice overnight, then centrifuged at 4,500 rpm for 10 min to separate serum. A portion of the serum was assayed immediately for osmolality (mmol kg⁻¹) and creatine kinase (µg L⁻¹), whilst the remainder was frozen at -20°C until the end of the experiment. The concentration of the acute phase protein al-acid glycoprotein, α 1AG, in serum was then determined using immunodiffusion kits (Cardiotech Services Inc, KY, USA). Corticosterone concentrations (ng ml⁻¹) were assessed by Cambridge Specialist Laboratories using RIA analysis.

Faecal sampling and analysis (pm on days 29–31 of each phase)

Each hen was placed in a wire cage with a plastic sheet floor, with at least one other hen visible in an adjacent cage, and left for 90 min to produce a faecal deposit. Faecal samples were placed in 50 ml plastic tubes and frozen at -20° C. The dry matter, ash, crude protein, fat and fibre content of each sample were determined using in-house laboratory analysis, using standard proximate analysis techniques. Dry matter digestibility and apparent fat and protein digestibility of the feed was estimated during week 37, using titanium dioxide as an inert marker.

Novel object tests (day 23 of each phase)

Novel object (NO) tests were conducted by opening the observation hatch and allowing birds to settle for 1 min. A novel object was then introduced to the pen and 30 scans of the activity of each bird taken at 10 s intervals. These scans were aggregated to give a total count for each behaviour pattern observed for each bird over the 300 s after the object was introduced. A different novel object was used for each phase of the experiment: striped rubber ball, green apple, inflated balloon, white plastic 200 ml bottle, aluminium can, transparent plastic 2 L bottle.

Mealworm resource competition tests (day 23 of each phase)

Mealworm resource competition (MW) tests were conducted by opening the observation hatch and allowing birds to settle for 1 min. A dish of mealworms was then introduced to the pen and 10 scans of the activity of each bird taken every 10 s. These scans were aggregated to give a total count for each behaviour pattern observed for each bird over the 100-s test.

Selection of behavioural indicators for further analysis

Our aim was to seek correlations and relationships amongst a large set of welfare indicators. However, our initial categorisation of behaviour into 31 separate activities recorded directly (DHP) and by video (VHP) in the home pen, 34 activities recorded in the novel object test and 35 activities recorded in the resource competition test resulted in an over-dominance of behavioural parameters (131 behavioural parameters in all). In our previous work examining associations between welfare indicators and bird preferences, we excluded rare activities but here, to examine relationships between the welfare indicators themselves, we wished to include all activities. We therefore first grouped them into 12 behavioural categories, based on their known functional relationships (Table 1). The duration of time spent in any of the activities within a category was summed to give an overall time spent in each behavioural category.

There were too few occurrences of behaviour in the 15 starred categories in Table 1 for further analysis, leaving us with 21 behavioural duration categories. Two other parameters, latency to approach mealworms and latency to approach the novel object were also included in the subsequent statistical analyses. A secondary aim was to compare the video observations with the direct observations, both for home pen behaviour and for the NO and MW tests. Neither collection method is *a priori* better than the other, but if information obtained using these two methods is comparable then the more convenient method could be used in future.

Table I Grouping of separate activities into 12 behavioural categories, and the number of observations of each category in the four sets of observations taken.

		Direct observation (2 × 300 s per phase)	Video observation (3 × 900 s per phase)	Mealworm test (100 s per phase)	Novel object test (300 s per phase)
LOCO	Fly, walk, run	340	359	200	273
COMFORT	Flap wings, stretch wings, dustbathe, feather raise, preen, scratch self, tail wag, shake head	279	354	4*	66
FORAGE	Feed, beak wipe, ground scratch, litter peck, furniture peck, egg peck, approach mealworm dish, eat worms, peck at dish	357	360	351	267
DRINK	Drink	104	349	2*	53
M-PECK-REC	Receive mild feather peck	59	142	I *	 *
M-PECK-GIVE	Give mild feather peck, give beak peck	76	146	2*	2*
S-PECK-REC	Receive severe peck	4*	38	0*	0*
S-PECK-GIVE	Give severe peck	7*	33	0*	0*
INACTIVE	Stand inactive, sit inactive	6*	21	0*	0*
ALERT	Stand alert, sit alert	359	360	280	355
INT-NO	Interaction with novel object, peck at novel object	n/a	n/a	n/a	189
NEST	Nest	0	0	0	0

Behavioural Activities included Number of non-zero occurrences of behaviour out of a possible 360 (60 hens × 6 phases)

Statistical analysis

All indicator variables were either continuous or ordered categorical. For estimating the relative variabilities of group, hen and phase within-hen we fitted 3-level normal response models in MLwiN version 2.16 (Rasbash et al 2009). For simplicity, we also treated the variables as continuous when calculating Pearson's correlations and cluster analyses. However, we also calculated non-parametric Spearman correlations to check whether the ordinal scaling of many of the variables affected the relationships found. For the cluster analyses, all variables were standardised prior to clustering, and the clustering was performed using the 'dist' and the 'hclust' commands in the R software package (R Development team 2009). The 'dist' function forms a matrix of distances between the standardised variables and this distance matrix is then the input used in the 'hclust' function which performs a (hierarchical) cluster analysis based on the given distance matrix. We used the default (Euclidean) distance metric in 'dist' and the average method of clustering in the 'hclust'

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function. For more details, see the R documentation (R Development team 2009).

