



Research Article

Cite this article: Narciso RB, Smit NJ, Perbiche-Neves G, da Silva RJ (2024). Integrative taxonomy approach to the study of parasitic ergasilids (Cyclopoida: Ergasilidae) of fishes from the Pardo River, Brazil with a redescription of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 and a molecular phylogeny for Ergasilidae. *Parasitology* 1–24. <https://doi.org/10.1017/S003118202400129X>

Received: 7 July 2024

Revised: 20 September 2024

Accepted: 2 October 2024



Keywords:

copepoda; freshwater fish parasite; *Gamispatulus*; mitochondrial gene; Neotropical region; ribosomal gene

Corresponding author:

Nico J. Smit;
Email: nico.smit@nwu.ac.za

Integrative taxonomy approach to the study of parasitic ergasilids (Cyclopoida: Ergasilidae) of fishes from the Pardo River, Brazil with a redescription of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 and a molecular phylogeny for Ergasilidae

Rodrigo Bravin Narciso^{1,2} , Nico J. Smit² , Gilmar Perbiche-Neves³ and Reinaldo José da Silva¹

¹Section of Parasitology, Institute of Biosciences, São Paulo State University (UNESP), Botucatu, São Paulo, Brazil;

²Water Research Group, Unit for Environmental Sciences and Management, North-West University, Potchefstroom, South Africa and ³Department of Hydrobiology/CCBS, Federal University of São Carlos (UFSCar), São Carlos, São Paulo, Brazil

Abstract

Brazil has the richest biodiversity of Ergasilidae on Earth, with 76 species in 19 genera. However, several authors suggested that there is still great potential for discoveries, especially regarding genetic data that are still scarce for ergasilids from this region. To expand our knowledge of this taxon, we conducted an integrative study involving morphological (light and scanning electron microscopy) and molecular analyses of ergasilids from fishes sampled in the Pardo River, São Paulo State, Brazil. Two ergasilid species were found, *Gamispatulus schizodontis* and *Rhinergasilus piranhus* parasitizing the nostrils of 5 fish species (native and exotic): (i) *R. piranhus* from *Astyanax lacustris*; *Cyphocharax modestus*; *Coptodon rendalli*; *Psalidodon bockmanni*; and (ii) *G. schizodontis* from *Serrasalmus maculatus*. Additionally, we redescribed *R. piranhus* herein based on novelties and discrepancies found in the antennules, antennae, body segments and ornamentation of the swimming legs. Novel molecular data of ribosomal (18S and 28S rDNA) and/or mitochondrial (cox 1) genes were provided for both species and the phylogenetic relationships among the ergasilids were tested. Relationships between species/genera are still uncertain in Ergasilidae, but it was possible to verify the formation of 5 well-supported terminal clades – i.e. including a clade formed only by Neotropical species. More molecular data are needed to test this hypothesis, but the clades observed here represent good targets for future taxonomic revisions. The occurrence of *R. piranhus* specimens in the introduced fish, *Co. rendalli* can indicate (for the first time) host switching in this parasite species and spillback from native to introduced hosts.

Introduction

Copepoda is a diverse monophyletic taxon of the subphylum Crustacea; it comprises over 14 000 valid species distributed throughout the world (Bernot *et al.*, 2021). These small crustaceans inhabit every type of aquatic habitat (i.e. from deep-sea communities to semi-terrestrial conditions such as damp moss/leaf litter in humid forests) and exhibit a wide range of body shapes with distinct life cycles (i.e. from free-living to obligate parasites) (Boxshall and Halsey, 2004). Parasitism appeared independently at least 14 times in the evolutionary history of this taxon and today over 5000 species are recognized as parasites (Bernot *et al.*, 2021).

Fourteen metazoan phyla, ranging from sponges to large cetaceans, act as hosts for 1 or more symbiotic stages of copepods (see Tables 3.8 and 3.9 in Boxshall and Hayes, 2019). Fish (including Chondrichthyes and mainly Osteichthyes) stand out as one of the most studied host groups. In addition to a large number of known host–parasite associations with copepods (i.e. over 890 associations in the Neotropical region alone, see Luque *et al.*, 2013), this group also includes several species that impact the economy as pests in aquaculture (Johnson *et al.*, 2004; Piasecki *et al.*, 2004).

In Brazil, copepods represent one of the most speciose taxa of fish ectoparasites comprising over 200 operational taxonomic units [since it includes both determined and undetermined species; see Table 1 in Luque *et al.* (2013)]. In freshwater, the parasitic family Ergasilidae Burmeister, 1835 stands out as the most important due to its high diversity and harmful aquaculture potential (Piasecki *et al.*, 2004; Luque *et al.*, 2013). These copepods are easily recognized by their prehensile antenna, the absence of maxillipeds in females and the morphology of mouthparts (Boxshall and Halsey, 2004). Most adult females (i.e. the only parasitic stage of their life cycle) can be frequently found attached to the gills, but they also occur inside nostrils, on the surface of the body, embedded into host tissues as mesoparasites or even

© The Author(s), 2024. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives licence (<http://creativecommons.org/licenses/by-nc-nd/4.0>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided that no alterations are made and the original article is properly cited. The written permission of Cambridge University Press must be obtained prior to any commercial use and/or adaptation of the article.

as endoparasites inside the urinary bladder (Thatcher, 2006; Tang and Kalman, 2008; Rosim *et al.*, 2013).

Despite the importance of ergasilids in Brazil, several authors believe that the current biodiversity of these copepods is still underestimated (Thatcher, 1998; Boxshall and Defaye, 2008; Luque *et al.*, 2013). More than 90% of Brazilian fishes (>4000 spp.) have never been studied for their parasitic copepod fauna, leaving an enormous gap in the country for discoveries (Luque *et al.*, 2013). During the parasitological survey of fishes from the Pardo River, São Paulo State, Brazil, copepods were found parasitizing the nostrils of 4 native and 1 introduced fish species: *Astyanax lacustris* (Lütken, 1875); *Cyphocharax modestus* (Fernández-Yépez, 1948); *Psalidodon bockmanni* (Vari and Castro, 2007); *Serrasalmus maculatus* Kner, 1858 and the introduced *Coptodon rendalli* (Boulenger, 1897). The use of scanning and light microscopy together with molecular and phylogenetic analyses revealed that these copepods belong to 2 distinct species of Ergasilidae: *Rhinergasilus piranhus* Boeger and Thatcher, 1988 and *Gamispatulus schizodontis* Thatcher and Boeger, 1984. Molecular data were provided (for the first time) for both copepods (COI mtDNA, 18S and 28S rDNA genes) and the newly generated sequences were used to generate a new phylogenetic hypothesis for this family. Careful analysis of present specimens of *R. piranhus* revealed the need to redescribe this species, which is also done here.

Materials and methods

Host sampling and morphological analysis of copepods

From June 2020 to December 2022, 354 fish from 5 different species were sampled along the Pardo River in São Paulo state, Brazil

(Fig. 1). Fish were sampled with the aid of (1) multi-panel seine nets (3–14 cm mesh) soaked from 4 to 12 h, (2) cast nets, (3) fishing sieves and (4) fishing rods. Each fish was placed in a marked plastic bag and frozen for necropsy. In the laboratory, gills, surfaces, fins and nostrils of each fish were examined under a stereomicroscope for the presence of parasitic copepods. Fish sampling was authorized by the Instituto Chico Mendes de Conservação da Biodiversidade – ICMBio and Sistema de Autorização e Informação em Biodiversidade – SISBIO # 60640-1 and all procedures followed the recommendations of the Ethical Commission for Animal Experimentation from the São Paulo State University (Unesp), Institute of Biosciences, Botucatu, Brazil (CEUA no 9415260520). Fish were identified at the species level with the help of experts and the use of identification keys present in specialized literature (van der Sleen and Albert, 2017; Ota *et al.*, 2018; Ribeiro *et al.*, 2019). The scientific names of the fish cited throughout the text are in accordance with the nomenclature recommended in Eschmeyer's Catalogue of Fishes (Fricke *et al.*, 2024).

Copepods were carefully extracted from organs/tissues using fine needles and then preserved in glass vials with 70 and 96% ethanol for morphological and molecular analysis, respectively. For morphological identification, some copepods were cleared in lactic acid (10–30 min) and then mounted in Hoyer's medium. Whenever necessary, some specimens were dissected in glycerol medium and then each part was mounted on individual slides (e.g. antennules, antennae, swimming legs, etc.). Coverslips were sealed with transparent nail varnish. Morphological analysis and measurements of whole/dissected copepods were made using a compound microscope with differential interference contrast optics (Leica DMLB 5000, Leica Microsystems). Drawings were

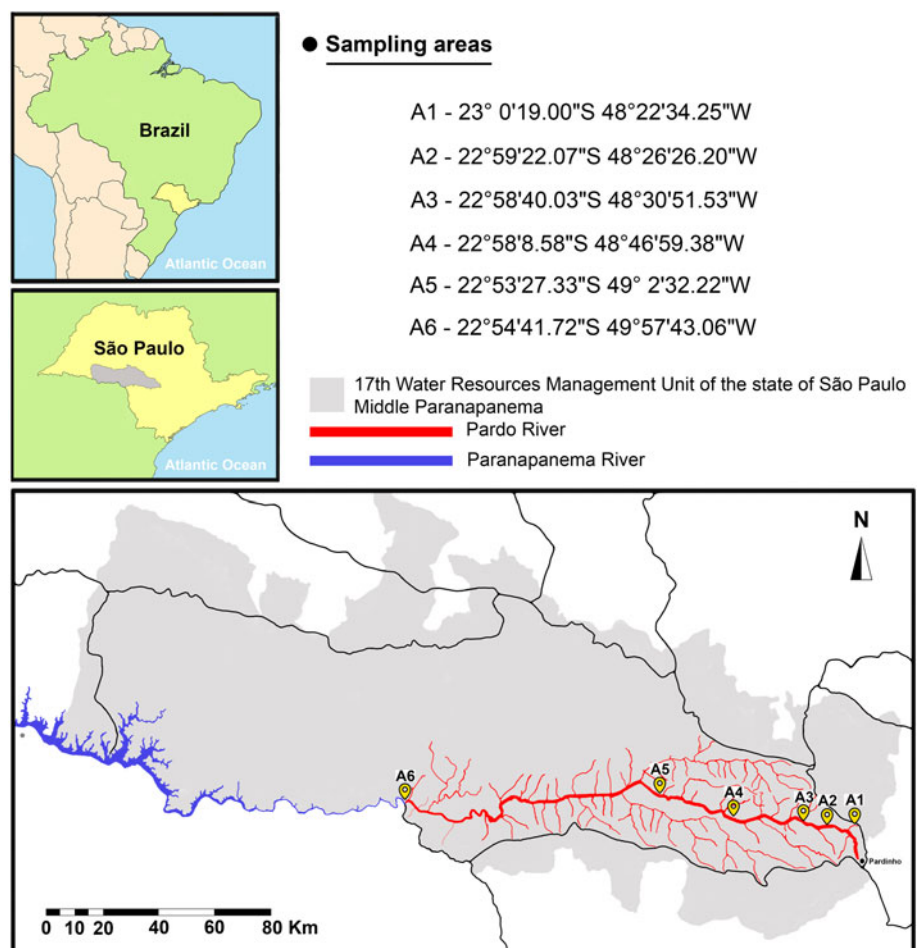


Figure 1. Sampling areas (A1 to A6) along the Pardo River, São Paulo state, Brazil.

