

SHORT REPORT

Staphylococcus aureus nasal carriage rate and associated risk factors in individuals in the community

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SUMMARY

The increasing prevalence of *Staphylococcus aureus* and methicillin-resistant *S. aureus* (MRSA) strains together with their disease impact on hospital patients and individuals in the community has posed a major challenge to healthcare workers. This study examined the prevalence of *S. aureus* nasal carriage, antimicrobial susceptibility patterns, and possible risk factors in the community. Of 500 studied subjects (aged from 6 to 65 years) in Lebanon, the overall *S. aureus* nasal carriage rate was 38·4%, the highest (57·1%) being in children aged 6–10 years. Only eight individuals (1·6%) were carriers of MRSA. Risk factors for *S. aureus* nasal colonization were male gender, young age, contact with healthcare workers, use of needle injections, and having asthma. A significant decrease in colonization rate was associated with nasal wash with water, use of nasal sprays, and the presence of acne. These findings may assist in better understanding of control measures to decrease nasal colonization with *S. aureus* in Lebanon and elsewhere.

Key words: Colonization, Lebanon, MRSA, risk factors, *Staphylococcus aureus*.

Staphylococcus aureus has long been recognized as one of the most common causes of both endemic and epidemic infections acquired in hospitals, resulting in substantial morbidity and mortality [1, 2]. This is exacerbated by the increasing appearance of multi-drug-resistant strains especially those with resistance to methicillin (MRSA) which represent a serious clinical threat and therapeutic challenge not only to hospitalized patients but also to adults and children in the community [3, 4]. The epidemiology of MRSA has changed radically in recent years with the definition of two groups of strain populations, hospital acquired and community acquired. The former is

mostly associated with infections in the healthcare setting while the latter has been increasingly reported in healthy persons living in the community and is not associated with traditional MRSA risk factors (i.e. contact with healthcare facilities, previous antimicrobial therapy). These infections most often present as skin and soft tissue infections but occasionally as primary pneumonia with high morbidity and mortality [5].

Carriage of *S. aureus* in the nasal passages appears to play a key role in the epidemiology and pathogenesis of infection [1]. Colonizing strains may serve as endogenous reservoirs for overt clinical infections or may spread to other patients [6]. Data on the carriage rate and antibiotic susceptibility pattern of *S. aureus* strains in the Lebanese community are scarce [7]. This prospective study was therefore undertaken to

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determine the current carriage rate of *S. aureus* [methicillin-susceptible *S. aureus* and MRSA] in school and university students and employees working in these institutions. The possible contributing risk factors involved in colonization were also assessed.

In total 500 nasal swab samples were collected from school and university students and employees in the cities of Beirut and Sidon between September 2006 and March 2007. All participants were interviewed and a questionnaire was completed detailing name, age, gender, previous or current antibiotic treatment, history of hospital admission and previous surgery, contact with healthcare workers (HCWs), needle injections, daily nasal wash with water, use of nasal medication sprays, having acne problems, smoking, and suffering from asthma.

Sterile cotton swabs with Stuart's transport medium (Deltalab, Spain) were used for collection and transport of specimens. Sampling was performed by rotating a swab pre-wetted with sterile saline in the nares of each participant. The swabs were transported at 4 °C to the microbiology laboratory for culture, most within 4 h of collection. They were streaked onto mannitol salt agar (MSA; Oxoid, UK), incubated at 35 °C and examined for growth after 24–48 h. Mannitol-fermenting colonies were subcultured to nutrient agar and incubated at 35 °C for 24 h. Isolates were identified as *S. aureus* by Gram stain, production of catalase, DNase, and coagulase in tube tests.