Results

Indicator variability

It was striking that for most indicators the largest source of variability occurred at the phase within-bird level, demonstrating sensitivity of these indicators to the changing environment and ages of the birds (Table 2). Despite this overarching effect, at least 20% of the variation in indicators, such as bodyweight and condition score, comb characteristics, claw length, posture, dry matter digestibility, creatine kinase levels, responses in the MW test and video observations of alert, foraging, drinking and mild pecking behaviours was ascribed at individual-bird level, demonstrating some stability in these individual bird traits. For birds housed in groups, group-level effects explained some 5 to 16% of the variation in indicators, such as comb size, foot characteristics, condition score, and of many behaviours.

TI Tonic immobility 360 0.06 (± 0.01) 0.00 3.56 96.44 BW Bodyweight 360 1.74 (± 0.02) 0.00 64.20 35.80 FTHD-BR Feather damage to breast 360 1.17 (± 0.08) 0.00 11.25 88.75 COMB-SZ Comb elevation 360 2.28 (± 0.10) 8.99 68.13 22.88 COMB-COL Comb colour 360 0.77 (± 0.07) 9.06 0.00 90.94 CLAW-L Claw length 360 0.75 (± 0.07) 9.06 0.00 90.94 CLAW-L Claw length 360 0.77 (± 0.07) 9.06 0.00 90.94 CLAW-L Claw length 360 1.95 (± 0.07) 12.06 31.65 56.29 TEMP-F Footr temperature 360 1.27 (± 0.06) 1.62 7.01 91.37 TEMP-F Footr temperature 360 1.42 (± 2.61) 0.00 34.59 65.41 LAT-WU Latency to approach mealworms 360	Variable	Description	n	Mean (± SEM)	% group	% bird	% phase within-bird
FTHD-BR Feature arange to breast 360 1.17 (± 0.08) 0.00 11.25 88.75 COMB-EL Comb elevacion 360 2.41 (± 0.08) 0.00 69.14 30.86 COMB-SZ Comb size 360 2.28 (± 0.10) 8.99 68.13 2.288 COMB-COL Comb colour 360 0.07 (± 0.07) 9.06 0.00 9.94 CLAW-L Claw length 360 1.51 (± 0.28) 13.15 35.50 51.34 POSTURE Posture 360 1.95 (± 0.07) 1.266 31.65 56.29 COMD-S Condition score 360 1.71 (± 0.66) 1.62 7.01 9.137 TEMP-F Foot temperature 360 1.82 (± 0.67) 1.24 83.03 1.414 66.54 LAT-MV Latency to approach mealworms 360 1.42 (± 0.60) 0.00 14.55 85.45 LAT-MO Latency to approach nealworms 361 80.33 (± 0.64) 1.71 6.53 91.77 SMO <t< td=""><td>TI</td><td>Tonic immobility</td><td>360</td><td>0.06 (± 0.01)</td><td>0.00</td><td>3.56</td><td>96.44</td></t<>	TI	Tonic immobility	360	0.06 (± 0.01)	0.00	3.56	96.44
COMB-EL Comb elevation 360 2.41 (± 0.08) 0.00 69.14 30.86 COMB-SZ Comb size 360 2.28 (± 0.10) 8.99 68.13 22.88 COMB-COL Comb colour 360 2.00 (± 0.07) 4.89 55.79 39.31 FEET-C Feet condition 360 0.77 (± 0.07) 9.06 0.00 9.04 CLAW-L Claw length 360 0.94 (± 0.20) 0.00 3.361 66.39 COND-S Condition score 360 1.71 (± 0.06) 1.62 7.01 91.37 TEMP-F Foot temperature 360 1.71 (± 0.06) 1.62 7.01 91.37 TEMP-F Foot temperature 360 1.424 (± 2.61) 0.02 31.44 68.54 LAT-MV Latency to approach mealworms 360 1.632 5.00 63.54 LAT <no< td=""> Latency to approach mealworms 310 1.604 (± 0.68) 0.00 1.455 85.45 LAC Blood glucose 319 3.104</no<>	BW	Bodyweight	360	I.74 (± 0.02)	0.00	64.20	35.80
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COND-S Condition score 360 1.95 (± 0.07) 12.06 31.65 56.29 TEMP-E Eye temperature 360 31.71 (± 0.06) 1.62 7.01 91.37 TEMP-F Foot temperature 360 28.20 (± 0.25) 4.25 12.73 83.03 LAT-MW Latency to approach mealworms 360 14.24 (± 2.61) 0.02 31.44 68.54 LAT-NO Latency to approach mealworms 369 16.27 (± 11.28) 3.78 0.00 96.22 GLUC Blood glucose 349 3.64 (± 0.06) 0.00 14.55 85.45 LACT Blood creatine kinase 301 805.38 (± 6.24) 0.00 31.65 91.77 APP Acute phase protein al AG 374 191.62 (± 5.85) 0.00 100.00 HL Heterophiliymphocyte ratio 352 1.00 (± 0.07) 3.18 2.71 94.11 % WATER % facal arbohydrate 351 1.49 (± 0.07) 1.80 6.25 91.95 % FAT % facala	CLAW-L	Claw length	360	15.13 (± 0.28)	13.15	35.50	51.34
TEMP-E Eye temperature 360 31.71 (± 0.06) 1.62 7.01 91.37 TEMP.F Foot temperature 360 28.20 (± 0.25) 4.25 12.73 83.03 LAT-MW Latency to approach mealworms 360 14.24 (± 2.61) 0.02 31.44 68.54 LAT-NO Latency to approach novel object 359 216.27 (± 11.28) 3.78 0.00 96.22 GLUC Blood glucose 349 3.04 (± 0.06) 0.00 14.55 85.45 LACT Blood creatine kinase 301 805.38 (± 6.240) 0.00 34.59 65.41 OSMO Blood creatine kinase 301 805.38 (± 6.41) 1.71 6.53 91.77 APP Acute phase protein al AG 374 191.62 (± 5.85) 0.00 100.00 HL Heterophil:/mphocyte ratio 352 1.00 (± 0.01) 3.18 2.71 94.11 % WATER % facal arbotydrate 351 1.49 (± 0.07) 1.80 6.25 91.95 % CARB <t< td=""><td>POSTURE</td><td>Posture</td><td>360</td><td>0.94 (± 0.02)</td><td>0.00</td><td>33.61</td><td>66.39</td></t<>	POSTURE	Posture	360	0.94 (± 0.02)	0.00	33.61	66.39