Table 1. Abbreviations of body parts and segments used throughout the text to describe copepods

Abbreviation	Meaning
PS-1 (2–5)	First (second to fifth) pedigerous somite
AS-1 (2, 3)	First (second, third) abdominal somite
L1 (2–5)	First (second to fifth) leg
enp	Endopod
exp	Exopod
enp-1 (2, 3)	First (second, third) endopodal segment
exp-1 (2, 3)	First (second, third) exopodal segment

made with the aid of a compound microscope (LeicaDMLS, Leica Microsystems, Wetzlar, Germany) equipped with a drawing tube. All measurements are in micrometres (μm) and presented as the range followed by the mean in parenthesis. Morphological nomenclature used throughout the text to describe copepods was adapted from the nomenclature used by Boxshall and Halsey (2004) and Damborenea *et al.* (2020). Abbreviations used for describing the copepod parts (e.g. body regions, segments and appendices) are summarized in Table 1. Ecological descriptors such as prevalence and mean intensity were calculated following Bush *et al.* (1997).

For scanning electron microscopy (SEM), copepods were dehydrated in a graded ethanol series to 100% ethanol and placed in hexamethyldisilazane. Dried specimens were mounted on carbon tape on aluminium stubs and sputter-coated with carbon (Emscope TB500, Quorum Technologies, Puslinch, ON, USA), followed by 20–30 nm gold/palladium (Eiko IB2 ion coater, Eiko, Japan). Micrographs were taken using an FEI Nova NanoSEM 450 scanning electron microscope (FEI, Hillsboro, OR, USA).

Type-material of *R. piranhus* (Cyclopoida: Ergasilidae) was burrowed from the Zoological Collection of the National Institute of Amazonian Research (INPA) (holotype INPA PA 309-1; paratypes INPA PA 309-2 to 309-5) (see Supplementary Fig. 1) and used for morphological comparison with present specimens. Regarding *G. schizodontis*, the specimens were compared with the illustrations and/or information present in the available literature (Thatcher and Boeger, 1984; Lacerda *et al.*, 2007; Narciso and Silva, 2020). Representative specimens of each species were deposited in the Zoological Collection of the Museum of Zoology of the University of São Paulo (MZUSP), municipality of São Paulo, São Paulo state, Brazil.

DNA extraction and PCR amplification

DNA extraction was performed using the posterior part of the body (=urosome) and/or the egg sacs of the copepods, except for *R. piranhus*, for which whole specimens were used due to their small size (<400 μm). When only a small portion of the specimens or the egg sacs was used for DNA extraction, the remainder (i.e. the entire copepod or the upper part of the body) was deposited as a hologenophore in the Zoological Collection of MZUSP. For *R. piranhus*, micrographs (i.e. photohologenophore) of each specimen used for DNA extraction were taken (see Supplementary Fig. 2).

Genomic DNA was extracted from specimens fixed at 96% ethanol using the following kits: (1) NucleoSpin Tissue XS (Macherey-Nagel, Düren, Germany) Kit for whole/dissected copepods; and (2) PCR Biosystems Rapid DNA Extraction (London, England) Kit for egg sacs. Extractions followed the

recommendations indicated in the manufacturer's protocol for each kit. Partial fragments of a mitochondrial gene (subunit I of cytochrome c oxidase – *cox1*) and 2 ribosomal genes (18S and 28S rDNA) were amplified using a set of primers available in the literature (summarized in Table 2). Polymerase chain reaction (PCR) was performed in 25 μm mixtures containing 22 μL of DreamTaq PCR Master Mix (comprising Dream Taq DNA Polymerase, optimized DreamTaq buffer, MgCl_2 and dNTPs), 1.25 μL of each primer (forward and reverse) and 0.5 μL of genomic DNA under the following conditions: (i) 95°C for 5 min; (ii) 30 cycles of 95°C for 30 s; (iii) 47°C for 30 s for primer annealing; (iv) 72°C for 1 min; and (v) 72°C for 7 min for final extension (adapted from Song *et al.*, 2008). The amplified PCR products were run on agarose gel (1%) stained with GelRedTM (Biotium Inc., Fremont City, CA, USA) and visualized with a UV transilluminator. Successful amplifications were sent for cleaning, purification and sequencing to Inqaba Biotechnical Industries (Pty) Ltd. (Pretoria, South Africa). Forward and reverse sequences were assembled, aligned and edited using Geneious 11.1.4 (Biomatters, Auckland, New Zealand). Novel sequence data have been deposited in GenBank (see Table 3).

DNA sequence alignment and phylogenetic reconstruction

The newly generated sequences of each gene (*cox1*, 18S and 28S) were assembled and inspected using 'De Novo Assembly' tool in Geneious Prime v2022.2. (<https://www.geneious.com>). The resulting consensus sequences were compared to available sequences using the Basic Local Alignment Tool (BLAST) (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>, accessed on 2 January 2024) (Altschul *et al.*, 1990) to confirm the identity of the present specimens (if possible) and define their closest congeners. For all alignments, available sequences of the lernaed copepods were chosen as the outgroup: *Lernaea cyprinacea* Hermann, 1783; *Lamproglena clariae* Fryer, 1956; and/or *Lamproglena orientalis* Markevich, 1936. Two distinct sequences of copepods were taken from 8 families of the Suborder Ergasilida and used herein as an internal group due to their phylogenetic proximity to Ergasilidae [see the phylogenetic tree present in Bernot *et al.* (2021)]. For ergasilids, all available sequences (maximum of 3 per species) were taken from the nucleotide database of the National Center for Biotechnology Information (NCBI) (<https://www.ncbi.nlm.nih.gov/>, accessed on 2 January 2024). Sequences shorter than 450 bp were excluded from the analyses in order to avoid the loss of informative characters. All sequences used in the analyses (including ergasilids, internal groups and outgroups) are summarized in Table 3. The alignments were made in Geneious using the default parameters of MAFFT (Kato *et al.*, 2002). Extremes of each sequence were subsequently trimmed to an alignment of (i) 510 bp for *cox1*; (ii) 1121 bp for 18S; and (iii) 659 bp for 28S. Indels greater than 3 consecutive characters and affecting more than 5% of sequences were also removed from the final alignment. Only sequences of 18S and 28S were used for subsequent phylogenetic analyses. The best nucleotide substitution model test for maximum likelihood (ML) and Bayesian inference (BI) were determined using jModelTest v2.1.10 based on the Akaike information criterion (AIC) (Darriba *et al.*, 2012). The chosen AIC model was GTR + I + G for all genes. The ML and BI analyses were run in RAxML v7.2.8 (Stamatakis, 2014) and MrBayes v3.2.2 (Ronquist *et al.*, 2012), respectively, via the CIPRES Science Gateway (available at <https://www.phylo.org/>) (Miller *et al.*, 2010). Nodal supports for ML analyses were estimated at 100 bootstrap replications and only nodes with values greater than 75% were considered as well-supported. The BI analyses were run for 10 million generations (ngen = 10 000 000) with 2 runs each

Table 2. List of primers (forward and reverse) used for DNA amplification of the partial mitochondrial (cox1) and ribosomal genes (18S and 28S rDNA) of parasitic ergasilids (Cyclopoida: Ergasilidae) found in the nostrils of fishes from Pardo River, São Paulo state, Brazil – including sequences and references

Gene regions	Primers	Sequences	References
Cox1	LCO1490	5-GGTCACAAATCATAAGATATTGG-3'	Folmer <i>et al.</i> (1994)
	HCO2198	5-TAAACTTCAGGGTGACCAAAAAATCA-3'	
18S	18SF	5-AAGGTGTGMCCTATCAACT-3'	Song <i>et al.</i> (2008)
	18SR	5-TTA CTTCTCTAAACGCTC-3'	
28S	28SF	5-ACAACTGTGATGCCCTTAG-3'	Song <i>et al.</i> (2008)
	28SR	5-TGGTCCGTGTTTCAAGACG-3'	

containing 4 simultaneous Markov Chain Monte Carlo (MCMC) chains (nchains = 4) and with a sampling tree topology every 1000 generations (samplefreq = 1000). Burn-in was set to the first 25% of generations (Burnfrac = 0.25). Bayesian analyses used the following parameters: nst = 6, rates = invgamma, ngammacat = 4 and the priors' parameters of the combined dataset were set to ratepr = variable. Samples of substitution model parameters, and tree and branch lengths were summarized using the parameters 'sump burnin = 3000' and 'sumt burnin = 3000'. Only nodes with posterior probabilities (pp) greater than 75% were considered as well-supported. The BI and ML trees were edited and visualized in FigTree v1.4.3 software (Rambaut, 2012). Pairwise genetic distances (*p*-distances) were estimated in MEGA 7.0 (Kumar *et al.*, 2016), and the number of base pair differences was calculated in Geneious.

Results

Two copepod species were found parasitizing the nostrils of 5 fish species from Rio Pardo (Fig. 2; Tables 4 and 5): (i) *R. piranhus*; and (ii) *G. schizodontis*. *Rhinergasilus piranhus* was found only in the first 2 sampling areas (close to the river spring) associated with both native and exotic fish. *Gamispatulus schizodontis* specimens were found inside the nostrils of a single host species, the native piranha *S. maculatus* – only at the river mouth (or A6 in Fig. 1).

Morphological characterization

Rhinergasilus piranhus Boeger and Thatcher, 1988 (Figs 3–7; Tables 6 and 7)

Type host: *Pygocentrus nattereri* Kner, 1858 (Characiformes: Serrasalminidae).

Other hosts: *Acestrorhynchus lacustris* (Lütken, 1875) (Characiformes: Acestrorhynchidae); *Acestrorhynchus falciostris* (Cuvier, 1819) (Characiformes: Acestrorhynchidae); *A. lacustris* (Lütken, 1875) (Characiformes: Characidae) – present study (PS); *Colossoma macropomum* (Cuvier, 1816) (Characiformes: Serrasalminidae); *Co. rendalli* (Boulenger, 1897) (Cichliformes: Cichlidae) – PS; *C. modestus* (Fernández-Yépez, 1948) (Characiformes: Curimatidae) – PS; *Prochilodus nigricans* Spix et Agassiz, 1829 (Characiformes: Prochilodontidae); *P. bockmanni* (Vari and Castro, 2007) (Characiformes: Characidae) – PS; and *Serrasalmus altispinis* Mercky, Jégu et Santos, 2000 (Characiformes: Serrasalminidae).

Type locality: Ilha da Marchantaria, Solimoes-Amazonas River near Manaus, Amazonas state, Brazil.

Sampled localities: Area 1 – initial section (close to the river spring) of the Pardo River located within the 'Boa Vista' Farm in the municipality of Pardo, state of São Paulo, Brazil (22° 59'59.8"S, 48°22'37.4"W); and area 2 – section of the Rio Pardo

located within the 'Paineiras' Farm in the municipality of Pardo, state of São Paulo, Brazil (22°59'22.1"S, 48°26'26.2"W).

Site of infestation: Nostrils.

Material examined: A total of 20 ♀♀ were examined for morphological description, all in nostrils, including (i) 7 ♀♀ from *A. lacustris* sampled in area 1 (4 ♀♀) and area 2 (3 ♀♀); (ii) 2 ♀♀ from *Co. rendalli* in area 2; (iii) 2 ♀♀ from *Cy. modestus* in area 2; and (iv) 9 ♀♀ from *P. bockmanni* in area 1 (2 ♀♀) and area 2 (7 ♀♀). Body and appendage measurements are listed in Table 6.

Additional material: 5 ♀♀ of the type-material of *R. piranhus* were borrowed from INPA's zoological collection and then used for morphological comparisons, including the holotype INPA PA 309-1 and 4 paratypes (INPA PA 309-2 to 309-5).

Representative DNA sequences: PQ442939, PQ452578, PQ452580-82.

Voucher material: A total of 20 whole ♀♀ (including 10 ♀♀ mounted in Hoyer's medium and 10 ♀♀ stored in glass vials with 70% ethanol) were deposited in the Zoological Collection of MZUSP (accession no. 46666-69, 46690, 46736, 46776, 46778).