Susceptibility testing was performed by the disk diffusion method according to Clinical and Laboratory Standards Institute (CLSI) guidelines [8] with *S. aureus* (ATCC 25923) as a control strain. Isolates were screened for susceptibility to amoxicillin/clavulanic acid, ampicillin, cefoxitin, cephalothin, ciprofloxacin, clindamycin, erythromycin, gentamicin, nitrofurantoin, oxacillin, penicillin, rifampicin, trimethoprim/sulfamethoxazole, tetracycline, fusidic acid, mupirocin, and vancomycin. Interpretation of zone inhibition diameters was according to CLSI guidelines except for fusidic acid [9] and mupirocin [10]. Isolates resistant to 1 µg oxacillin and 30 µg cefoxitin were classified as MRSA [8]. Induced clindamycin resistance was inferred by the appearance of the inhibition zone between erythromycin and clindamycin disks [8]. Isolates were screened for resistance to vancomycin on brain heart infusion agar containing 6 µg/ml vancomycin [8].

Data were analysed using Yates corrected χ^2 test. *P* values <0.05 were taken as significant. Statistical analysis was performed using Minitab and Epi-Info

3.4.3 soft ware (Centers for Disease Control and Prevention and World Health Organization).

Of the 500 participants in this study, 277 (55.4%) were females and 223 (44.6%) were males ranging in age from 6 to 65 years. Overall, 192 individuals (38.4%, 95% CI 34.1–42.8) were colonized by *S. aureus*, of which eight isolates (4.2%) were confirmed as MRSA. The age of individuals was significantly associated with *S. aureus* carriage ($P < 0.001$). Subjects aged 6–10 years had a higher carriage rate (57.1%, 95% CI 48.0–65.9), compared to 11–17 years (34.9%, 95% CI 28.1–42.3), 18–25 years (24.8%, 95% CI 17.8–32.9), 26–40 years (37.0%, 95% CI 19.4–57.6), and 41–65 years (45.8%, 95% CI 25.6–67.2), respectively. The higher colonization rate therefore extended to those of school age (aged 6–17 years), declined during university attendance, and increased again in later life (aged 18–65 years) (43.9% vs. 29.3%, $P = 0.001$). Males showed significantly higher ($P = 0.02$) *S. aureus* colonization rates (43.5%) than females (34.3%).

The overall carriage rate for *S. aureus* of 38.4% exceeded rates reported from several other countries such as Saudi Arabia (20.2%) [11], Italy (25.9%) [12] and the USA (31.6%) [13]. Moreover, the current carriage rate is higher than the 20% rate found in a survey of Lebanese individuals in the community in 1993 [7]. The latter study reported a MRSA colonization rate of 0.3% compared to the 1.6% rate found in the present study.

Children have been reported to have increasing rates of infection with *S. aureus* and MRSA [5]. In Lebanon and the wider region, the effect of age on colonization rate had not been previously addressed. Reports from other countries also cite high rates of *S. aureus* colonization in children, e.g. 35% in children aged 3–11 years in Italy [12], and 28.4% in the 4–6 years age group in Turkey [14]. The significant difference in carriage between the school community subjects and the adult general population may be due to the closed school community environment facilitating the spread of bacteria.

Table 1 shows that the risk factors for a high carriage rate of *S. aureus* were: use of antibiotics in the last 6 weeks, hospital admittance in the last year, regular contact with HCWs, using needle injections, or having asthma. Lower colonization rates were significantly associated with participants having nasal wash with water more than twice daily, regular use of nasal sprays, and the presence of acne ($P \leq 0.02$). Smoking and previous surgical operations did not

Table 1. Factors associated with *Staphylococcus aureus* colonization in a Lebanese community