TEMP-FFoot temperature36028.20 (± 0.25)4.251.2.7383.03LAT-MWLatency to approach mealworms36014.24 (± 2.61)0.0231.4468.54LAT-NOLatency to approach novel object359216.27 (± 11.28)3.780.0096.22GLUCBlood glucose34913.04 (± 0.06)0.0014.5585.45LACTBlood latate3493.63 (± 0.08)5.907.2886.82CKBlood creatine kinase301805.38 (± 62.40)0.0034.5965.41OSMOBlood osmolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α I AG347191.62 (± 5.85)0.00100.00HLHeterophil:lymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faccal arter35674.90 (± 0.37)4.7525.6869.57% FAT% faccal fat3511.49 (± 0.07)1.806.2091.95% CARB% faccal fat35151.05 (± 0.40)0.350.0094.65DIGEST-PMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FMDigestible fat34676.72 (± 1.41)3.0916.7680.15CORTCormicon-direct3608.95 (± 0.61)7.352.7489.91LGCO-DLacomotion-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDini	COND-S	Condition score	360	l.95 (± 0.07)	12.06	31.65	56.29
LAT-MWLatency to approach mealworms36014.24 (± 2.61)0.0231.4468.54LAT-NOLatency to approach novel object359216.27 (± 11.28)3.780.0096.22GLUCBlood glucose34913.04 (± 0.06)0.0014.5585.45LACTBlood lactate3493.63 (± 0.08)5.907.2886.82CKBlood creatine kinase301805.38 (± 62.40)0.0034.5965.41OSMOBlood somolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α1 AG347191.62 (± 5.85)0.000.00100.00HLHeterophil:lymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faecal vater35674.90 (± 0.37)4.7525.6869.57% PROT% faecal fat3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35059.42 (± 1.22)4.2394.960.81DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat3606.95 (± 1.04)3.9916.7680.15CORTCorricosterone3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3605.96 (± 1.04)8.297.8983.82FORAGE-DForage-direct direct3605.96 (± 1.97)5.448.5685.99 <td>TEMP-E</td> <td>Eye temperature</td> <td>360</td> <td>31.71 (± 0.06)</td> <td>1.62</td> <td>7.01</td> <td>91.37</td>	TEMP-E	Eye temperature	360	31.71 (± 0.06)	1.62	7.01	91.37
LAT-NOLatency to approach novel object359216.27 (± 11.28)3.780.0096.22GLUCBlood glucose34913.04 (± 0.06)0.0014.5585.45LACTBlood lactate3493.63 (± 0.08)5.907.2886.82CKBlood creatine kinase301805.38 (± 62.40)0.0034.5965.41OSMOBlood osmolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein αl AG347191.62 (± 5.85)0.000.00100.00HLHeterophil:lymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faecal vater35674.90 (± 0.37)4.7525.6869.57% PROT% faecal protein3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblifty fat3606.95 (± 1.04)3.0916.7680.15CORTCorricosterone3621.09 (± 0.08)2.071.00.287.90LOCO-DLocomotion-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct3605.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3600.30 (± 0.07)0.009.959.05M-PECK-REC-D <td>TEMP-F</td> <td>Foot temperature</td> <td>360</td> <td>28.20 (± 0.25)</td> <td>4.25</td> <td>12.73</td> <td>83.03</td>	TEMP-F	Foot temperature	360	28.20 (± 0.25)	4.25	12.73	83.03
GLUCBlood glucose34913.04 (± 0.06)0.0014.5585.45LACTBlood lactate3493.63 (± 0.08)5.907.2886.82CKBlood creatine kinase301805.38 (± 62.40)0.0034.5965.41OSMOBlood smolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α1 AG347191.62 (± 5.85)0.000.00100.00HLHeterophiltlymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faccal water35674.90 (± 0.37)4.7525.6869.57% PROT% faccal protein35423.55 (± 0.29)0.0011.1388.87% FAT% faccal atr35151.05 (± 0.40)0.350.009.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat34676.72 (± 1.41)3.0916.7680.15CORTCorricosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 1.41)3.9183.8283.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3603.05 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.55M-PECK-GIVE-DMild-	LAT-MW	Latency to approach mealworms	360	14.24 (± 2.61)	0.02	31.44	68.54
