

2016

Neuroscience 2016
November 12–16, 2016
San Diego, CA
www.sfn.org

Laboratory Diagnosis of Haematological Malignancies
November 16, 2016
London, UK
www.rms.org.uk/discover-engage/eventcalendar/laboratory-diagnosis-2016.html

2016 MRS Fall Meeting & Exhibit
November 27–December 2, 2016
Boston, MA
www.mrs.org/fall2016

American Society for Cell Biology (ASCB) 2016 Annual Meeting
December 3–7, 2016
San Francisco, CA
<http://ascb.org/future-ascb-annual-meetings>

MSSA2016: Microscopy Society of Southern Africa Annual Conference
December 5–9, 2016
Port Elizabeth, South Africa
www.mssa2016.co.za

2017

AMAS XIV - 14th Australian Microbeam Biennial Symposium
February 6–10, 2017
Brisbane, Australia
<http://microscopy.org.au/amas/>

Microscopy & Microanalysis 2017
August 6–10, 2017
St. Louis, MO
www.microscopy.org

2018

Microscopy & Microanalysis 2018
August 5–9, 2018
Baltimore, MD
www.microscopy.org

2019

Microscopy & Microanalysis 2019
August 4–8, 2019
Portland, OR
www.microscopy.org

2020

Microscopy & Microanalysis 2020
August 2–6, 2020
Milwaukee, WI
www.microscopy.org

2021

Microscopy & Microanalysis 2021
August 1–5, 2021
Pittsburgh, PA
www.microscopy.org

More Meetings and Courses

Check the complete calendar near the back of this magazine.

Carmichael's Concise Review

An Underwater Microscope

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Approximately three-quarters of the Earth's surface is covered by oceans. In spite of the enormous size and importance of this ecosystem, we still have much to learn about the biologic systems at play in the ocean. These shortcomings include events that occur at microscopic levels. A major limiting factor is the necessity of studying microscopic events in the laboratory where simulated marine environments are only approximations. A microscope that can be used in the ocean by human scientists could go a long way to fill in this void. Such a microscope has been developed recently by Andrew Mullen, Tali Treibitz, Paul Roberts, Emily Kelly, Rael Horwitz, Jennifer Smith, and Jules Jaffe [1]. They have labeled this exciting instrument the Benthic Underwater Microscope (BUM), and it promises to fill the distinct need to make non-invasive observations of important environmental processes under natural conditions.

The BUM is an imaging system that provides the first *in situ* underwater observations of benthic environments at nearly micrometer resolution. This diver-deployed portable microscope can visualize spatial phenomena in marine environments such as coral reefs and kelp forests. In addition, temporal resolution is achieved with extended time-series recordings lasting several hours. This can reveal gradual or periodic activities and processes, allowing for studies of individual coral polyps and