Redescription (based on adult females; male unknown): Body cycloform (Figs 3A and 5A), comprising prosome, urosome and caudal rami. Prosome consisting of cephalosome and PS-1; both segments separated dorsally by flexible cuticle; and 3 free pedigerous somites (PS-2 to PS-4). Cephalosome (Figs 3A and 5D) widening posteriorly, maximum width at level of buccal apparatus, anterior margin protruded, dorsal surface with 3 elliptical integumental windows (anterior, medial and posterior); posterior window with diameter about 2.0× times larger than previous window, with slightly rough surface (Figs 5E and 7H); armed with paired spine-like projections (Figs 5F and 7G), and ornamented with several pores symmetrically distributed throughout segment; pores with or without bristles. PS-1 (Figs 3A and 7C) with rounded lateral margins, ornamented with pores along posterior margin; pores lacking bristles. Rostrum (Figs 3B and 5E) with rounded posterior margin, ventrally ornamented with pair of pores. Free pedigerous somites (Figs 3A and 7B) decreasing gradually in width (on transverse axis) from anterior to posterior; each somite ornamented with 1 or more pairs of pores (Figs 3A and 7C); pores with or without bristles; PS-2 (Figs 3A and 7C) narrower than previous somite, with paired integumental windows laterally on tergite (arrowhead on Fig. 7C); PS-3 and PS-4, both lacking integumental windows; PS-4 reduced, smaller and thinner than previous prosome somites.

Urosome consisting of PS-5, genital double-somite and 3 free abdominal somites (AS-1 to AS-3). PS-5 (Figs 4A and 7C) reduced, similar in size to PS-4, unornamented. Genital double-somite (Fig. 4A) narrowing posteriorly, with rounded lateral margins, ornamented ventrally with posterior row of spinules. Abdominal somites (Figs 4A, 6B and 7E) decreasing gradually in width (on transverse axis) from anterior to posterior; each somite ornamented ventrally with posterior row of spinules; AS-3 (=anal somite) deeply incised posteriorly (=anal opening

Table 3. List of sequences from the partial mitochondrial (subunit I of cytochrome c oxidase or *cox1*) and ribosomal genes (18S and 28S rDNA) included in the phylogenetic analyses

Copepoda	Host species	Locality	Cox	18S	28S	Reference
Outgroup						
Lernaeidae						
<i>Lamproglena clariae</i> Fryer, 1956	<i>Clarias gariepinus</i> (Burchell, 1822)	Kenya	OR232208 OR232209	–	–	Rindoria et al. (2023)
<i>Lamproglena cleopatra</i> Humes, 1957	<i>Labeo victorianus</i> Boulenger, 1901	Kenya	OR232207	–	–	Rindoria et al. (2023)
<i>Lamproglena orientalis</i> Markevich, 1936	<i>Squaliobarbus curriculus</i> (Richardson, 1846) <i>Chanodichthys erythropterus</i> (Basilevsky, 1855)	China	–	DQ107552	DQ107541	Song et al. (2008)
<i>Lernaea cyprinacea</i> Linnaeus, 1758	<i>Hemiculter leucisculus</i> (Basilevsky, 1855) <i>Opsariichthys bidens</i> Günther, 1873	China	–	DQ107554	DQ107548	Song et al. (2008)
Ingroup						
Chondracanthidae						
<i>Acanthochondria spirigera</i> Shiino, 1955	–	South Korea	KR049020	KR048753	KR048824	Unpublished
<i>Chondracanthus distortus</i> Wilson, 1922	–	South Korea	KR049023	KR048756	KR048827	Unpublished
Clausidiidae						
<i>Conchylurus quintus</i> Tanaka, 1961	–	South Korea China	OQ725275	KR048763	KR048839	Unpublished
<i>Hemicyclops tanakai</i> Itoh and Nishida, 2002	–	South Korea	KR049027	KR048769	KR048854	Unpublished
Lichomolgidae						
<i>Herrmannella hoonsooi</i> Kim, 1992	–	South Korea	KR049039	–	KR048849	Unpublished
<i>Herrmannella longicaudata</i> Avdeev, 1975	–	South Korea	–	KR048757	–	Unpublished
<i>Lichomolgus nakaii</i> Matsuzaki and Ogawa, 1989	–	South Korea	–	–	KR048845	Unpublished
<i>Lichomolgus similis</i> Ho and Kim, 1991	–	South Korea China	OQ725276	KR048758	–	Unpublished
Mycolidae						
<i>Ostrincola koe</i> Tanaka, 1961	–	South Korea	OQ725292	KR048750	KR048821	Unpublished
<i>Pseudomyicola spinosus</i> (Raffaele and Monticelli, 1885)	–	South Korea China	OQ725301	KR048751	KR048822	Unpublished
Mytilicolidae						
<i>Mytilicola orientalis</i> Mori, 1935	<i>Magallana gigas</i> (Thunberg, 1793)	Germany China	OQ725290	HM775190	–	Elsner et al. (2011)
<i>Trochicola enterica</i> Dollfus, 1914	<i>Calliostoma zizyphinum</i> (Linnaeus, 1758)	France	–	AY627006	–	Huys et al. (2006)
<i>Trochicola japonica</i> Shimura and Kuwabara, 1984	–	China	OQ725305	–	–	Unpublished
Rhynchomolgidae						
<i>Critomolgus vicinus</i> Kim, 1996	–	South Korea	–	KR048766	KR048848	Unpublished
<i>Zamolgus cavernularius</i> Kim, 2000	–	South Korea	–	KR048761	KR048832	Unpublished

(Continued)

Table 3. (Continued.)

Copepoda	Host species	Locality	Cox	18S	28S	Reference
Schminkepinellidae						
<i>Cyclopinella</i> sp.	–	Pacific Ocean	MK370282	MK370225	–	Khodami <i>et al.</i> (2019)
<i>Schminkepinella</i> sp.	–	Pacific Ocean	MK370288	MK370222	MK370276	Khodami <i>et al.</i> (2019)
Taeniacanthidae						
<i>Clavisodalis dilatatus</i> Dojiri and Humes, 1982	–	Japan	–	–	LC752235	Unpublished
<i>Clavisodalis sentifer</i> Dojiri and Humes, 1982	–	Japan	–	LC752238	LC752237	Unpublished
<i>Pseudotaeniacanthus congeri</i> Yamaguti and Yamasu, 1959	–	South Korea	KR049030	–	–	Unpublished
<i>Taeniacanthus yamagutii</i> (Shiino, 1957)	–	South Korea	KR049032	KR048748	–	Unpublished
Ergasilidae						
<i>Acusicola margulisiae</i> Santacruz <i>et al.</i> , 2020	<i>Amphilophus citrinellus</i> (Günther, 1864) <i>Parachromis managuensis</i> (Günther, 1867) <i>Oreochromis</i> sp. <i>Poecilia mexicana</i> Steindachner, 1863	Nicaragua	MN854869 MN854870	MN852696	MN852849 MN852851	Santacruz <i>et al.</i> (2020)
<i>Acusicola</i> sp.	–	Iraq	LC599903	–	–	Unpublished
<i>Ergasilus anchoratus</i> Markevich, 1946	<i>Tachysurus fulvidraco</i> (Richardson, 1846)	China	–	DQ107564 MZ558064 OP076959	DQ107528	Song <i>et al.</i> (2008)
<i>Ergasilus briani</i> Markevich, 1933	<i>Misgurnus anguillicaudatus</i> (Cantor, 1842)	China	–	DQ107572	DQ107532	Song <i>et al.</i> (2008)
<i>Ergasilus caparti</i> Mič <i>et al.</i> , 2023	<i>Neolamprologus brichardi</i> (Poll, 1974)	Burundi	–	OQ407469	OQ407474	Mič <i>et al.</i> (2023)
<i>Ergasilus caeruleus</i> Wilson, 1911	–	USA	OP830263 OP830265	–	–	Unpublished
<i>Ergasilus centrarchidarum</i> Wright, 1882	–	USA	OP830032 OP830192 OP830132	–	–	Unpublished
<i>Ergasilus centrarchidarum</i> Wright, 1882	–	USA	OP830234 OP830262	–	–	Unpublished
<i>Ergasilus danjiangensis</i> Song <i>et al.</i> , 2008	<i>Zacco platypus</i> (Temminck and Schlegel, 1846)	China	–	–	DQ107533	Song <i>et al.</i> (2008)
<i>Ergasilus hypomesi</i> Yamaguti, 1936	<i>Acanthogobius hasta</i> (Temminck and Schlegel, 1845)	China	–	DQ107573	DQ107539	Song <i>et al.</i> (2008)
<i>Ergasilus jaraquensis</i> Thatcher and Robertson, 1982	–	Brazil	MF651988 MF651989	–	–	Unpublished
<i>Ergasilus luciopercarum</i> Henderson, 1926	–	USA	OP830298 OP830347	–	–	Unpublished
<i>Ergasilus macrodactylus</i> (Sars, 1909)	<i>Gnathochromis permaxillaris</i> (David, 1936)	Burundi	–	OQ407465	OQ407470	Mič <i>et al.</i> (2023)
<i>Ergasilus megaceros</i> Wilson, 1916	–	USA	OP830321 OP830295	–	–	Unpublished

<i>Ergasilus megacheir</i> (Sars, 1909)	<i>Simochromis diagramma</i> (Günther, 1894)	Burundi	–	OQ407466	OQ407471	Mič <i>et al.</i> (2023)
<i>Ergasilus mirabilis</i> Oldewage and van As, 1987	<i>Clarias gariepinus</i> (Burchell, 1822)	South Africa Zambia	OR448769 OR448770	OR449753	OR449756	Fikiye <i>et al.</i> (2023)
<i>Ergasilus parasarsi</i> Mič <i>et al.</i> , 2023	<i>Simochromis diagramma</i> (Günther, 1894)	Burundi	–	OQ407467	OQ407473	Mič <i>et al.</i> (2023)
<i>Ergasilus parasiluri</i> (Yamaguti, 1936)	<i>Silurus asotus</i> Linnaeus, 1758 <i>Tachysurus fulvidraco</i> (Richardson, 1846)	China	–	DQ107567	DQ107536	Song <i>et al.</i> (2008)
<i>Ergasilus parvitergum</i> Ho <i>et al.</i> , 1992	–	USA	OP871074	–	–	Unpublished
<i>Ergasilus parvus</i> Mič <i>et al.</i> , 2023	<i>Spathodus erythrodon</i> Boulenger, 1900	Burundi	–	OQ407468	OQ407472	Mič <i>et al.</i> (2023)
<i>Ergasilus peregrinus</i> Heller, 1865	<i>Siniperca chuatsi</i> (Basilewsky, 1855)	China	–	DQ107577	DQ107531	Song <i>et al.</i> (2008)
<i>Ergasilus scalaris</i> Markevich, 1940	<i>Tachysurus dumerili</i> (Bleeker, 1864)	China	–	DQ107565	DQ107538	Song <i>et al.</i> (2008)
<i>Ergasilus sieboldi</i> Nordmann, 1832	<i>Perca fluviatilis</i> Linnaeus, 1758 <i>Sparus aurata</i> Linnaeus, 1758	Czech Republic Egypt	–	MW810238	OM812074 ON706996	Kvach <i>et al.</i> (2021) Abou-Okada <i>et al.</i> (2023)
<i>Ergasilus tumidus</i> Markevich, 1940	<i>Acheilognathus taenianalis</i> (Günther, 1873)	China	–	DQ107569	DQ107535	Song <i>et al.</i> (2008)
<i>Ergasilus versicolor</i> Wilson, 1911	–	USA	OP830223 OP830335	–	–	Unpublished
<i>Ergasilus wilsoni</i> Markevich, 1933	–	South Korea	KR049036	KR048765	KR048843	Unpublished
<i>Ergasilus yaluzangbus</i> Kuang and Qian, 1985	<i>Oxygymnocypris stewartii</i> (Lloyd, 1908)	China	–	DQ107578	DQ107540	Song <i>et al.</i> (2008)
<i>Ergasilus yandemontei</i> Waicheim <i>et al.</i> , 2021	<i>Odontesthes hatcheri</i> (Eigenmann, 1909)	Argentina	–	MT969345	–	Waicheim <i>et al.</i> (2021)
<i>Ergasilus</i> sp.	<i>Mugil liza</i> Valenciennes, 1836	South Korea Chile	KR049035 KU557411 KU557412	–	KR048842	Castro-Romero <i>et al.</i> (2016)
Ergasilidae gen. sp.	–	Mexico Nicaragua	MG448691 MG448706 OM541914	–	–	Elías-Gutiérrez <i>et al.</i> (2018) Santacruz <i>et al.</i> (2022)
<i>Gamispatus schizodontis</i> Thatcher and Boeger, 1984	<i>Serrasalmus maculatus</i> Kner, 1858	Brazil	PQ442941 PQ442938 PQ442940 PQ450081	PQ452576 PQ452577	PQ452579	Present study
<i>Gamispinus diabolicus</i> Thatcher and Boeger, 1984	–	Brazil	MF651982 MF651983	–	–	Unpublished
<i>Miracetyma</i> sp.	–	Brazil	MF651987 MF651985	–	–	Unpublished
<i>Neoergasilus japonicus</i> (Harada, 1930)	<i>Lepomis gibbosus</i> (Linnaeus, 1758) <i>Scardinius erythrophthalmus</i> (Linnaeus, 1758)	South Korea Czech Republic USA	MZ964938 OP830068	KR048752 MH167970	KR048823 MW810241	Ondračková <i>et al.</i> (2019) Kvach <i>et al.</i> (2021) Vasquez <i>et al.</i> (2022)
<i>Neoergasilus</i> sp.	–	USA	MZ964932 MZ964933	–	–	Vasquez <i>et al.</i> (2022)
<i>Paeonodes subviridis</i> El-Rashidy and Boxshall, 2001	–	India	OP425700	–	–	Unpublished