LACTBod locBod creatine kinase3493.63 (± 0.08)5.907.288.682CKBlood creatine kinase301805.38 (± 62.40)0.0034.5965.41OSMOBlood osmolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α1 AG347191.62 (± 5.85)0.000.00100.00HLHeterophil:lymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% facal water35674.90 (± 0.37)4.7525.6869.57% PROT% facal protein35423.55 (± 0.29)0.0011.1388.87% FAT% facal fat3511.49 (± 0.07)1.806.2591.95% CARB% facal carbohydrate35059.42 (± 1.22)4.2394.960.81DIGEST-DMDigestibility fat36676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3221.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct3600.50 (± 0.18)3.498.143.49	LAT-NO	Latency to approach novel object	359	216.27 (± 11.28)	3.78	0.00	96.22
CK OSMOBlood creatine kinase Blood somolality301805.38 (± 62.40)0.0034.5965.41OSMOBlood somolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α1 AG347191.62 (± 5.85)0.000.00100.00HLHeterophil/ymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faecal water35674.90 (± 0.37)4.7525.6869.57% PROT% faecal protein35423.55 (± 0.29)0.0011.1388.87% FAT% faecal fat3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35059.42 (± 1.22)4.2394.960.81DIGEST-DMDigestible dry matter36059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat34676.72 (± 1.41)3.0916.7680.15CORTCorricosterone3241.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct36050.5 (± 0.31)6.740.2792.99M-PECK-REC-DMid-peck received direct3603.01 (± 0.07)0.009.959.005M-PECK-GIVEDMid-peck given direct3600.50 (± 0.18)3.49 <t< td=""><td>GLUC</td><td>Blood glucose</td><td>349</td><td>13.04 (± 0.06)</td><td>0.00</td><td>14.55</td><td>85.45</td></t<>	GLUC	Blood glucose	349	13.04 (± 0.06)	0.00	14.55	85.45
OSMOBlood osmolality324323.68 (± 0.61)1.716.5391.77APPAcute phase protein α I AG347191.62 (± 5.85)0.000.00100.00HLHeterophildymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% facal water35674.90 (± 0.37)4.7525.6869.57% PROT% facal protein35423.55 (± 0.29)0.0011.1388.87% FAT% facal carbohydrate3511.49 (± 0.07)1.806.2591.95% CARB% facal carbohydrate35059.42 (± 1.22)4.2394.960.81DIGEST-DMDigestible dry matter36059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat3608.95 (± 0.41)3.0916.7680.15CORTCorricosterone3608.95 (± 0.41)3.027.908.32LOCO-DLocomotion-direct3608.95 (± 0.41)8.297.898.32FORAGE-DForage-direct3605.06 (± 1.99)5.448.5685.99DRINK-DDrink-direct3603.03 (± 0.07)0.009.959.05M-PECK-REC-DMild-peck given direct3603.06 (± 0.18)3.498.24ALERT-DAlert direct3603.06 (± 0.18)3.498.288.24	LACT	Blood lactate	349	3.63 (± 0.08)	5.90	7.28	86.82
APPAcute phase protein α1 AG347191.62 (± 5.85)0.000.00100.00HLHeterophiliymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% faecal water35674.90 (± 0.37)4.7525.6869.57% PROT% faecal protein35423.55 (± 0.29)0.0011.1388.87% FAT% faecal fat3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.77)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 1.18)3.498.2888.24ALERT-DAlert direct3600.50 (± 1.37)3.0910.1186.80	СК	Blood creatine kinase	301	805.38 (± 62.40)	0.00	34.59	65.41
HLHeterophil:lymphocyte ratio3521.00 (± 0.04)3.182.7194.11% WATER% facal water35674.90 (± 0.37)4.7525.6869.57% PROT% facal protein35423.55 (± 0.29)0.0011.1388.87% FAT% facal fat3511.49 (± 0.07)1.806.2591.95% CARB% facal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestiblity fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-REC-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct3603.069 (± 1.37)3.0910.1186.80	OSMO	Blood osmolality	324	323.68 (± 0.61)	1.71	6.53	91.77
% WATER% faecal water35674.90 (± 0.37)4.7525.6869.57% PROT% faecal protein35423.55 (± 0.29)0.0011.1388.87% FAT% faecal fat3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestibility fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck given direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct3603.06 (± 1.37)3.0910.1186.80	APP	Acute phase protein $\alpha I AG$	347	191.62 (± 5.85)	0.00	0.00	100.00
% PROT % faecal protein 354 23.55 (± 0.29) 0.00 11.13 88.87 % FAT % faecal fat 351 1.49 (± 0.07) 1.80 6.25 91.95 % CARB % faecal carbohydrate 351 51.05 (± 0.40) 0.35 0.00 99.65 DIGEST-DM Digestible dry matter 350 59.42 (± 1.22) 4.23 94.96 0.81 DIGEST-F Digestiblity fat 366 76.72 (± 1.41) 3.09 16.76 80.15 CORT Corticosterone 342 1.09 (± 0.08) 2.07 10.02 87.90 LOCO-D Locomotion-direct 360 8.95 (± 1.61) 7.35 2.74 89.91 COMFORT-D Comfort behaviour-direct 360 50.66 (± 1.99) 5.44 8.56 85.99 DRINK-D Drink-direct 360 1.55 (± 0.31) 6.74 0.27 92.99 M-PECK-REC-D Mild-peck received direct 360 0.30 (± 0.07) 0.00 9.95 90.05 M-PECK-GIVE-D Mild-peck given direct 360 0.50 (± 0.18) 3.49 8.28 88	HL	Heterophil:lymphocyte ratio	352	1.00 (± 0.04)	3.18	2.71	94.11