Factors	Total participants (n)	Carriers of <i>S. aureus</i> , % (95% CI)	P
Factors associated with high carriage rate			
Taking antibiotic during last 6 weeks			0.12
No	389	37.0 (32.2–42.1)	
Yes	111	43.2 (33.9–53.0)	
Hospital admission in past 12 months			0.29
No	440	38.0 (33.4–42.7)	
Yes	60	41.7 (29.1–55.1)	
Contact with healthcare workers			0.006
No contact or rarely	468	37.6 (33.2–42.2)	
Always	32	50.0 (31.9–68.1)	
Using needle injections			0.01
Never	420	36.2 (31.6–41.0)	
Rarely or always	80	50.0 (38.6–61.4)	
Asthma			0.068
No	484	37.8 (33.5–42.3)	
Yes	16	56.3 (29.9–80.2)	
Factors associated with low carriage rate			
Nasal wash (by water)			
Never	163	44.8 (37.0–52.8)	
Twice daily	130	40.0 (31.5–49.0)	0.2*
More than twice daily	207	32.4 (26.0–39.2)	0.007*
Using nasal spray			
Never	397	39.5 (34.7–44.6)	
Sometimes	86	38.4 (28.1–49.5)	0.4*
Always	17	11.8 (1.5–36.4)	0.02*
Acne			
Never	381	42.5 (37.5–47.7)	
Sometimes	81	28.4 (18.9–39.5)	0.009*
Always	38	18.4 (7.7–34.3)	0.002*
Factors not associated with carriage rate			
Smoking			
Never	434	(38.7) (34.1–43.5)	
Sometimes	14	(28.6) (8.4–58.1)	0.7*
Always	36	(38.9) (23.1–56.5)	0.5*
Hubble bubble smoking	16	(37.5) (15.2–64.6)	0.5*
History of surgery			0.7
No	382	(38.0) (33.1–43.1)	
Yes	118	(39.8) (30.9–49.3)	

CI, Confidence interval.

* Comparing the corresponding factor to the never factor.

appear to be associated with *S. aureus* nasal colonization.

The higher colonization rate in individuals in regular contact with HCWs may be explained by contact of the latter with patients, colonized or infected by *S. aureus*, who as a result have higher rates of nasal carriage [15]. This reinforces the need for HCWs to be regularly screened for MRSA and, if positive,

undergo decontamination to prevent further transmission of these organisms to others in the general population and the hospital. Impaired immune function or corticosteroid use by asthmatics, and/or their more frequent contact with HCWs could be an explanation for increased carriage in this group.

To our knowledge, this is the first time that nasal washing with water has been shown to affect *S. aureus*

nasal colonization. Water may decrease the adherence of *S. aureus* by interrupting the physicochemical forces including hydrophobic interaction needed for bacterial adherence [16] resulting in the elimination of the organism. Similarly, nasal medication sprays may mechanically decrease adherence of *S. aureus* to nasal mucosa but the contribution of the vehicle or medication to lower carriage rates should be considered. The correlation of acne with low carriage rates is an interesting finding and may be related to competition between *S. aureus* and *Propionibacterium* spp., which are frequent colonists of the anterior nares of individuals, as increased staphylococcal numbers have been reported to be associated with a parallel reduction of the number of *Propionibacterium* [17]. Tobacco smoking appears to have a noticeable effect on the microbial ecology of the nose [18]. Generally, smokers harboured a greater number of *S. aureus* (judged by the primary growth on MSA) but no statistical association was confirmed for their colonization rates over non-smokers.

Apart from a very low rate of susceptibility (5.7%) to penicillin and ampicillin, most isolates were susceptible ($\geq 94\%$) to all other antimicrobials tested except for erythromycin (82.8%), clindamycin (86.5%), fusidic acid (91.1%), and tetracycline (92.2%); no resistance to trimethoprim/sulfamethoxazole, mupirocin and vancomycin was observed. In addition, all MRSA isolates were susceptible to gentamicin, ciprofloxacin and nitrofurantoin.

In conclusion, this study sheds light on *S. aureus* carriage rates in a Lebanese community and establishes a basis for future surveillance of carriage. Despite a relatively high overall rate compared to some other countries, the level of MRSA carriage remains very low. Further investigations are needed to evaluate the effectiveness of nasal wash with water and use of nasal sprays in reducing colonization rates by *S. aureus* especially in HCWs. This might be useful as a hygiene measure in tandem with effective hand washing. Finally, the unexpected relationship between acne and decreased *S. aureus* carriage rate and the possible influence of *P. acnes* warrants further investigation.