(Continued)

Table 3. (Continued.)

Copepoda	Host species	Locality	Cox	18S	28S	Reference
<i>Paraergasilus brevidigitus</i> Yin, 1954	<i>Cyprinus carpio</i> Linnaeus, 1758	China	-	DQ107576	DQ107530_	Song et al. (2008)
<i>Paraergasilus longidigitus</i> Yin, 1954	<i>Abramis brama</i> (Linnaeus, 1758) <i>Perca fluviatilis</i> Linnaeus, 1758 <i>Scardinius erythrophthalmus</i> (Linnaeus, 1758)	Czech Republic	-	MW810239	-	Kvach et al. (2021)
<i>Paraergasilus medius</i> Yin, 1956	<i>Ctenopharyngodon idella</i> (Valenciennes, 1844)	China	-	DQ107574	DQ107529	Song et al. (2008)
<i>Rhinergasilus piranhus</i> Boeger and Thatcher, 1988	<i>Astyanax lacustris</i> (Lütken, 1875) <i>Psalidodon bockmanni</i> (Vari and Castro, 2007)	Brazil	PQ442939	-	PQ452578 PQ452580 PQ452581 PQ452582	Present study
<i>Sinergasilus major</i> (Markevich, 1940)	<i>Ctenopharyngodon idella</i> (Valenciennes, 1844) <i>Silurus glanis</i> Linnaeus, 1758	China Hungary	-	DQ107558 MZ047814	DQ107524 MZ047815	Song et al. (2008) Dos Santos et al. (2021)
<i>Sinergasilus polycolpus</i> (Markevich, 1940)	<i>Hypophthalmichthys molitrix</i> (Valenciennes, 1844)	China	-	DQ107563	DQ107525	Song et al. (2008)
<i>Sinergasilus undulatus</i> (Markevich, 1940)	<i>Cyprinus carpio</i> Linnaeus, 1758	China	-	DQ107561 MZ575163	DQ107527	Song et al. (2008)
<i>Therodamas longicollum</i> Oliveira et al., 2021	<i>Leporinus fasciatus</i> (Bloch, 1794)	Brazil	-	MW652731	-	Oliveira et al. (2021)

Taxa represented in bold fonts are the novel sequences provided in the present study.

or anus), dorsal surface with a pair of pores carrying bristles located laterally to anus (highlighted in Fig. 7E).

Caudal rami (Fig. 4A), each ramus ornamented with 2 rows of spinules; spinule rows located immediately before seta 1 and 3; armed with 4 bare setae (s-1 to s-4); s-1 and s-3 similar in size, both setae inserted on ventral surface; s-2 and s-4 inserted on posterior margin; s-4 longest.

Antennule 5-segmented (Figs 3F and 5B); proximal segment about 2.0 × longer than others, incompletely subdivided into 2 subsegments; setal formula: 6, 4, 4, 2, 5 + 2 ae (total 23). Antenna (Figs 3E and 5C) 4-segmented comprising coxobasis, and 3-segmented endopod (enp-1 to enp-3). Coxobasis broad, armed with short outer seta. Enp-1 slightly longer than coxobasis, unornamented. Enp-2 triangular, maximum width at level of proximal margin, unornamented. Enp-3 reduced, unornamented. Antennal claw strongly curved, similar in size to enp-1 with fossa on concave margin; tip rounded and covered by cap. Antennal support (Figs 3C and 5D) ventrally articulated with antenna by coxobasis, armed with lateral spine-like projection.

Buccal apparatus (Fig. 3D) comprising mandible, maxillule and maxilla. Labrum not observed. Mandible armed with 2 blades (=anterior and posterior blade), both blades ornamented with spinules along posterior margin. Maxillule reduced, unornamented. Maxilla 2-segmented, comprising syncoxa (=first segment) and basis (=second segment); syncoxa broad, with distal pore; basis ornamented with multiple spinules.

Three pairs of biramous swimming legs (L1 to L3), each leg comprising coxa, basis, enp (=inner ramus) and exp (=outer ramus). L1 (Fig. 4B). Coxa ornamented with 2 clusters of spinules (lateral and central); central spinules arranged in horizontal line ($n = 3$ spinules) near posterior margin; lateral spinules minute, located on posterolateral margin ($n = 3-4$ spinules). Basis carrying bare outer seta and ornamented with row of spinules located immediately above inner ramus ($n = \sim 9$ spinules); spinules similar in size and shape as central spinules of coxobasis. Enp 2-segmented, all segments with spinules along outer margin; enp-1 rectangular, armed with 1 plumose seta, ornamented with bristles along outer margin; enp-2 oar-shaped, with 3 plumose setae. Exp 3-segmented, all segments with spinules along outer margin; enp-1 armed with serrated spine, ornamented with bristles along inner margin; enp-2 with plumose seta; enp-3 armed with 2 serrated spines, 1 semi-plumose seta (=serrated outer margin) and 4 plumose setae.

L2 (Fig. 4C). Coxa ornamented with minute spinules on outer margin. Basis with bare outer seta. Enp 3-segmented; enp-1 ornamented with bristles on outer margin, armed with plumose seta; enp-2 and -3 both ornamented with spinules along outer margin; enp-2 with 2 plumose setae; enp-3 with 1 spine and 4 plumose setae. Exp 3-segmented, all segments with spinules along outer margin; exp-1 armed with serrated spine, ornamented with bristles along inner margin; exp-2 with 1 plumose seta; exp-3 with 1 serrated spine, 1 semi-plumose seta and 5 plumose setae. L3 (Fig. 4D) with same ornamentation and armament described for L2, except for single plumose seta on enp-2 and lacking spine on enp-3.

L4 and L5, both legs reduced and represented by single seta (Figs 4A and 6C). Spine and setal formula of biramous swimming legs are summarized in Table 7.

Intercostal sclerites unornamented, with both lateral ends directed posteriorly (Figs 4E and 6A). Interpodal plate of L1 and L2, both ornamented with transverse row of spinules; interpodal plate of L3, unornamented. L4 and L5, absent. Egg sac (Fig. 4F) paired, multiseriate.

Remarks

The specimens studied were identified as members of the genus *Rhinergasilus* by having the following combination of diagnostic

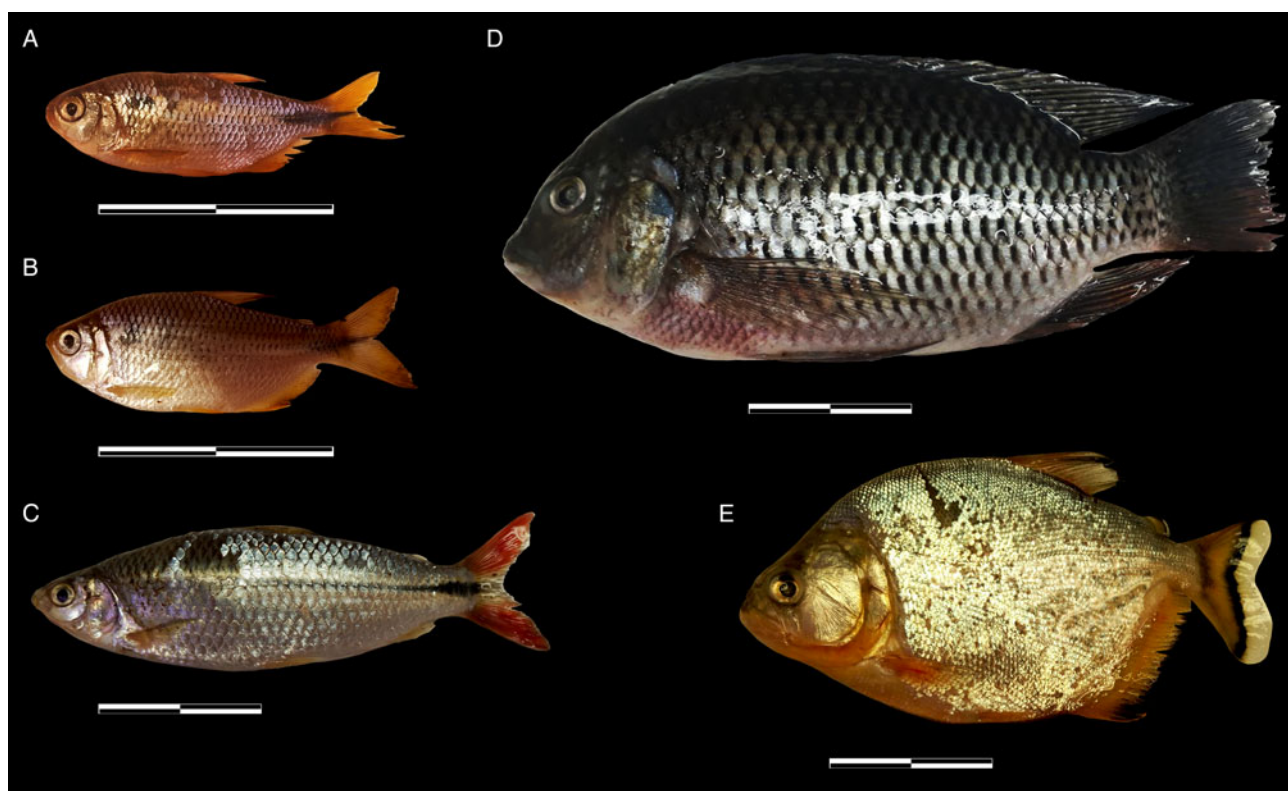


Figure 2. Fish collected and analysed in this study, (A) *Astyanax lacustris* (Lütken, 1875), (B) *Psalidodon bockmanni* (Vari and Castro, 2007), (C) *Cyphocharax modestus* (Fernández-Yépez, 1948), (D) *Coptodon rendalli* (Boulenger, 1897), (E) *Serrasalmus maculatus* Kner, 1858. Scale bars = 5 cm.