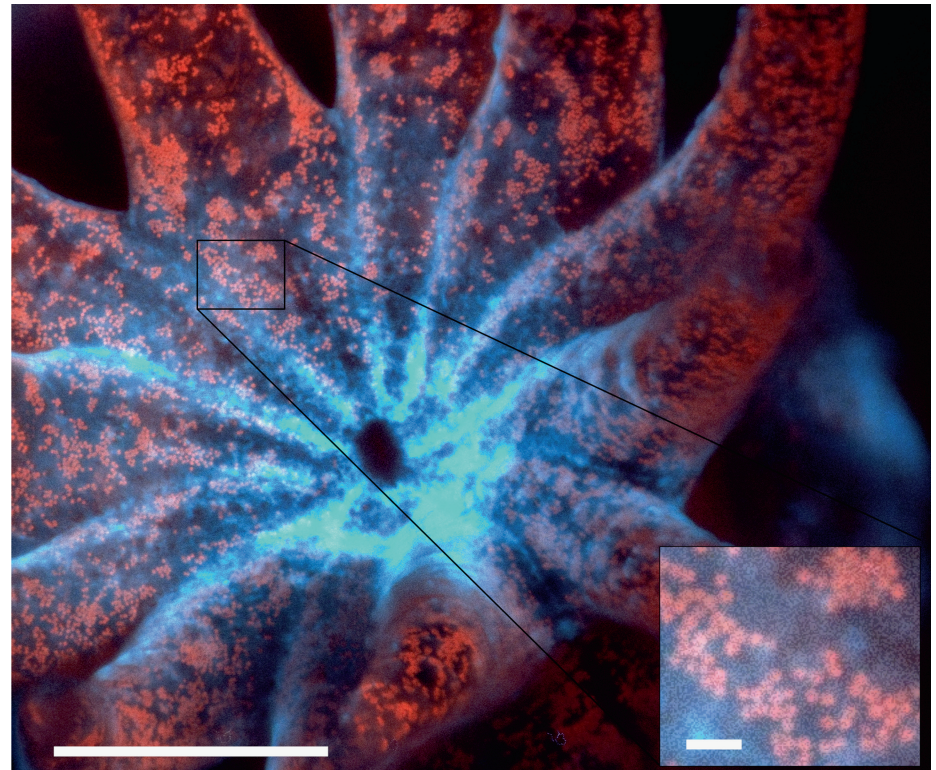
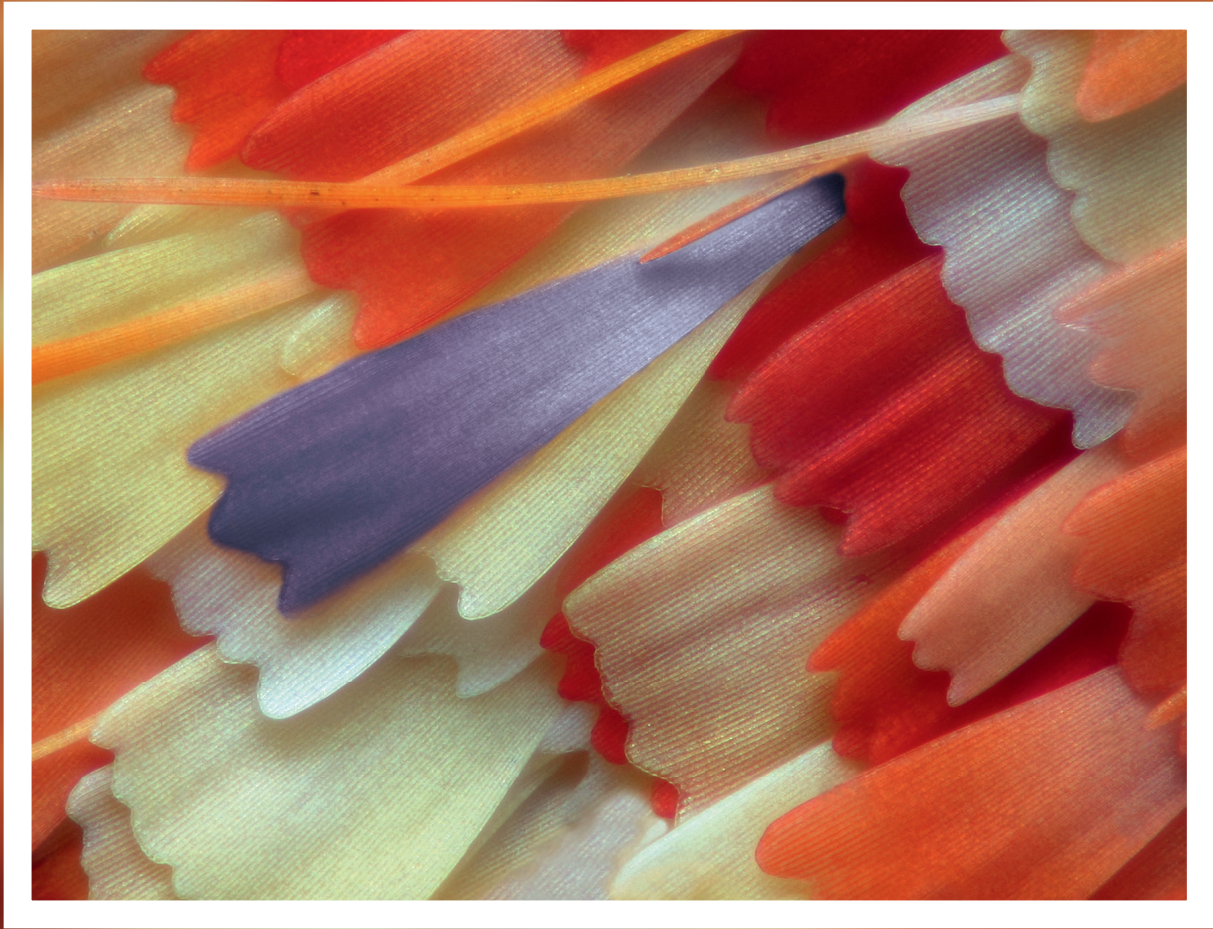