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DECLARATION OF INTEREST

None.

REFERENCES

1. Kluytmans J, van Belkum A, Verbrugh H. Nasal carriage of *Staphylococcus aureus*: epidemiology, underlying mechanisms, and associated risk. *Clinical Microbiology Review* 1997; **10**: 505–520.
2. Klein E, Smith DL, Laxminarayan R. Hospitalizations and deaths caused by methicillin-resistant *Staphylococcus aureus*, United States, 1999–2005. *Emerging Infectious Diseases* 2007; **13**: 1840–1846.
3. Lowy FD. *Staphylococcus aureus* infections. *New England Journal of Medicine* 1998; **339**: 520–532.
4. Lu PL, et al. Risk factors and molecular analysis of community methicillin-resistant *Staphylococcus aureus* carriage. *Journal of Clinical Microbiology* 2005; **43**: 132–139.
5. Gorwitz RJ. Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and update. *Pediatric Infectious Disease Journal* 2008; **27**: 925–926.
6. Eiff CV, et al. Nasal carriage as a source of *Staphylococcus aureus* bacteremia. Study group. *New England Journal of Medicine* 2001; **344**: 11–16.
7. Araj GF, et al. Carriage rates, antimicrobial susceptibility and molecular characterization of *Staphylococcus aureus* in Lebanon. *British Medical Journal (Middle East)* 1997; **4**: 17–23.
8. National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial disk susceptibility testing: approved standards, 16th edn. Wayne, PA: NCCLS, 2006; document M100-S16.
9. Andrews JM. BSAC Working party on susceptibility testing. BSAC standardized disc susceptibility testing method. *Journal of Antimicrobial Chemotherapy* 2001; **48**: S43–S47.
10. Udo EE, Jacob LE, Mathew B. Genetic analysis of methicillin-resistant *Staphylococcus aureus* expressing high- and low-level mupirocin resistance. *Journal of Medical Microbiology* 2001; **50**: 909–915.
11. Panhotra BR, Saxena AK, Al Mulhim AS. Prevalence of methicillin-resistant and methicillin-sensitive *Staphylococcus aureus* nasal colonization among patients at the time of admission to the hospital. *Annals of Saudi Medicine* 2005; **25**: 304–308.
12. Zanelli GA, et al. *Staphylococcus aureus* nasal carriage in the community: a survey from central Italy. *Epidemiology and Infection* 2002; **129**: 417–420.
13. Graham III PL, Lin SX, Larson EL. A U.S. population-based survey of *Staphylococcus aureus* colonization. *Annals of Internal Medicine* 2006; **144**: 318–325.
14. Ciftci IH, et al. Nasal carriage of *Staphylococcus aureus* in 4–6 age groups in healthy children in Afyonkarahisar, Turkey. *Acta Paediatrica* 2007; **96**: 1043–1046.

15. **Cesur S, Cokca F.** Nasal carriage of methicillin-resistant *Staphylococcus aureus* among hospital staff and outpatients. *Infection Control and Hospital Epidemiology* 2004; **25**: 169–171.
16. **Beachy H.** Bacterial adherence: adhesin receptor interaction mediating the attachment of bacteria to mucosal surface. *Journal of Infectious Diseases* 1981; **143**: 325–344.
17. **Coates P, et al.** Efficacy of oral isotretinoin in the control of skin and nasal colonization by antibiotic-resistant propionibacteria in patients with acne. *British Journal of Dermatology* 2005; **153**: 1126–1136.
18. **Durmaz RM, et al.** Nasal carriage of methicillin-resistant *Staphylococcus aureus* among smokers and cigarette factory workers. *New Microbiology* 2001; **24**: 143–147.