% FAT% faecal fat3511.49 (± 0.07)1.806.2591.95% CARB% faecal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestibility fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck given direct3600.30 (± 0.07)0.009.9590.05ALERT-DAlert direct3603.069 (± 1.37)3.0910.118.80	% WATER	% faecal water	356	74.90 (± 0.37)	4.75	25.68	69.57
% CARB% faecal carbohydrate35151.05 (± 0.40)0.350.0099.65DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestibility fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3603.06 (± 1.37)3.498.2888.24ALERT-DAlert direct3603.06 (± 1.37)3.0910.1186.80	% PROT	% faecal protein	354	23.55 (± 0.29)	0.00	11.13	88.87
DIGEST-DMDigestible dry matter35059.42 (± 1.22)4.2394.960.81DIGEST-FDigestibility fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct36030.69 (± 1.37)3.0910.1186.80	% FAT	% faecal fat	351	1.49 (± 0.07)	1.80	6.25	91.95
DIGEST-FDigestibility fat34676.72 (± 1.41)3.0916.7680.15CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct36030.69 (± 1.37)3.0910.1186.80	% CARB	% faecal carbohydrate	351	51.05 (± 0.40)	0.35	0.00	99.65
CORTCorticosterone3421.09 (± 0.08)2.0710.0287.90LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct36030.69 (± 1.37)3.0910.1186.80	DIGEST-DM	Digestible dry matter	350	59.42 (± 1.22)	4.23	94.96	0.81
LOCO-DLocomotion-direct3608.95 (± 0.61)7.352.7489.91COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct36030.69 (± 1.37)3.0910.1186.80	DIGEST-F	Digestibility fat	346	76.72 (± 1.41)	3.09	16.76	80.15
COMFORT-DComfort behaviour-direct3606.95 (± 1.04)8.297.8983.82FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct36030.69 (± 1.37)3.0910.1186.80	CORT	Corticosterone	342	1.09 (± 0.08)	2.07	10.02	87.90
FORAGE-DForage-direct36050.66 (± 1.99)5.448.5685.99DRINK-DDrink-direct3601.55 (± 0.31)6.740.2792.99M-PECK-REC-DMild-peck received direct3600.30 (± 0.07)0.009.9590.05M-PECK-GIVE-DMild-peck given direct3600.50 (± 0.18)3.498.2888.24ALERT-DAlert direct36030.69 (± 1.37)3.0910.1186.80	LOCO-D	Locomotion-direct	360	8.95 (± 0.61)	7.35	2.74	89.91
DRINK-D Drink-direct 360 1.55 (± 0.31) 6.74 0.27 92.99 M-PECK-REC-D Mild-peck received direct 360 0.30 (± 0.07) 0.00 9.95 90.05 M-PECK-GIVE-D Mild-peck given direct 360 0.50 (± 0.18) 3.49 8.28 88.24 ALERT-D Alert direct 360 30.69 (± 1.37) 3.09 10.11 86.80	COMFORT-D	Comfort behaviour-direct	360	6.95 (± 1.04)	8.29	7.89	83.82
M-PECK-REC-D Mild-peck received direct 360 0.30 (± 0.07) 0.00 9.95 90.05 M-PECK-GIVE-D Mild-peck given direct 360 0.50 (± 0.18) 3.49 8.28 88.24 ALERT-D Alert direct 360 30.69 (± 1.37) 3.09 10.11 86.80	FORAGE-D	Forage-direct	360	50.66 (± 1.99)	5.44	8.56	85.99
M-PECK-GIVE-D Mild-peck given direct 360 0.50 (± 0.18) 3.49 8.28 88.24 ALERT-D Alert direct 360 30.69 (± 1.37) 3.09 10.11 86.80	DRINK-D	Drink-direct	360	1.55 (± 0.31)	6.74	0.27	92.99
ALERT-D Alert direct 360 30.69 (± 1.37) 3.09 10.11 86.80	M-PECK-REC-D	Mild-peck received direct	360	0.30 (± 0.07)	0.00	9.95	90.05
	M-PECK-GIVE-D	Mild-peck given direct	360	0.50 (± 0.18)	3.49	8.28	88.24
LOCO-NO Locomotion in novel object test 359 10.26 (± 1.21) 37.45 0.00 62.55	ALERT-D	Alert direct	360	30.69 (± 1.37)	3.09	10.11	86.80
	LOCO-NO	Locomotion in novel object test	359	10.26 (± 1.21)	37.45	0.00	62.55
COMFORT-NO Comfort behaviour in novel object test 359 3.48 (± 1.16) 28.11 0.00 71.89	COMFORT-NO	Comfort behaviour in novel object test	359	3.48 (± 1.16)	28.11	0.00	71.89
FORAGE-NO Forage in novel object test 359 20.89 (± 1.90) 9.22 0.00 90.78	FORAGE-NO	Forage in novel object test	359	20.89 (± 1.90)	9.22	0.00	90.78
INT-NO Interact with novel object 359 9.25 (± 0.91) 2.49 0.00 97.51	INT-NO	Interact with novel object	359	9.25 (± 0.91)	2.49	0.00	97.51
DRINK-NO Drink in novel object test 359 I.38 (± 0.33) 6.37 0.00 93.63	DRINK-NO	Drink in novel object test	359	I.38 (± 0.33)	6.37	0.00	93.63
ALERT-NO Alert behaviour in novel object test 359 55.12 (± 3.08) 13.97 0.00 86.03	ALERT-NO	Alert behaviour in novel object test	359	55.12 (± 3.08)	13.97	0.00	86.03
LOCO-MW Locomotion in mealworm test 360 9.30 (± 0.67) 1.33 2.98 95.69	LOCO-MW	Locomotion in mealworm test	360	9.30 (± 0.67)	1.33	2.98	95.69
FORAGE-MW Forage in mealworm test 360 69.34 (± 2.31) 4.77 28.22 67.01	FORAGE-MW	Forage in mealworm test	360	69.34 (± 2.31)	4.77	28.22	67.01

