Standard Paper

Aptrootia khaoyaiensis (Trypetheliaceae), a new corticolous lichen from the dry dipterocarp forest in central Thailand

Wetchasart Polyiam^{1,4} (b), Santi Watthana¹ (b), Nooduan Muangsan¹ (b), Sittiporn Parnmen² (b) and Robert Lücking³ (b)

¹School of Biology, Institute of Science, Suranaree University of Technology, Suranaree Sub-district, Muang District, Nakhon Ratchasima 30000, Thailand;
²Toxicology Center, National Institute of Health, Department of Medical Sciences, Ministry of Public Health, Nonthaburi 11000, Thailand;
³Botanischer Garten und Botanisches Museum Berlin, Freie Universität Berlin, 14195 Berlin, Germany and ⁴Lichen Research Unit, Department of Biology, Faculty of Science, Ramkhamhaeng University, Bangkok 10240, Thailand

Abstract

Aptrootia khaoyaiensis Polyiam & Lücking is described as new to science from dry dipterocarp forest in central Thailand, based on morphological assessment and sequence data of the mitochondrial small subunit (mtSSU). The new species is characterized by a corticolous habit, a corticate thallus, ascomata typically immersed in the thallus, with a brown to blackish ostiolar region, an inspersed hamathecium, and dark brown, muriform ascospores occurring 1-2 per ascus. Phylogenetic analyses support placement of the new species in *Aptrootia*; it is morphologically close to *A. elatior* but differs in the smaller ascospores and the inspersed hamathecium.

Keywords: biodiversity; phylogeny; taxonomy; tropical rainforest

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Introduction

The lichenized genus *Aptrootia* Lücking & Sipman was introduced by Lücking *et al.* (2007). It is mainly characterized by a crustose thallus, immersed to erumpent ascomata and large, dark brown, muriform ascospores (Aptroot & Lücking 2016). *Aptrootia* is a small genus belonging to the family *Trypetheliaceae*, with only three species known so far: *Aptrootia elatior* (Stirt.) Aptroot, *A. robusta* (P. M. McCarthy & Kantvilas) Aptroot and *A. terricola* (Aptroot) Lücking *et al.* (Lücking *et al.* 2007; Aptroot 2009; Aptroot & Lücking 2016). Members of this genus are corticolous, terricolous or muscicolous and found in tropical areas (Aptroot 1999, 2009; Aptroot & Lücking 2016). Up to now, the genus has not been reported from Thailand (Buaruang *et al.* 2017).

During intensive fieldwork along the gradient of elevational ranges in the dry dipterocarp forest of Khao Yai National Park in central Thailand, an unidentified *Aptrootia* species was discovered which did not correspond to any of the known species. Consequently, we describe this material here as a new species, *Aptrootia khaoyaiensis* sp. nov., based on diagnostic morphological characters and supported by molecular phylogenetic analyses.

Material and Methods

Morphology and anatomy

All specimens studied were collected from tree bark in the dry dipterocarp forest of Khao Yai National Park, Thailand, in

2020–2021. Morphology was examined using an Olympus SZ30 dissecting microscope. Sections of thalli and ascoma were handcut with a razor blade and studied using an Olympus BX41 compound microscope. All measurements were made on material mounted in water. Images were captured with an Olympus Tough TG-6 digital camera. Lichen substances were analyzed by spot tests using K (10% aqueous solution of potassium hydroxide), ultraviolet light (365 nm), and thin-layer chromatography (Orange *et al.* 2010).

DNA extraction, PCR and sequencing

Perithecia were carefully cut from the thallus with a razor blade and ground with a pestle tissue grinder. Genomic DNA was extracted from 5-10 perithecia using the Qiagen DNeasy Plant Mini Kit (Qiagen, Venlo, The Netherlands), according to the manufacturer's instructions. A portion of mycobiont mitochondrial small subunit (mtSSU) was amplified and sequenced using combinations of the primers mrSSU1-AGCAGTGAGGAATAT TGGTC and mrSSU3R-ATGTGGCACGTCTATAGCCC (Zoller et al. 1999). Amplifications were performed in a 25 µl volume and the amplification mixture consisted of 1 µl total DNA, 9.5 µl MasterMix (TaqDNA Polymerase, 25 mM MgCl₂, 8 mM dNTPs, 10× buffer; Thermo Fisher Scientific Inc., USA), 2.5 μ l of each primer (10 μ M) and 9.5 μ l nuFree H₂O. The cycle sequencing conditions were as follows: initial denaturation for 5 min at 96 °C, followed by 1 min at 95 °C, 1 min at 55 °C, 1 min 30 s at 72 °C, and a final extension step of 10 min at 72 °C. The amplification products were visualized on a 1% agarose gel and PCR products were purified using the FavorPrep[™] GEL/PCR Purification Kit (Favorgen, Ping-Tung, Taiwan). Sanger sequencing of the purified amplicons was