Figure 1: Fluorescent image of the coral *Pocillopora* taken in a lab aquarium using a 5× objective. Individual zooxanthellae (~6–13 μm in size) can be seen emitting red fluorescence from their chlorophyll. Image is a composite focus stack. Main figure scale bar = 500 μm. Inset scale bar = 50 μm. Image credit: Jaffe Lab for Underwater Imaging, Scripps Institution of Oceanography at UC San Diego.



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the behavior of other animals. These impressive capabilities allow both temporal and spatial analysis of ecologically significant phenomena at scales never before seen in the marine environment.

Key challenges in the design of the BUM were overcome through the use of a microscope objective with a long working distance, an electrically tunable lens (ETL) that changes shape to achieve rapid focusing. Focused reflectance illumination is provided by light emitting diodes (LEDs). The desired resolving power requires a high numerical aperture of the objective lens that results in a shallow depth of field. The ETL consists of a flexible polymer encasing an optical fluid. An integrated actuator exerts a variable pressure to rapidly change the lens curvature and focal length on a millisecond time scale. This provides a compact means to bring the subject of interest into precise focus, which is a principle challenge in benthic underwater microscopy. The ETL rapidly scans the focal plane of the optical system, and the frames are then combined using image-processing techniques commonly known as focus stacking. Whereas the 3× and 5× microscope objective lenses provide narrow depths of field (34 μm and 16 μm, respectively), the ETL yields an apparent focus over large depths, 18.4 mm and 6.9 mm, respectively. The ring of LEDs around the lens provides the high-intensity light required for brief reflectance illumination without disturbing the environment.

Several observations were made by Mullen et al. that proved the usefulness of their optical system. For example,

images of live coral polyps in the ocean reveal the distribution and discrimination of individual symbiotic single-celled dinoflagellates, commonly known as zooxanthellae, living inside the coral. With this imaging capability they examined corals experiencing varying levels of bleaching (which is caused by expulsion of zooxanthellae) and checked for the presence of remaining symbionts. Fluorescence imaging was conducted with the microscope in laboratory aquariums by equipping the instrument with blue LEDs combined with a filter in the imaging system to further enhance observations. These and many other studies demonstrated the ability of the BUM to collect ecologically significant spatial and temporal data that can be used for quantitative analysis.

Mullen et al. pointed out that the BUM offers a platform for technology development to facilitate future studies. This technological breakthrough promises to bring laboratory research into the ocean. Such an *in situ* viewpoint enables novel and important investigations on basic marine research and will provide the means to connect theoretical lab work to the natural environment!

References

- [1] A.D. Mullen et al., DOI 10.1038/ncomms12093 (*Nature Communications*, 2016).
- [2] The author gratefully acknowledges Andrew Mullen and Dr. Jules Jaffe for reviewing this article.