Table 2The percentage variation in each indicator that can be ascribed at the group level, the individual bird level,and the phase within-bird level.

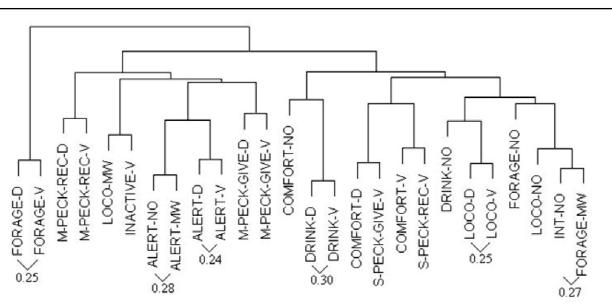
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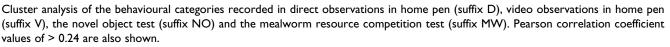
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Table 2 (cont)

Variable	Description	n	Mean (± SEM)	% group	% bird	% phase within bird
ALERT-MW	Alert behaviour in mealworm test	360	20.73 (± 2.27)	7.60	28.51	63.89
LOCO-V	Locomotion-video	360	8.13 (± 0.33)	5.02	14.01	80.97
COMFORT-V	Comfort behaviour-video	360	8.16 (± 0.78)	5.03	6.69	88.28
FORAGE-V	Forage-video	360	63.14 (± 1.55)	2.71	30.55	66.74
DRINK-V	Drink-video	360	4.44 (± 0.27)	0.00	24.30	75.70
M-PECK-REC-V	Mild-peck received-video	360	0.34 (± 0.14)	1.30	23.61	75.09
M-PECK-GIVE-V	Mild-peck given-video	360	0.44 (± 0.20)	5.91	26.37	67.72
S-PECK-REC-V	Severe-peck received-video	360	0.04 (± 0.03)	0.00	0.00	100.0
S-PECK-GIVE-V	Severe-peck given-video	360	0.03 (± 0.02)	3.98	0.00	96.02
INACTIVE-V	Inactive-video	360	0.08 (± 0.04)	1.32	0.00	98.68
ALERT-V	Alert-video	360	15.13 (± 0.82)	0.12	36.80	63.08

Figure I





Comparison of video and direct behaviour observations

An initial cluster analysis examining only the behavioural categories was completed to examine similarities between video and direct observations in all four contexts where behavioural data were collected (ie the DHP, VHP, MW and NO observations). This analysis revealed that the DHP and VHP observations were closely related for the behavioural categories of foraging, alert behaviour, mild pecking, drinking and locomotion (Figure 1).

Relationships between behaviours

Before considering how the behavioural, physiological and physical indicators were related, we examined relationships amongst the behavioural categories in all four contexts where behavioural data were collected (ie the DHP, VHP, MW and NO observations). As animals have a limited time budget, time spent on one activity necessarily reduces the time spent on other categories. We therefore expected, and found, a number of strong negative correlations between

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different behavioural categories. Spearman and Pearson's correlation analyses gave similar results, except for comfort behaviour, and mild and severe pecking, where the number of observations was low and where Spearman tests gave higher coefficient values. Given the large number of tests performed, we present results only where Pearson's values exceed 0.24. At this value, the correlations are significant at P < 0.01 when n = 115. Due to some missing data the number of observations in our tests varied from 301 to 360 resulting in *P*-values of < 0.001.

The time spent foraging was negatively correlated with time spent alert in all four contexts where behavioural data were collected (DHP, VHP, MW and NO observations) with Pearson's correlation coefficients ranging from -0.74 to -0.89. Foraging in the home pen was also negatively correlated with comfort behaviour for the DHP and VHP observations (-0.47, -0.71, respectively), and with locomotion for the VHP, MW and DHP observations (-0.34, -0.40, -0.44). Alert behaviour in the NO test was negatively correlated with drinking, locomotion and interacting with the novel object (-0.31, -0.45, -0.59). The latency to approach the novel object was positively correlated with time spent alert (0.65) and negatively correlated with time spent drinking (-0.29), foraging (-0.39) and interacting with the novel object (-0.66) during the same test. The latency to approach the mealworms was positively correlated with time spent alert during the same test (0.57).

Positive relationships were examined by both correlations and cluster analysis. In addition to the close relationships between direct and video recordings of the same behavioural categories (reported above and in Figure 1), alert behaviour and mild feather pecking appeared to be related, as did comfort behaviour and severe pecking (Figure 1). Alert behaviour in the DHP and VHP observations clustered relatively well with alert behaviour in the NO and MW test, but in contrast, foraging in the home pen appeared almost entirely unrelated to foraging behaviour in the NO and MW tests.

Relationships among welfare indicators

Our main aim was to examine positive and negative relationships between the behavioural, physiological and physical indicators taken. For the undisturbed home pen observations, we used only the behavioural categories recorded by video, because they represented substantially longer periods of observation than the direct observations, and because we knew that the information obtained from direct and video methods was closely related. To reduce the number of variables, we used eye temperature as a proxy for head and comb temperature measures, as eye temperature was positively correlated with both head (0.36) and comb (0.27) temperature. There were some missing observations for the blood indicators (shown in column 3 of Table 2). We dealt with these by performing mean imputation and replacing missing data with the mean values for these indicators. The low number of missing observations meant there was little effect of this procedure on the resulting cluster analysis and allowed us to use all the observations.