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Corresponding author: Wetchasart Polyiam; Email: wetchasart@ru.ac.th

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performed by the Toxicology Center, National Institute of Health, Thailand.

Sequence alignment and phylogenetic analyses

Alignments were carried out using MUSCLE v. 3.8.31 (Edgar 2004). Ambiguously aligned positions were corrected manually. Additional mtSSU sequences belonging to different genera of Trypetheliaceae were downloaded from GenBank (Table 1), their selection based on the present phylogenetic classification of the family (Del Prado et al. 2006; Nelsen et al. 2009, 2014; Luangsuphabool et al. 2016, 2018; Lücking et al. 2016). For the final analysis, we used a subset of selected species from the genera Aptrootia, Architrypethelium, Astrothelium, Bathelium, Pseudopyrenula and Trypethelium. Phylogenetic analyses were conducted using maximum likelihood (ML) and maximum parsimony (MP). ML analysis was performed in IQ-TREE (Trifinopoulos et al. 2016) using the GTR+I+G model, whereas MP was analyzed in MEGA X (Kumar et al. 2018). Branch support obtained from both methods was estimated using 1000 bootstrap pseudoreplicates. Only clades that received bootstrap support $\ge 70\%$ were considered supported. Phylogenetic trees were visualized using FigTree v. 1.4.3 (http://tree.bio.ed.ac.uk/ software/figtree).

Results

Phylogeny

Both ML and MP analyses showed a congruent tree topology, and therefore only the ML tree ($\ln L = -2865.74$) with bootstrap support (BS) > 70% is shown (Fig. 1). The phylogenetic position of *Aptrootia khaoyaiensis* was resolved as an early diverging lineage within *Aptrootia* (ML-BS = 88%; MP-BS = 73%), forming a sister group with *A. elatior*, *A. robusta* and *A. terricola* (Fig. 2). The

relationships within *Aptrootia* were supported, but the backbone relationships between the various genera were largely not supported.

The Species

Aptrootia khaoyaiensis Polyiam & Lücking sp. nov.

MycoBank No.: MB 849988

Similar to *Aptrootia elatior* (Stirt.) Aptroot in growing on bark and producing a corticate thallus, but differing in the smaller ascospores without obvious ornamentation, the fully inspersed hamathecium, and the uneven to verrucose but not bullate thallus.

Type: Thailand, Prachinburi, Prachantakham District, Khao Yai National Park, Klong Peka, dry dipterocarp forest, 14° 11′51″N, 101°29′31″E, *c*. 253 m a.s.l., on bark of *Gluta usitata*, 19 July 2020, *W. Polyiam* 33238 (RAMK—holotype). GenBank Accession no.: OR392814.

(Fig. 2)

Thallus crustose, corticolous, corticate, $35-50 \,\mu\text{m}$ thick, pale yellow-brown to green, uneven to irregularly verrucose, without prothallus. *Photobiont* trentepohlioid.

Ascomata trypethelioid, solitary or irregularly confluent, with apical ostioles, ampulliform (in section), 0.5–1.2 mm diam., immersed to erumpent, largely covered by thallus, ostiolar area black. Ascomata wall black, 40–60 µm thick. Hamathecium gelatinous, fully inspersed with irregular, large oil droplets. Paraphysoids anastomosing, 400–550 µm long. Asci clavate to subcylindrical, with 1–2 ascospores per ascus. Ascospores densely muriform, not ornamented, 80–150 × 30–50 µm, ellipsoid, hyaline but becoming dark brown when mature, I– to I+ pale violet when still hyaline.