Table 4. Data from fish sampled along the Pardo River, São Paulo state, Brazil

Fish species	Fish family	L (cm)	W (g)	SpL	PC	Copepod species	SI	HS
<i>Astyanax lacustris</i> (Lütken, 1875)	Characidae	4–16 (10)	1–70 (21)	A1–28	P	<i>Rhinergasilus piranhus</i> Boeger and Thatcher, 1988	Nostrils	Native
				A2–11	NO	–	–	
				A4–24	NO	–	–	
				A6–30	NO	–	–	
<i>Psalidodon bockmanni</i> (Vari and Castro, 2007)	Characidae	2–16 (8)	0.5–50 (9.5)	A1–25	P	<i>Rhinergasilus piranhus</i>	Nostrils	Native
				A2–50	P	<i>Rhinergasilus piranhus</i>	Nostrils	
				A3–23	NO	–	–	
				A4–5	NO	–	–	
				A5–39	NO	–	–	
<i>Cyphocharax modestus</i> (Fernández-Yépez, 1948)	Curimatidae	8–18 (13)	8–93 (40)	A1–20	P	<i>Rhinergasilus piranhus</i>	Nostrils	Native
				A2–30	NO	–	–	
				A5–4	NO	–	–	
<i>Coptodon rendalli</i> (Boulenger, 1897)	Cichlidae	3–26 (11)	1–314 (32)	A1–34	P	<i>Rhinergasilus piranhus</i>	Nostrils	Introduced
				A4–17	NO	–	–	
				A5–3	NO	–	–	
<i>Serrasalmus maculatus</i> Kner, 1858	Serrasalmidae	10–27 (17)	19–635 (170)	A6–11	P	<i>Gamispatalus schizodontis</i> Thatcher and Boeger, 1984	Nostrils	Native

HS, host status on this river; L, total length in centimetres represented by the amplitude and the average in parentheses (cm); NO, not observed; P, present; SpL, fish specimens per locality; W, weight in grams represented by the amplitude and the average in parentheses (g). A1 to A6 = sampling areas along the river.

Table 5. Parasitological descriptors of ergasilids found in nostrils of fish from the Rio Pardo, São Paulo, Brazil

Host	N	Ergasilidae	P%	MI ± s.e. (R)	MA ± s.e. (R)
<i>Astyanax lacustris</i> (Lütken, 1875)	93	<i>Rhinergasilus piranhus</i> Boeger and Thatcher, 1988	9.7	8.1 ± 2.9 (1–22)	0.7 ± 0.3 (0–22)
			4.7	1.25 ± 0.25 (1–2)	0.06 ± 0.03 (0–2)
<i>Cyphocharax modestus</i> (Fernández-Yépez, 1948)	54	<i>Rhinergasilus piranhus</i>	1.8	2 (1–2)	0.04 ± 0.04 (0–2)
<i>Coptodon rendalli</i> (Boulenger, 1897)	54	<i>Rhinergasilus piranhus</i>	3.7	2 (1–2)	0.07 ± 0.05 (0–2)
<i>Psalidodon bockmanni</i> (Vari and Castro, 2007)	142	<i>Rhinergasilus piranhus</i>	10	5.8 ± 1.8 (1–25)	0.6 ± 0.2 (0–25)
			3.5	3.6 ± 1.6 (1–9)	0.1 ± 0.07 (0–9)
<i>Serrasalmus maculatus</i> Kner, 1858	11	<i>Gamispatulus schizodontis</i> Thatcher and Boeger, 1984	25	1–1	0.25 ± 0.1 (0–1)

MA, average abundance of infestation; MI, mean intensity of infestation; N, number of fish analysed; P%, prevalence in percentage; R, range of parasites; s.e., standard error. Calculations were made following the recommendations of Bush et al. (1997).

features: (1) PS-4 and PS-5 reduced, both somites narrower than previous ones; (2) 3 pairs of biramous swimming legs (L1 to L3); and (3) L4 and L5 reduced, each leg being represented by single seta (Boeger and Thatcher, 1988). Except for 4 genera (*Abergasilus* Hewitt, 1978; *Brasergasilus* Thatcher and Boeger, 1983; *Mugilicola* Tripathi, 1960; and *Urogasilus* Rosim et al., 2013), the reduction in the number of swimming legs allows the easy differentiation of these copepods from other ergasilids. The genera *Abergasilus* and *Mugilicola* do not occur in the Neotropical region [see Figs 4 and 5 in Amado et al. (1995)] and both differ from *Rhinergasilus* by having the first endopod with 3 segments instead of 2. *Rhinergasilus* can also be distinguished from *Mugilicola* and *Abergasilus* by the absence of a long 'neck' separating the cephalothorax in 2 distinguished areas (i.e. post-oral and pre-oral regions) or a robust spine on the second antennal segment, respectively. Regarding the Neotropical genera, it can be differentiated from *Urogasilus* by the preferred attachment site (i.e. nostrils/gills instead of the urinary bladder as in *Urogasilus*), the shape of the maxilla (i.e. maxilla carrying 2 blades in *Rhinergasilus* instead of a single blade as in *Urogasilus*) and the absence of a posterior 'trunk'. In *Urogasilus brasiliensis* Rosim et al., 2013, the final thoracic segments (PS-4 and PS-5) and genital somite are fused to form an elongated structure called 'trunk' (Rosim et al., 2013), which does not occur in any of the 3 known species of *Rhinergasilus* – i.e. even though the reduction in the size of PS-4 and PS-5 occurs in *Rhinergasilus*, these segments remain unfused in all species as can be seen in Fig. 7E. *Brasergasilus* is the Neotropical genus that most resembles *Rhinergasilus* by the general shape of the body, number/segmentation of legs, and attachment site. Despite that, these 2 genera can be differentiated from each other by the shape of the antennal claw and the presence/absence of the L4. In *Rhinergasilus*, the claw is relatively short and has a simple (sharp) tip rather than longer (i.e. usually longer than other antennal segments combined) and with a modified tip as present in *Brasergasilus* species. Regarding L4, in *Rhinergasilus* this leg is represented by a single seta, while in *Brasergasilus* it is completely absent.

Among the 3 known *Rhinergasilus* species (*Rhinergasilus digitus* Narciso et al., 2020; *R. piranhus* – type-species; and *Rhinergasilus unguilongus* Narciso et al., 2021), the present specimens were identified as *R. piranhus* since they possess the same leg structure as described by Boeger and Thatcher (1988). In *R. piranhus*, the L4 and L5 are both represented by a small papilla carrying a single seta and each leg arises independently from their respective somites [see Fig. 10 in Boeger and Thatcher (1988) and Figs 4A and 6C in

the present paper]. On the other hand, in the other 2 species, the papillae are apparently fused and both setae arise from the same somite (PS-5). Additionally, the present specimens also resemble *R. piranhus* regarding the size [287–347 (309) long in present specimens vs 237–282 (263) in the original description] and by sharing other diagnostic features with the type-species such as (1) seta-1 and -3 of caudal ramus long – each seta about $\frac{3}{4}$ the length of seta-2; (2) interpodal plates 1 and 2, coxa of L1, and base of L1 ornamented with a transverse row of spinules; (3) antenna with first endopodal segment (or enp-1) unornamented; and (4) claw being as long as enp-1 (i.e. latter segment considered the longest antennal segment).

The diagnostic features allowed the identification of the present copepods as *R. piranhus*; however, careful analysis of them also showed some discrepancies regarding the original description as well as the visualization of new features. Regarding the discrepancies, our specimens showed the PS-1 separated dorsally from cephalosome rather than both segments being completely fused as proposed by Boeger and Thatcher (1988). The setation of antennules was different, in present specimens the antennule setation is 6, 4, 4, 2, 5 + 2 ae (total 23) vs 4, 1, 4, 4, 5 (total 18) as in the original description. The setation of legs also differs, in present specimens the L1 enp-3 and L3 exp-3 possess 3 and 6 plumose setae, respectively, instead of 2 and 5 as in the original description. Finally, the egg disposal varied from uni- to multi-serial instead of just uniserial as in the original description.

Regarding the novelties, we observed new features associated with the thoracic/abdominal somites, antennae, maxilla and swimming legs. The dorsal cephalic shield is equipped with a pair of horn-like structures located immediately below the antennules (see Figs 5F and 7G). Ventrally, the antennal joint (also called antennal support here, as this structure seems to articulate the cephalosome with the antennal coxobase) carried lateral spine-like projections (see Figs 3C and 5D). Several pores (carrying bristles or without bristles) were also observed on the dorsal surface of the prosome and urosome somites – with a higher concentration of pores observed on the dorsal surface of the cephalosome (Fig. 3A). In addition to the pores, 3 elliptical windows were also observed for the first time in the cephalosome (as represented in Figs 5E and 7H). The maxilla showed spinules along the posterior margin of both blades (anterior and posterior blades) rather than only on the anterior blade as in the original description. Finally, careful analysis of the antennal claw showed the presence of fossa and a hyaline cap covering the tip. Other novelties such as the presence of serrated spines and semi-plumose setae were observed for all swimming legs.

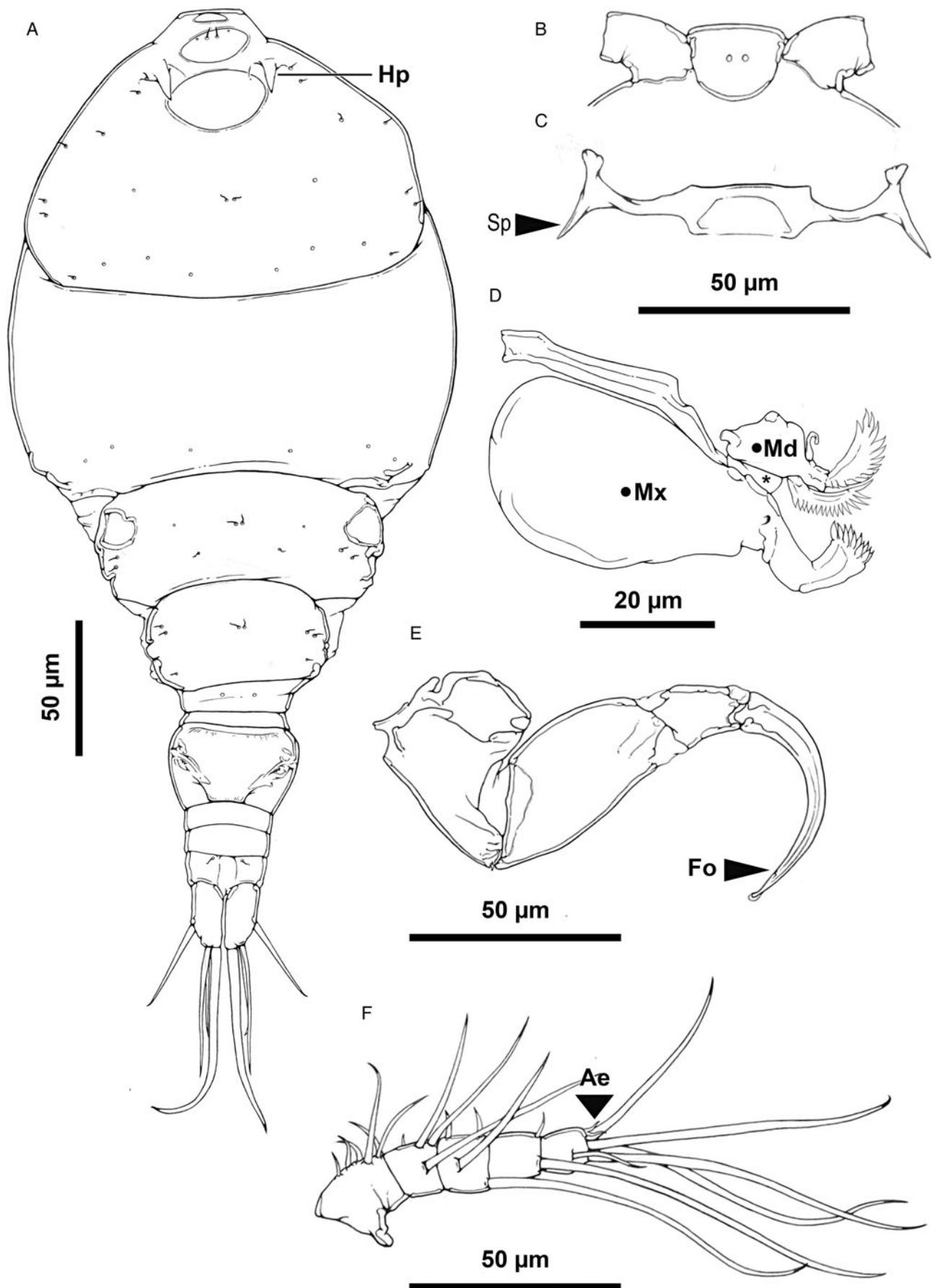


Figure 3. *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female, (A) body, dorsal view, cephalosome with a pair of horn-like projections (Hp), (B) rostrum, ventral view, (C) antennal support armed with lateral spine-like projections (Sp), (D) buccal apparatus comprising maxilla (Mx), maxillule (*) and mandible (Md), (E) antenna, claw with fossa (Fo) on concave margin, (F) antennule, distal segment 2 aesthetasc (Ae). Scale bars in micrometres (μm).