Table 3Positive correlations between behavioural,
physiological and physical indicators (acronyms described
in Table 2). Physical measures (green), physiological
measures (red), behavioural measures (blue).

Variable I	Variable 2	Spearman correlation coefficient	Pearson correlation coefficient
BW	COND-S	0.55	0.56
FTHD-BR	ALERT-NO	0.46	0.44
BW	CLAW-L	0.39	0.41
BW	ALERT-NO	0.34	0.36
FTHD-BR F	LAT-NO	0.38	0.35
DIGEST-DM	DIGEST-F	0.39	0.33
TEMP-F	GLUCOSE	0.35	0.32
%WATER	%PROT	0.35	0.32
TEMP-F	%CARB	0.33	0.30
BW	COMB-SZ	0.28	0.29
LACTATE	OSMO	0.44	0.27
TEMP-E	%CARB	0.30	0.27
INT-NO	APP	0.29	0.25
CLAW-L	LAT-NO	0.25	0.27
APP	HL	0.25	0.26
BW	LAT-NO	0.24	0.25
GLUCOSE	LACTATE	0.21	0.25
TEMP-F	APP	0.28	0.24
TEMP-F	HL	0.27	0.24

Positive and negative correlations between the indicators measured, again using cut-offs of 0.24 in magnitude, are shown in Tables 3 and 4. There was generally very good agreement between Spearman and Pearson's correlation coefficients. Relationships between the welfare indicators were examined further by the cluster analysis presented in Figure 2. A joint consideration of the results shown in Table 3 and Figure 2 suggests a cluster of relatively strong links between greater bodyweight, condition score, claw length, breast feather damage and relatively nervous behaviour in the MW and NO tests (indicated by alert behaviour and long latencies to approach). A second important cluster links higher acute phase protein and heterophil:lymphocyte measures with each other, and with relatively confident behaviour in the MW and NO tests, higher body temperature and greater blood glucose concentrations. Lactate and osmolality were closely related (as expected, since osmolality is a measure of blood ion concentration), as were the two digestibility measures. Apparent links between feather pecking and comfort behaviour (Figure 2) were based on low numbers of observations and were not supported by strong correlation coefficient values.

Table 4 Negative between-test correlations (acronyms described in Table 2). Physical measures (green), physiological measures (red), behavioural measures (blue).

Variable I	Variable 2	Spearman	Pearson
		correlation coefficient	
D14/	CLUCOST		
BW	GLUCOSE	-0.38	-0.40
FTHD-BR	APP	-0.47	-0.39
FTHD-BR	INT-NO	-0.39	-0.34
ALERT-NO	APP	-0.38	-0.34
ALERT-NO	FORAGE-MW	-0.35	-0.32
BW	CORT	-0.31	–0.3 l
LAT-NO	TEMP-F	-0.30	-0.29
CLAW-L	GLUCOSE	-0.31	-0.28
LAT-NO	FORAGE-MW	-0.26	-0.27
LAT-NO	APP	-0.30	-0.26
ALERT-NO	TEMP-F	-0.28	-0.26
ALERT-NO	TEMP-F	-0.28	-0.26
LAT-NO	TEMP-H	-0.26	-0.26
FTHD-BR	DRINK-NO	-0.26	-0.25
LAT NO	TEMP-E	-0.26	-0.25
FTHD-BR	HL	-0.28	-0.24
BW	HL	-0.27	-0.24
ALERT-NO	GLUCOSE	-0.26	-0.24

Discussion

Others have reported intra-situational consistency for individual birds in measures of fearfulness, such as tonic immobility and reaction to a novel object (Jones 1988; Ghareeb et al 2008) and we also know that the hens used in this experiment could be grouped into 'personality types' based on differences in their indicator profiles averaged over the entire experimental period (Nicol et al 2009). Different types can be maintained in populations because each has advantages under different environmental conditions. More passive (reactive or 'dove') individuals tend to show lower sympathetic reactivity, higher hypothalamic pituitary adrenal (HPA) reactivity and greater behavioural flexibility than more active (proactive or 'hawk') animals (Koolhaas et al 1999; Korte et al 2009). Given this, the amount of variability in indicator values that occurred at the phase withinbird level was somewhat unexpected. Clearly, the existence of underlying personality traits did not preclude greatly varying responses to housing at different phases of the experiment. However, different behavioural strategies can also be maintained within populations by the same individuals making conditional decisions to play either 'hawk' or 'dove' depending on the context, something not fully

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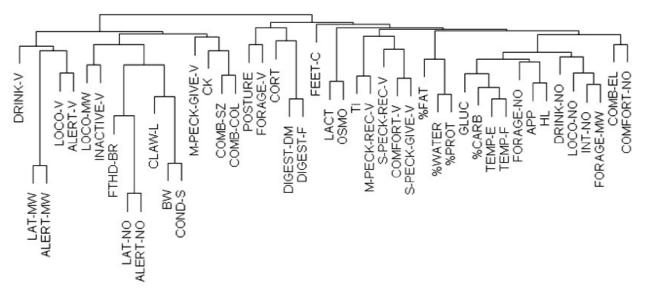
explored by Korte *et al* (2009). We simply draw attention to the fact that the response clusters found here do not perfectly map onto the stable bird 'types' identified previously (Nicol *et al* 2009). We considered ranking responses within phases and looking for consistency in bird rankings. However, because of the design of this experiment, different birds experienced different environments in each phase and we were unable to account for changes in bird age and environment simultaneously to examine individual consistency.