Table 1. GenBank Accession numbers and voucher information for samples of *Trypetheliaceae* used in the phylogenetic analyses (Fig. 1). New sequence generated for the present study is indicated in bold.

Taxon	Country of origin	Specimen voucher	GenBank Accession no.	Reference
Aptrootia elatior	New Zealand	Knight O61815 (OTA)	KM453821	Nelsen <i>et al.</i> 2014
A. khaoyaiensis	Thailand	Polyiam 33238 (RAMK)	OR392814	Present study
A. robusta	Australia	Lumbsch 20012 (F)	KM453822	Nelsen <i>et al.</i> 2014
A. terricola	Costa Rica	Lücking 17211 (F)	DQ328995	Del Prado et al. 2006
Architrypethelium lauropaluanum	Peru	Nelsen Cit1P (F)	KX215566	Lücking et al. 2016
A. murisporum	Thailand	Luangsuphabool 031332 (RAMK)	LC361339	Luangsuphabool et al. 2018
A. nitens	Panama	Lücking 27038 (F)	KM453823	Nelsen et al. 2014
Astrothelium aenascens	Thailand	Luangsuphabool 027887 (RAMK)	LC128018	Luangsuphabool et al. 2016
A. macrostiolatum	Thailand	Luangsuphabool 027895 (RAMK)	LC128022	Luangsuphabool et al. 2016
A. siamense	Thailand	Luangsuphabool 027901 (RAMK)	LC128020	Luangsuphabool et al. 2016
Bathelium lineare	Vietnam	Gueidan 2078 (F)	KM453839	Nelsen et al. 2014
B. madreporiforme	Brazil	Lücking 23290 (F)	KM453840	Nelsen et al. 2014
Pseudopyrenula diluta	Venezuela	Lücking 26062 (F)	KM453861	Nelsen et al. 2014
P. subgregaria	Thailand	Lücking 24079 (F)	GU327699	Nelsen et al. 2009
Trypethelium eluteriae	Australia	Lumbsch 19112 g (F)	DQ328990	Del Prado et al. 2006
T. subeluteriae	Costa Rica	Lücking 17611 (F)	DQ329009	Del Prado et al. 2006

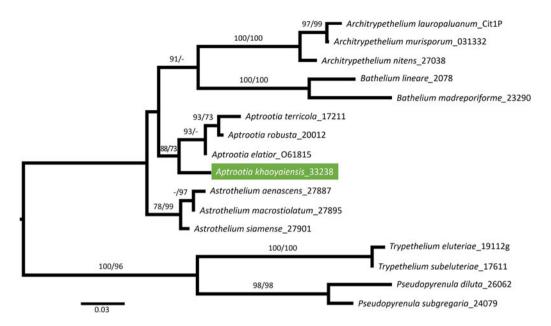


Figure 1. Maximum likelihood (ML) tree based on mtSSU gene sequences showing the position of *Aptrootia khaoyaiensis* within the *Trypetheliaceae*. The numbers at branch nodes indicate bootstrap percentages resulting from 1000 replications. ML and MP bootstrap values \geq 70% are indicated above the branches (ML/MP). Specimen voucher numbers are given after species names. In colour online.

Conidiomata not observed.

Chemistry. K-, UV-. TLC: no lichen substances detected.

Etymology. The specific epithet is a reference to Khao Yai National Park where the holotype was collected.

Ecology and distribution. The specimens studied were collected from tree bark between 200 and 400 m altitude in dry dipterocarp forest with open canopy, growing on the bark of *Dipterocarpus obtusifolius* and *Gluta usitata*, indicating that they prefer hard, thick and acid tree barks (Wolseley & Aguirre-Hudson 1997). The topography of the area consists of inclining slopes, sandy soil and sandy rocks on the ground (Fig. 1). The area is frequently affected by forest fires during the dry season.