Table 6. Measurements in micrometres (μm) of adult females of *Rhinergasilus piranhus* Boeger and Thatcher, 1988

Measured part	Range (mean)
Total length ^a	287–347 (309)
Cephalosome length	101–142 (109)
Cephalosome width	113–145 (131)
Pedigerous somite 1 length	41–51 (46)
Pedigerous somite 1 width	119–145 (139)
Antennule length	62–72 (67)
Antenna segment 1 length	40–52 (45)
Antenna segment 2 length	48–54 (52)
Antenna segment 3 length	19–25 (22)
Antenna segment 4 length	5–6 (6)
Claw length	42–52 (48)
Pedigerous somite 2 length	22–37 (28)
Pedigerous somite 2 width	93–109 (101)
Pedigerous somite 3 length	27–41 (32)
Pedigerous somite 3 width	61–69 (64)
Pedigerous somite 4 length	8–12 (11)
Pedigerous somite 4 width	34–42 (38)
Pedigerous somite 5 length	5–9 (7)
Pedigerous somite 5 width	31–42 (39)
Genital double-somite length	27–35 (27)
Genital double-somite width	39–48 (46)
Abdominal somite 1 length	8–12 (11)
Abdominal somite 1 width	25–35 (32)
Abdominal somite 2 length	9–14 (12)
Abdominal somite 2 width	26–33 (30)
Abdominal somite 3 length	10–14 (12)
Abdominal somite 3 width	25–31 (29)
Caudal ramus length	22–26 (25)
Caudal ramus width	11–14 (13)
Caudal ramus seta 1 length	28–39 (32)
Caudal ramus seta 2 length	45–65 (53)
Caudal ramus seta 3 length	22–36 (31)
Caudal ramus seta 4 length	56–82 (68)

^aExcluding the length of the caudal setae.

These novelties/discrepancies noted above were not considered sufficient to propose a new taxon (as discussed in detail in the next session) hence only a redescription of this species was provided herein.

Table 7. Armature of swimming legs (L1 to L3) of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female

Swimming leg	Coxa	Basis	Endopod	Exopod
L1	0–0	0–1	0–1; 0–3	I–0; 0–1; II–5
L2	0–0	0–1	0–1; 0–2; I–4	I–0; 0–1; I–6
L3	0–0	0–1	0–1; 0–1; I–4	I–0; 0–1; 0–6

(Roman numeral = spines; Arabic numerals = setae).

Gamispatulus schizodontis Thatcher and Boeger, 1984 (Fig. 8; Table 8)

Type host: *Schizodon fasciatus* Spix and Agassiz, 1829 (Characiformes: Anostomidae).

Other hosts: *Hoplias lacerdae* Miranda Ribeiro, 1908 (Characiformes: Erythrinidae); *Hoplias malabaricus* (Bloch, 1794) (Characiformes: Erythrinidae); *Leporinus friderici* (Bloch, 1794) (Characiformes: Anostomidae); *Leporinus lacustris* Amaral Campos, 1945 (Characiformes: Anostomidae); *Megaleporinus elongatus* (Valenciennes, 1850) (Characiformes: Anostomidae); *Megaleporinus obtusidens* (Valenciennes, 1837) (Characiformes: Anostomidae); *Pimelodus maculatus* Lacepède, 1803 (Siluriformes: Pimelodidae); *S. maculatus* Kner, 1858 (Characiformes: Serrasalminidae) – PS; *Serrasalmus marginatus* Valenciennes, 1837 (Characiformes: Serrasalminidae); *Schizodon borellii* (Boulenger, 1900) (Characiformes: Anostomidae); and *Schizodon intermedius* Garavello and Britski, 1990 (Characiformes: Anostomidae).

Type locality: Ilha da Marchantaria, Solimoes-Amazonas River near Manaus, Amazonas state, Brazil.

Sampled localities: Area 6 – Mouth of the Pardo River in the municipality of Salto Grande, state of São Paulo, Brazil (22° 54'41.7"S, 49°57'43.1"W).

Site of infestation: Nostrils.

Material examined: A total of 12 ♀♀ from nostrils of *S. maculatus* sampled in area 6 were examined for morphological description.

Representative DNA sequences: PQ442938, PQ442940–41, PQ450081, PQ452576–77, PQ452579.

Voucher material: A total of 6 whole ♀♀ stored in glass vials with 96% ethanol were deposited in the Zoological Collection of MZUSP (accession no. 46663 and 46792).

Identification (based on adult females; male unknown): Present specimens showed all diagnostic features proposed by Thatcher and Boeger (1984) for *G. schizodontis*, as well as the new features observed by Narciso and Silva (2020) when redescribing this species. Spine and setal formula of biramous swimming legs are summarized in Table 8.

Remarks

The copepods found inside the nostrils of *S. maculatus* were identified as members of the 'vaigamid' subgroup by having the combination of cephalosome armed with dorsolateral stylets (or retrostylets) and leg 1 with endopod 2-segmented. To date, 5 ergasilid genera are included in this subgroup, namely: *Gamidactylus* Thatcher and Boeger, 1984; *Gamispatulus*; *Gamispinus* Thatcher and Boeger, 1984; *Pseudovaigamus* Amado et al., 1995; and *Vaigamus* Thatcher and Robertson, 1984. Among all vaigamids, the present copepods were identified as members of the genus *Gamispatulus* by having the following combination of diagnostic features: (i) rostrum armed with a rostral spine; (ii) antennule 5-segmented and armed with 2 claws (middle and inner claw); and (iii) L4 with enp 2-segmented and exp 1-segmented.

Currently, the genus *Gamispatulus* consists of 2 species, *G. schizodontis* (type-species) and *Gamispatulus ferrilongus* Narciso and Silva, 2020. The present copepods were identified as *G. schizodontis* due to the possession of a spatulate process on the retrostylets (see Fig. 7) – which is an exclusive feature of this species. Additionally, they also resemble the type species by the size of the rostral spine (i.e. short as in *G. schizodontis* rather than long as in *G. ferrilongus*), the ornamentation of thoracic appendages (including antennule, antenna and swimming legs), and by the number of setae on caudal ramus (i.e. 2 setae as in *G. schizodontis* rather than 4 as in its congener).

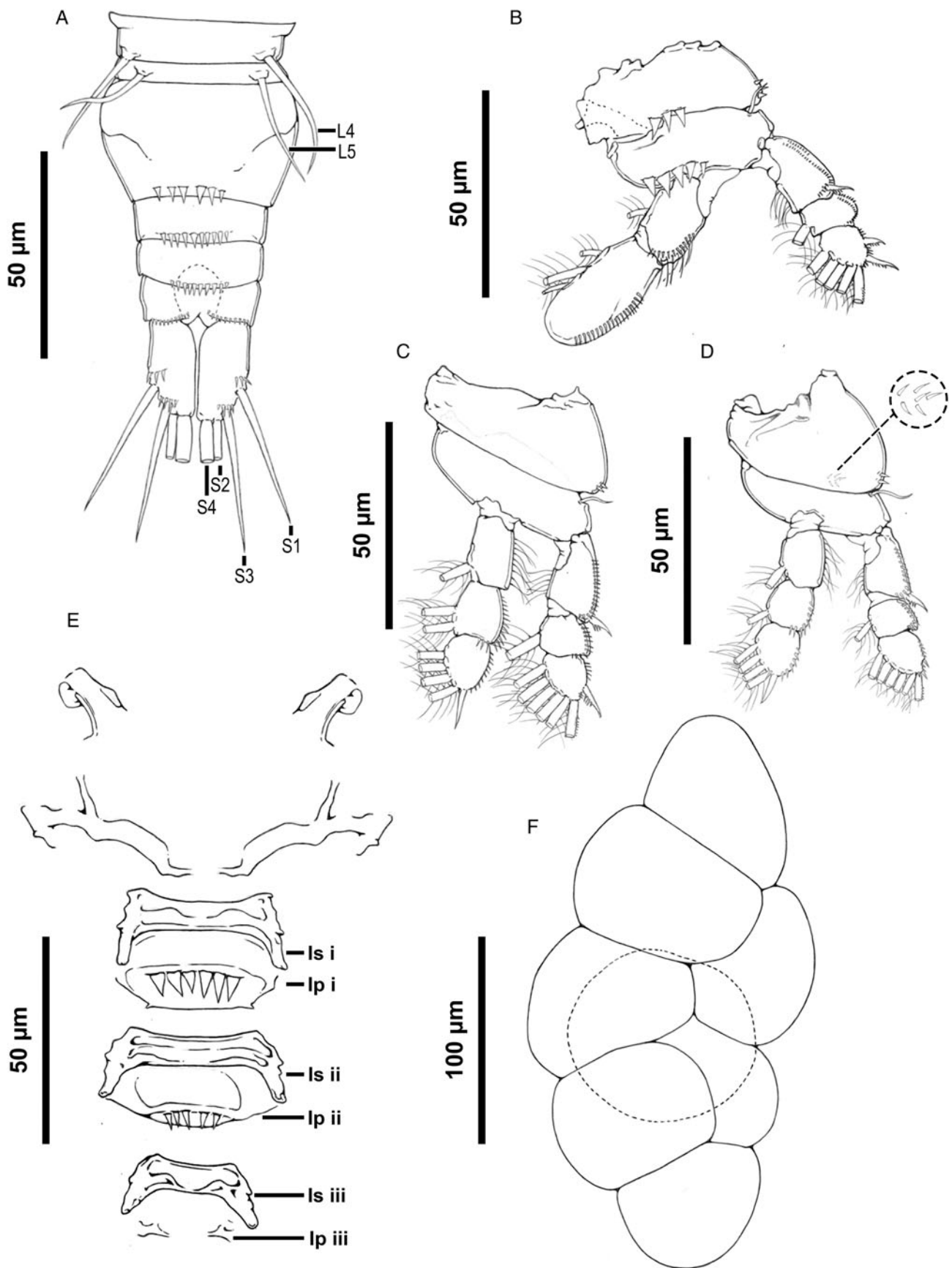


Figure 4. *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female, (A) fourth pedigerous somite, urosome and caudal rami with setae (S1 to S4), ventral view: fourth and fifth somites with leg 4 (L4) and leg 5 (L5) each reduced to a single seta, respectively, (B) leg 1, (C) leg 2, (D) leg 3 with coxa ornamented with a patch of spinules near the posterior margin (dashed circle), (E) intercoxal sclerites (Isi to Isiii) and interpodal plates (Ipi to Ipiii), (F) egg sac. Scale bars in micrometres (μm).