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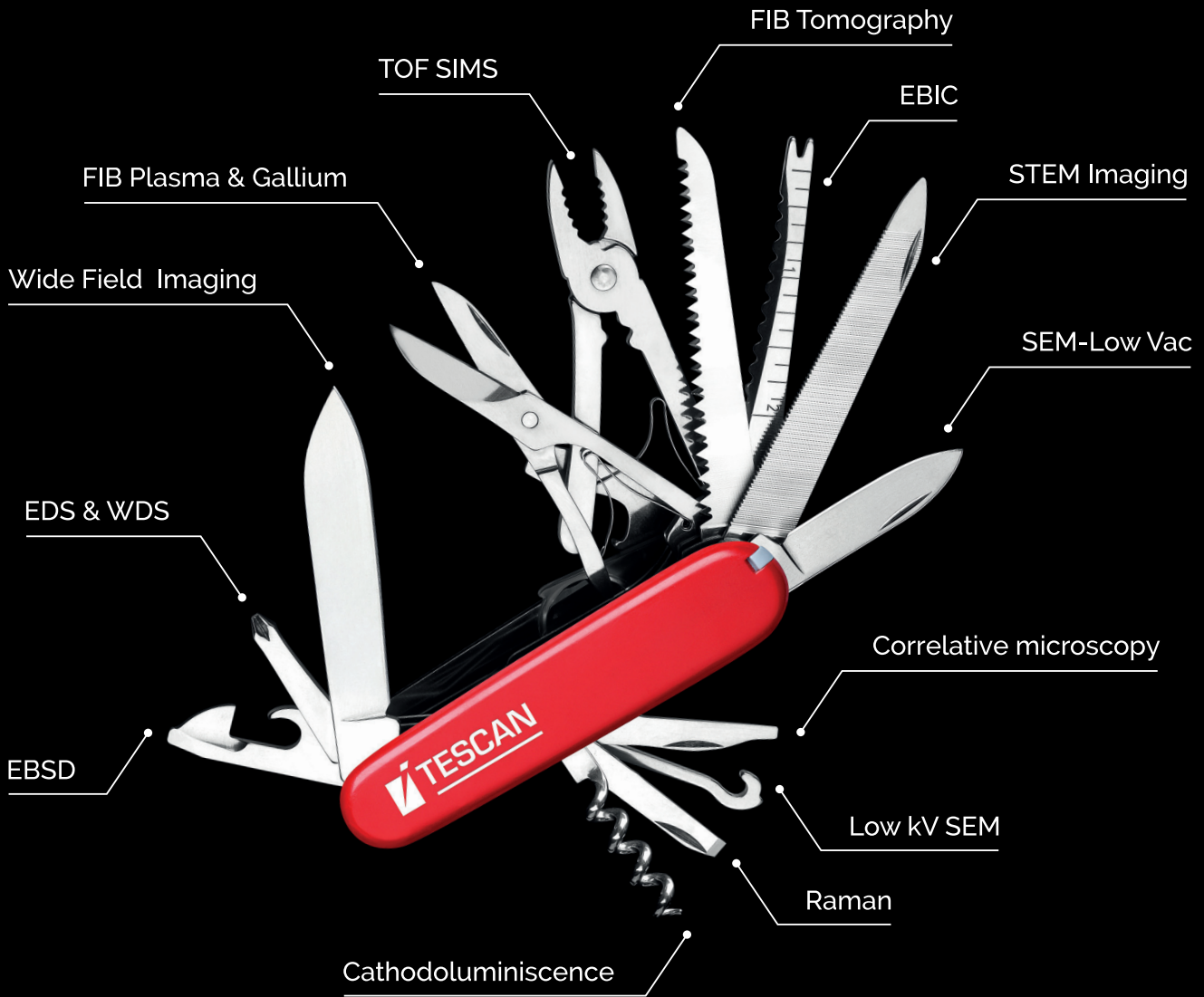
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