Remarks. Aptrootia khaoyaiensis is characterized by its corticolous habit, its corticate, smooth to uneven thallus, its inspersed hamathecium and dark brown, densely muriform ascospores. The new species is most similar to *A. elatior* in external morphology and substratum ecology (Aptroot & Lücking 2016). The

latter, however, differs in having ornamented and much larger ascospores $(200-330 \times 60-90 \ \mu\text{m})$, a largely clear hamathecium (except for oil droplets near the ostiole), and the thallus is bullate. *Aptrootia terricola* and *A. robusta* grow on soil or bryophytes, their thallus is smooth to uneven, rather thin and greyish, they have a clear hamathecium and their ascospores, although also with smooth walls, are larger $(150-400 \times 40-140 \ \mu\text{m})$ (Aptroot & Lücking 2016). The new species is also superficially similar to *Anthracothecium macrosporum* (Hepp) Müll. Arg. in the erumpent perithecia covered by thallus tissue, but differs in the conical shape of the perithecia, the unbranched periphysoids, and the clear hamathecium (Awasthi 1991; Aptroot *et al.* 2008).

Additional specimens examined. **Thailand:** Prachinburi Province: Prachantakham District, Khao Yai National Park, 14° 11′51″N, 101°29′31″E, 253 m a.s.l., dry dipterocarp forest, on bark of *D. obtusifolius* Teijsm. ex Miq., 2021, *W. Polyiam* 33237 (RAMK); *ibid.*, 14°11′54″N, 101°29′26″E, 214 m a.s.l., dry dipterocarp forest, on bark of *D. obtusifolius*, 2020, *W. Polyiam* 32239 (RAMK).

Key to the species of Aptrootia

1	Thallus corticolous 2 Thallus terricolous or muscicolous 3
2(1)	Thallus smooth to uneven; hymenium fully inspersed; ascospores not ornamented, $80-150 \times 30-50 \ \mu m$ A. khaoyaiensis
	Thallus vertucose to bullate; hymenium inspersed only near the ostiole; ascospores ornamented, $200-330 \times 60-90 \ \mu m$ A. elation
3(1)	Ascospores 170–230 × 40–70 μm

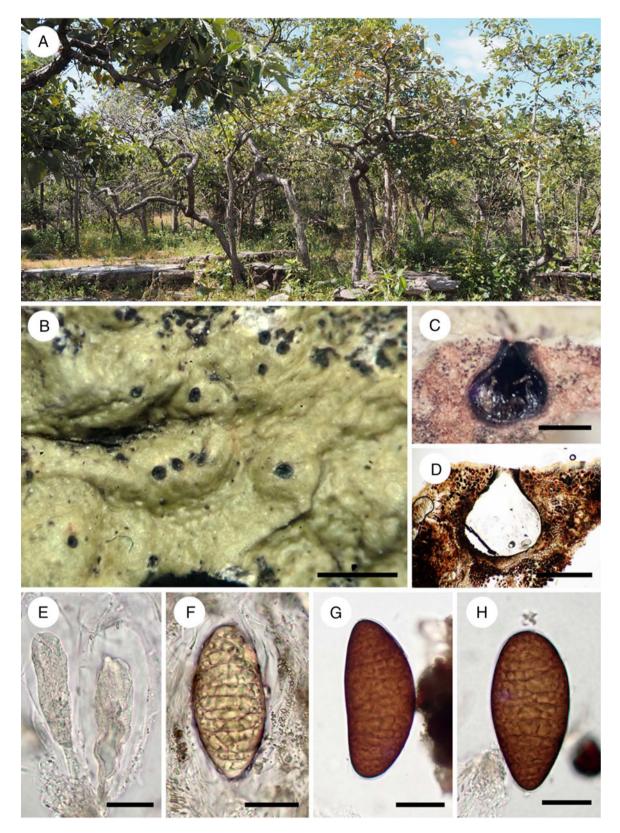


Figure 2. Aptrootia khaoyaiensis (RAMK 33238—holotype). A, habitat of the holotype dominated by *Dipterocarpus obtusifolius*. B, perithecia and thallus structure. C & D, sections through perithecia. E, asci. F, pale brown ascospore surrounded by paraphysoids and oil drops. G & H, dark brown ascospores. Scales: B = 1 mm; C & D = 300 μm; E–H = 30 μm. In colour online.

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Author ORCIDs. D Wetchasart Polyiam, 0009-0002-5593-4508; Santi Watthana, 0000-0002-8837-1820; Nooduan Muangsan, 0000-0002-4669-3119; Sittiporn Parnmen, 0000-0002-0116-9436; Robert Lücking, 0000-0002-3431-4636.