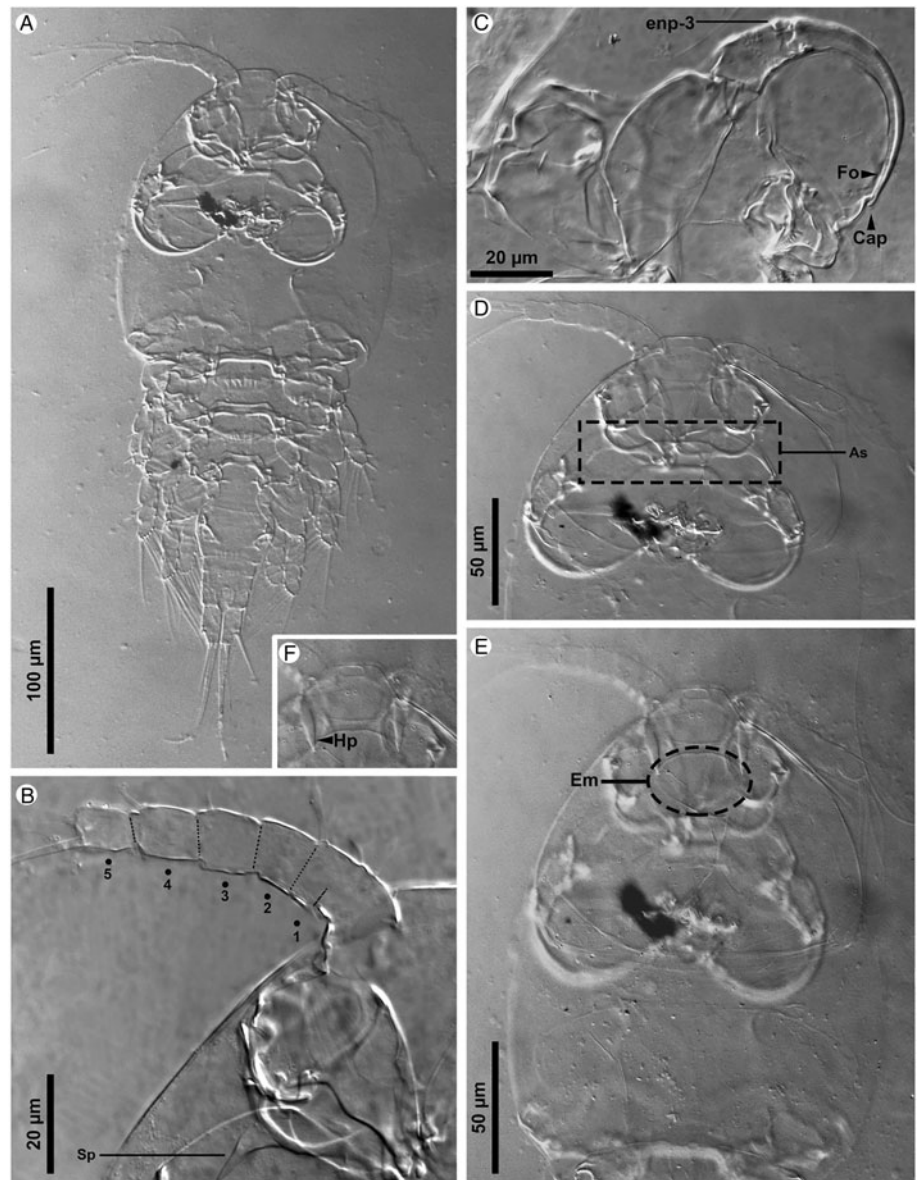


Figure 5. Light micrographs of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female, (A) body, ventral view, (B) antennule segments (1–5) and the antennal support armed with lateral spine-like projection (Sp), (C) antenna with reduced third endopodal segment (enp-3) and claw with fossa (Fo) and distal hyaline cap (Cap), (D) cephalosome, ventral view, with antennal support (As), (E) cephalothorax, dorsal view, with dorsal cephalic shield with elliptical mark (Em), (F) horn-like projections (Hp). Scale bars in micrometres (μm).

Molecular and phylogenetic analyses

A total of 12 novel sequences were generated (Table 3). Several attempts were made to obtain 18S rDNA sequences for *R. piranhus* (i.e. including variations in the number of cycles and temperature during PCR), but no positive results were obtained. The phylogenetic reconstructions using ribosomal genes (18S and 28S) recovered the monophyly of Ergasilidae with high support values (i.e. 100 and 0.99 for ML and BI analyses, respectively) and positioned the 2 studied species (*R. piranhus* and *G. schizodontis*) as members of this family.

The final alignment of the 28S rDNA dataset comprised a total of 49 sequences and generated 659 characters for analyses (Supplementary Table 1). The estimated distance between the novel sequences of *R. piranhus* and *G. schizodontis* and other ergasilids ranged between 10.7–12.5 and 9.0–11.8%, respectively. Among the *R. piranhus* sequences, the intraspecific distance varied between 0.5 and 0.1%. The unique 28S rDNA sequence of *G. schizodontis* diverged from those of *R. piranhus* by 7.0–7.2%. Regarding the 18S rDNA, the final alignment comprised a total of 52 sequences and resulted in a length of 1.121 bases (Supplementary Table 2). The 2 sequences obtained from *G. schizodontis* diverged from each other by 1.38% and from the other ergasilids by 1.2–3.9%.

Overall, the topologies for the ML and BI analyses for both ribosomal genes (18S and 28S rDNA) were congruent. Strong support values were obtained from 5 terminal clades in Ergasilidae, but relationships between those clades showed (mostly) low support (Figs. 9 and 10). The monophyly of type-genus, *Ergasilus* Nordmann, 1832 was not recovered.

The well-supported (>0.7 ML/70 BI) terminal clades were: clade I or Sub-Saharan clade – including *Ergasilus* spp. from Lake Tanganyika plus *Ergasilus mirabilis* Oldewage and van As, 1987 from Burundi, South Africa and Zambia; clade II – including *Paraergasilus* spp. plus *Ergasilus wilsoni* Markevich, 1933; clade III – including *Ergasilus* spp. plus *Neoergasilus japonicus* (Harada, 1930); clade IV – constituted by *Sinergasilus* spp. plus *Ergasilus anchoratus* Markevich, 1946. For 28S, a clade composed of Neotropical species (or clade V) was also recovered – it includes the novel sequences provided herein (for *R. piranhus* and *G. schizodontis*) plus sequences of *Acusicola marguliseae* Santacruz *et al.*, 2020 (Fig. 9). The latter clade was also recovered for 18S but with low support (Fig. 10). Except for *Ergasilus*, the other multi-sequenced genera (i.e. genera with more than 1 species with available sequences in NCBI database) had their monophyly recovered with high support values, including (i) *Neoergasilus* Yin, 1956; (ii) *Paraergasilus* Markevich, 1937; and (iii) *Sinergasilus*.

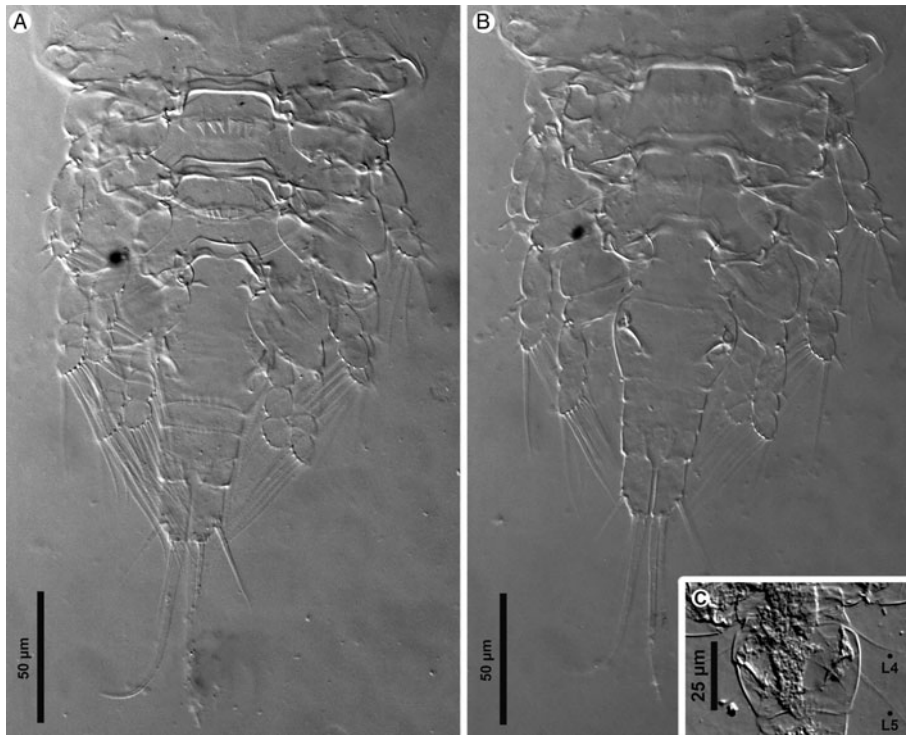


Figure 6. Light micrographs of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female, (A) posterior part of the body, ventral view, (B) posterior part of the body, dorsal view, (C) fourth and fifth pedigerous somites with legs reduced to a single seta (L4 and L5). Scale bars in micrometres (μm).

The final cox 1 alignment comprised a total of 59 sequences and the final dataset, after trimming the extremes, resulted in 510 bases. This dataset was only used to calculate genetic difference matrices. The BI and ML analyses produced phylograms with unsupported clades and unsatisfactory placements of the sequences used in the dataset (see Supplementary Fig. 3). The intraspecific distance observed between *G. schizodontis* sequences ranged from 0.9 to 2.1%, while the only cox 1 sequence obtained for *R. piranhus* differed from other ergasilids by 20.5–21.6%. Within Ergasilidae, the observed interspecific divergence varied from 0.9 to 22.4% (see Supplementary Table 3).

Discussion

Morphological comparisons and redescription

Several morphological variations were found between the present specimens and the original description of *R. piranhus*. Examination of the type-material was also made; however, due to their poor state of preservation it was not possible to verify the condition of most of the new features in those copepods. Despite that, some comparisons were made concerning the morphology of the urosome, caudal rami and part of the antennae (including claw) as these parts were relatively well preserved in some of the type specimens (see Supplementary Fig. 1). In the caudal ramus of the holotype, it was possible to visualize the lateral seta (or s-1) and, as proposed herein, this seta is relatively long when compared to the same seta present in *R. digitus* and *R. unguilongus* (i.e. s-1 is several times smaller when compared to s-2 and s-4 in the latter 2 species). Something similar was also noted concerning the shape and size of the antennal claw (preserved in paratypes): in *R. piranhus* the claw has the same length as the first endopodal segment (or enp-1), and its internal angle is about 180 degrees which gives it a more rounded shape, whereas, in its 2 congeners, the claw has a different size (i.e. being smaller than enp-1 in *R. digitus* while in *R. unguilongus* it is the longest antennal structure) and the internal angle is always acute (apparently less than 45 degrees in both species). The above features were also observed in our specimens and agree with the

original description of *R. piranhus*. Therefore, most of our comparisons were primarily made relying on drawings and the description text proposed for this species by Boeger and Thatcher (1988).