Information about the amount of variation in each indicator that occurred at the various levels in the dataset is important in distinguishing how different indicators can be used in a welfare context. Responses that are good indicators of a stable individual personality or body condition could be used to select birds with more desirable traits (Ghareeb *et al* 2008). In this study, the most stable traits were physical indicators, such as comb elevation and colour. More labile responses that vary with age, or as birds react to different environments, are potentially sensitive indicators of how well a bird is faring in its current living environment.

Our analysis reveals considerable lability in most of the behavioural and physiological measures. The behavioural measures recorded during the MW and NO tests had slightly more stable 'individual bird' or 'group' components than the behavioural measures recorded during undisturbed home pen observations, but all measures still varied greatly between experimental phases. Given the correlations between the MW and NO test responses and the physiological measures, we suggest that these behavioural test responses are more useful welfare indicators than previously acknowledged.

There has been much debate about the degree of independence that is likely when observations are taken on animals living in groups, and how non-independence should be treated statistically (Iason & Elston 2002; Knowles & Green 2002; Phillips 2002). By using hierarchical statistical modelling techniques, we found that variability at the group level was rather low, except for locomotion during the NO test. This suggests that the analysis of welfare indicators based on group means to avoid perceived (but not measured) problems of non-independence, can sometimes unnecessarily reduce the power of a study.

Our main aim was to examine correlations between different classes of welfare indicators. If many different indicators are closely linked this suggests redundancy in the data collection process and it could lead to problems in combining non-independent indicators to draw overall conclusions. We found at least two interesting patterns linking the different indicators across a range of different environments and bird ages. First, slow approach responses and alert behaviour during the MW and, especially, the NO test, were linked with higher bodyweight and condition score, longer claws and greater plumage damage. This 'slow response' cluster was negatively associated with indicators of stress — including lower body temperature, APP, H:L ratio and glucose concentration. Unexpectedly, therefore, a greater reluctance to approach



Cluster analysis of the behaviour recorded in the video observations in the home pen, the novel object test, and the mealworm resource competition test with the physical and physiological indicators recorded in the home pens at each phase.

novel stimuli in a behavioural test situation was strongly and consistently linked with a suite of physiological indicators of a lower stress profile. This conclusion is supported by our previous finding that a longer latency to approach the NO in a particular environment was associated with a positive choice for that same environment (Nicol *et al* 2009) and by more general evidence that there are no clear associations between fear and stress in birds (see Cockrem 2007 for a review). This suggests that novel object responses may be rather good summary indicators of welfare, but may require a very different interpretation from that usually provided.

Some measures, thought to be important for welfare, were seemingly unrelated to others recorded during this study. The most notable example was comfort behaviour, which we know is highly motivated and associated with positive environmental choice in hens (Nicol 1987a,b; Nicol et al 2009) but which appeared uncorrelated with other measures. This suggests that comfort behaviour should be included as an independent measure in welfare assessment protocols. Finally, there were some measures that, in this experiment, were not related to the birds' environmental preferences, or to other welfare indicators. These included the number of attempts to induce a state of tonic immobility and basal blood corticosterone concentrations. Basal corticosterone levels can fluctuate rapidly in response to internal and external events and its usefulness as a welfare measure in hens has been questioned previously (D'Eath et al 2009), but it remains the most widely used physiological parameter in welfare studies. It would be useful in future work to determine whether corticosterone measured after challenge (eg challenge with different dose levels of ACTH) might show stronger associations with other welfare indicators and/or bird preferences.

Another approach to summarising behavioural variation is to use principal components analysis (PCA). Van Reenen et al (2004), for example, used this technique to examine relationships between eight behavioural variables in a dataset examining reactivity of heifer calves. They found principal components (PCs) that loaded heavily on two or three of their eight variables and were fairly interpretable. We also looked at PCA on our larger datasets but the PCs produced were not very easy to interpret and we therefore preferred the cluster diagrams described here. It is our experience that, with larger datasets, PCA is a useful technique for dimension reduction but at a price of interpretability. Also, when using PCA on a set of predictor variables, the PCs produced may explain much of the variability in the predictors themselves, without being any more related to an external response variable than the original predictors. The ideal is to use dimension reduction techniques that capture the variability in the dataset while remaining interpretable. For example, we have recently (Browne et al 2010) worked on dimension reduction techniques that use knowledge of the data and modelling to summarise mass spectrograms as a set of parameters representing important peaks in the spectrograms. These are much easier to interpret and use than PCs from a PCA.

Figure 2

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Animal welfare implications

Statistical analysis of large datasets can inform decisions about whether to record welfare-relevant information at an individual animal or group level, and about which indicators are closely related or independent. This can be used to improve, refine and reduce protocols for experimental and on-farm assessments of animal welfare. Finally, there are welfare implications if the interpretation of commonly used novel object tests proves to be incorrect, especially if such tests form the basis of genetic selection programmes.

Conclusion

Consistent results from correlational and cluster analysis revealed that slow approach responses and alert behaviour in novel object tests were associated with lower levels of physiological stress. Comfort behaviour was an independent measure of welfare not linked to this set. The statistical techniques used revealed the level at which most variation occurred in each welfare indicator and highlighted patterns of independence and redundancy in the measures taken.

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