References

- Aptroot A (1999) Thelenella terricola, a new saprobic ascomycete from upland Papua New Guinea. Fungal Diversity 2, 43–46.
- Aptroot A (2009) *Trypetheliales*. In McCarthy PM and Kuchlmayr B (eds), *Flora of Australia. Vol. 57, Lichens 5.* Melbourne: ABRS and CSIRO Publishing, pp. 535–552.
- Aptroot A and Lücking R (2016) A revisionary synopsis of the Trypetheliaceae (Ascomycota: Trypetheliales). Lichenologist 48, 763–982.
- Aptroot A, Lücking R, Sipman HJM, Umaña L and Chaves JL (2008) Pyrenocarpous lichens with bitunicate asci. A first assessment of the lichen biodiversity inventory in Costa Rica. *Bibliotheca Lichenologica* 97, 1–162.
- Awasthi DD (1991) A key to the microlichens of India, Nepal and Sri Lanka. Bibliotheca Lichenologica 40, 1–337.
- Buaruang K, Boonpragob K, Mongkolsuk P, Sangvichien E, Vongshewarat K, Polyiam W, Rangsiruji A, Saipunkaew W, Naksuwankul K, Kalb J, et al. (2017) A new checklist of lichenized fungi occurring in Thailand. MycoKeys 23, 1–91.
- Del Prado R, Schmitt I, Kautz S, Palice Z, Lücking R and Lumbsch HT (2006) Molecular data place *Trypetheliaceae* in *Dothideomycetes*. *Mycological Research* 110, 511–520.
- Edgar RC (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* **32**, 1792–1797.
- Kumar S, Stecher G, Li M, Knyaz C and Tamura K (2018) MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* 35, 1547–1549.

- Luangsuphabool T, Lumbsch HT, Aptroot A, Piapukiew J and Sangvichien E (2016) Five new species and one new record of *Astrothelium* (*Trypetheliaceae, Ascomycota*) from Thailand. *Lichenologist* **48**, 727–737.
- Luangsuphabool T, Lumbsch HT, Piapukiew J and Sangvichien E (2018) Architrypethelium murisporum (Ascomycota, Trypetheliaceae), a remarkable new lichen species from Thailand challenging ascospore septation as an indicator of phylogenetic relationships. *MycoKeys* **34**, 25–34.
- Lücking R, Sipman HJM, Umaña L, Chaves J-L and Lumbsch HT (2007) Aptrootia (Dothideomycetes: Trypetheliaceae), a new genus of pyrenocarpous lichens for Thelenella terricola. Lichenologist 39, 187–193.
- Lücking R, Nelsen MP, Aptroot A, Barillas de Klee R, Bawingan PA, Benatti MN, Binh NQ, Bungartz F, Cáceres MES, Canêz LS, et al. (2016) A phylogenetic framework for reassessing generic concepts and species delimitation in the lichenized family *Trypetheliaceae (Ascomycota: Dothideomycetes)*. *Lichenologist* 48, 739–762.
- Nelsen MP, Lücking R, Grube M, Mbatchou JS, Muggia L, Rivas Plata E and Lumbsch HT (2009) Unravelling the phylogenetic relationships of lichenised fungi in *Dothideomyceta*. Studies in Mycology 64, 135–144.
- Nelsen MP, Lücking R, Aptroot A, Andrew CJ, Cáceres MES, Rivas Plata E, Gueidan C, Canêz LS, Knight A, Ludwig LR, et al. (2014) Elucidating phylogenetic relationships and genus-level classification within the fungal family *Trypetheliaceae (Ascomycota: Dothideomycetes)*. Taxon 63, 974–992.
- **Orange A, James PW and White FJ** (2010) *Microchemical Methods for the Identification of Lichens*, 2nd edn. London: British Lichen Society.
- Trifinopoulos J, Nguyen LT, von Haeseler A and Minh BQ (2016) W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Research 44, W232–W235.
- Wolseley PA and Aguirre-Hudson B (1997) Fire in tropical dry forests: corticolous lichens as indicators of recent ecological changes in Thailand. *Journal of Biogeography* 24, 345–362.
- Zoller S, Scheidegger C and Sperisen C (1999) PCR primers for the amplification of mitochondrial small subunit ribosomal DNA of lichen-forming ascomycetes. *Lichenologist* **31**, 511–516.