Differences in the number of setae (or setation) and the segmentation of cephalothorax were the 2 main discrepancies observed between our specimens and the original description of *R. piranhus*. Discrepancies in setation are one of the most common discrepancies pointed out in ergasilid redescrptions (see Chernysheva and Purasjoki, 1991; Kim and Nagasawa, 2006; Marques *et al.*, 2015; Marques and Boeger, 2018; Narciso and Silva, 2020). Due to their small size, setae can be easily lost due to breakages during the copepod handling or even being covered by other structures in non-dissected specimens (Boxshall, 2016). Despite the segmentation of cephalothorax being considered more evident when compared with the presence/absence of a certain seta, the correct description of this segment is also ‘tricky’ in Ergasilidae and can vary substantially depending on the taxonomist interpretation. In Marques *et al.* (2015), the authors verified several incongruences in the cephalothorax segmentation when reassessing the type-material of *Ergasilus* species. Similar to the setation, such inconsistencies on cephalothorax may be related to the copepod handling when preparing them for morphological identification – e.g. covering copepods with a coverslip on a flat microscope slide may cause them to be crushed/smashed and consequently obscuring the real segmentation of the cephalothorax (personal observation). Less invasive mounting techniques, such as the wooden slide proposed by Humes and Gooding (1964), might be viable alternatives to avoid such errors, which is particularly important for ‘big’ ergasilids – i.e. since they are more affected by such preparations. For small size species such as *R. piranhus*, the use of SEM proved useful to achieve a more accurate description of this somite – i.e. in the present study, the SEM allowed for the visualization of small details like the pores and elliptical marks, as well as the ‘flexible cuticle’ (or FC) that separates the cephalosome from the PS-1 which corroborates our observation that the cephalothorax in this species is subdivided (see Fig. 7B).

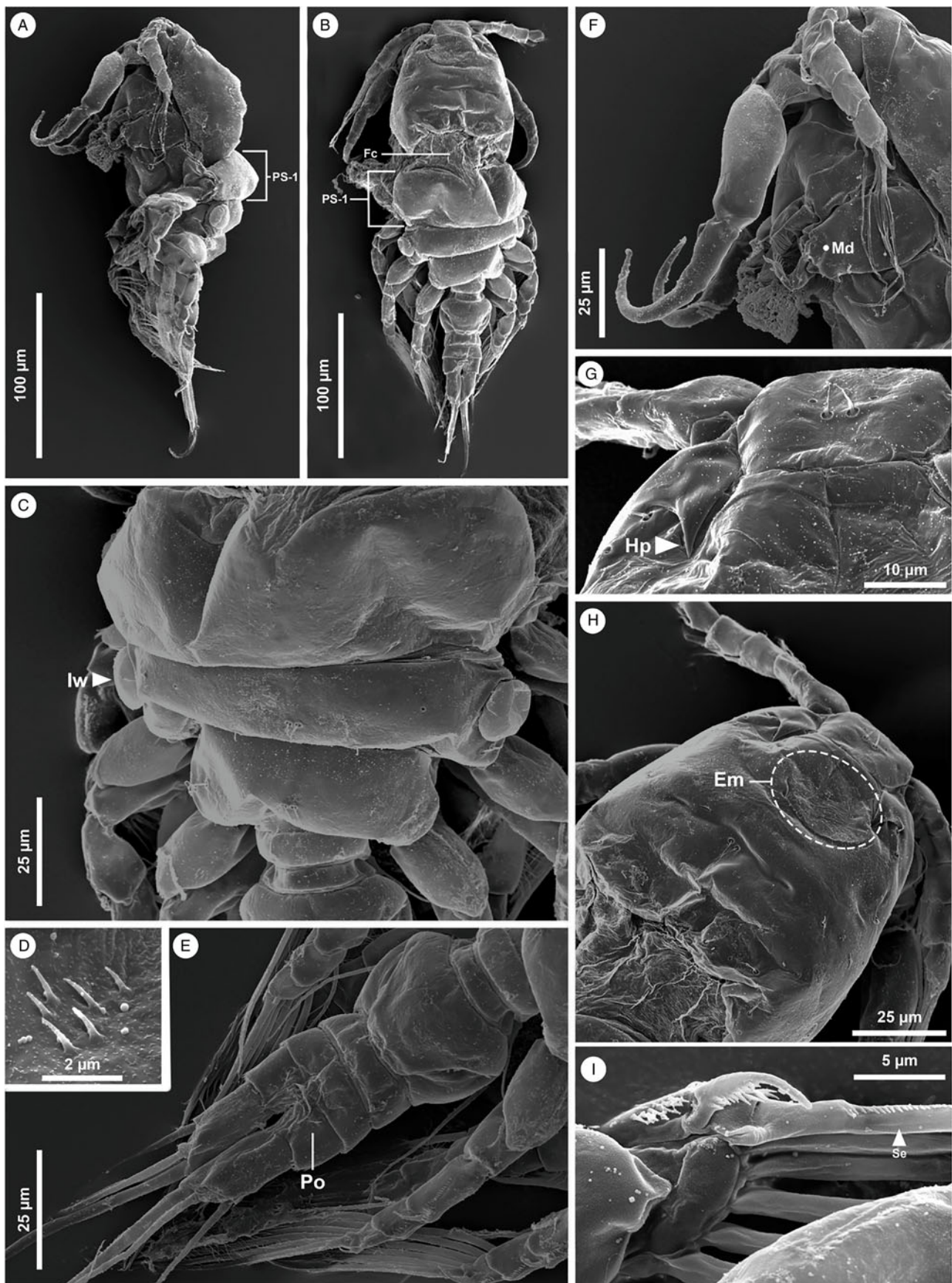


Figure 7. Scanning electron micrographs of *Rhinergasilus piranhus* Boeger and Thatcher, 1988 – adult female, (A) body, lateral view, with first pedigerous somite (PS-1) separated from cephalosome, (B) body, dorsal view, with flexible cuticle (Fc) separating cephalosome from prosome (PS-1), (C) pedigerous somite, dorsal view, with second pedigerous somite with paired integumental windows laterally on tergite (Iw), (D) detail of the patch of spinules from coxa of leg 3, (E) urosome, dorsal view, with third abdominal segment ornamented with a pair of pores (Po) carrying bristles (Po), (F) cephalothorax, lateral view, showing some parts of buccal apparatus such as the mandible (Md); (G) horn-like projections (Hp), (H) cephalothorax, dorsal view, with dorsal cephalic shield with elliptical mark (Em), (I) detail of the spinules (Se) on the first seta of third exopodal segment of leg 2. Scale bars in micrometres (μm).

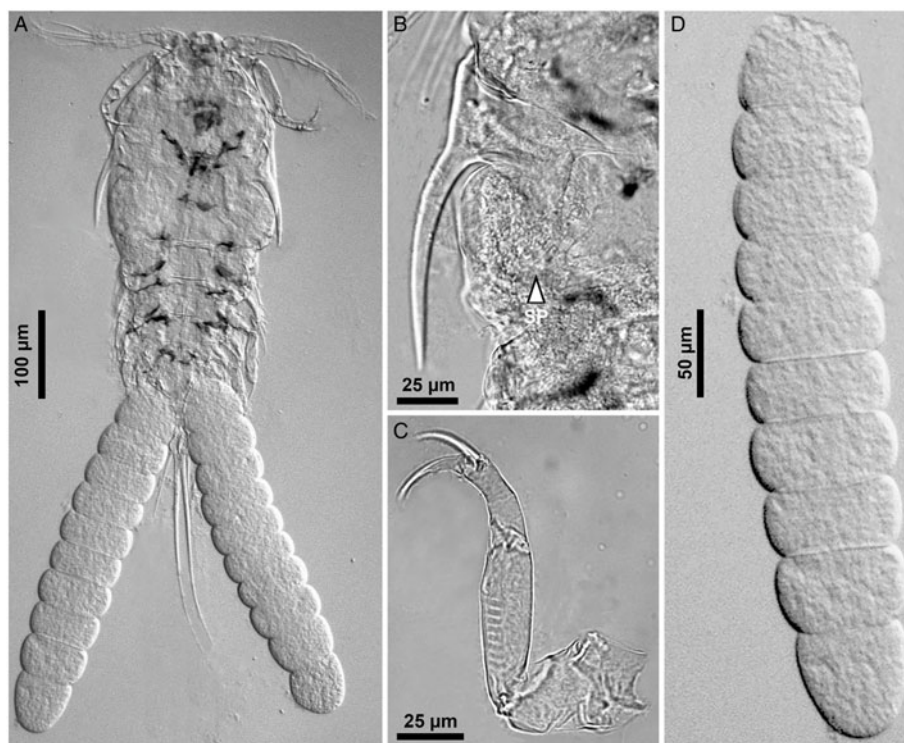


Figure 8. Light micrographs of *Gamispatulus schizodontis* Thatcher and Boeger, 1984 – adult female, (A) body, ventral view, (B) retrostylet armed with spatulate process (SP), (C) antenna, (D) uniseriate egg sac. Scale bars in micrometres (μm).

Our analyses also showed some novel features not previously described for *R. piranhus*. The dorsal cephalic shield is equipped with a pair of horn-like structures located immediately below the antennules (Figs 5F and 7G). Ventrally, spine-like projections were also observed for both sides of the antennal support (i.e. structure present on the ventral side of cephalosome that articulates with both antennae) (Fig. 5D). Although such ‘acute’ structures do not have any defined function, they can act as a complementary apparatus for its species attachment inside the nostrils – acting similarly to the retrostylets and/or rostral spine present in all ‘vaigamids’ like *G. schizodontis* (discussed in details below) (Narciso and Silva, 2020). The other 2 *Rhinergasilus* species were described as gill parasites and lacked such structures (Narciso *et al.*, 2021). However, additional data such as behavioural or *in situ* observations of living specimens are still necessary to prove this assumption. Other minor novelties were also observed regarding the ornamentation of the body and appendages.

Similarly to the discrepancies, the previous novelties were probably also difficult to visualize due to their relatively small size in the original description. Most of the novelties were found in the cephalothorax, which is a segment that is generally not dissected and has various ‘obstacles’ that compromise its correct characterization (e.g. internal organs, presence of pigments, etc.). As noticed in the type-material (see Supplementary

Fig. 1), the presence of internal lumps and/or tissues obscured our observation of most diagnostic features. In our analyses under the compound microscope, it was possible to observe most of the proposed novelties (as seen in Figs. 5 and 6), but it was only achieved due to the use of clarifying agents – e.g. lactic acid, Hoyer’s medium, etc.

Descriptions that detail the ornamentation of the body somites are relatively recent and still little practiced in taxonomic studies of ergasilids. Alston *et al.* (1993) provided the first detailed description of pores throughout the body of an *Ergasilus* species (i.e. made during the redescription of *Ergasilus briani* Markevich, 1933). In recent descriptions of Brazilian ergasilids, similar pores were also observed on the cephalothorax of different species (including species from distinct genera), which indicates that these ornaments are probably very common in the Ergasilidae (Narciso *et al.*, 2022; Couto *et al.*, 2023). Regarding the antennal support, *Ergasilus bryconis* Thatcher, 1981 is the only species in which this structure was represented in its description. Details of the ventral side are even scarcer in Ergasilidae since most descriptions only represent/describe the dorsal side of the copepods. Despite their current low taxonomic importance, structures such as the antennal support can provide relevant diagnostic features (as presented herein) and should therefore be better explored in future descriptions.

In the Neotropical region, the cephalosome armed with dorso-lateral stylets (or retrostylets) is a diagnostic feature unique to the copepods from the ‘vaigamid’ subgroup. Similar stylets were also reported for 5 species of *Paraergasilus* Markevich, 1937 (*Paraergasilus brevidigitus* Yin, 1954; *Paraergasilus longidigitus* Yin, 1954; *Paraergasilus medius* Yin, 1956; *Paraergasilus mimus* Yin, 1962; and *Paraergasilus rylovi* Markevich, 1937); however, these structures are usually described as spinous protrusions derived from the posterolateral margin of cephalosome instead of well-defined, mobile structures carrying sensory organs (i.e. pores and sensilla), as occurs in all vaigamids (Thatcher and Robertson, 1984). No developmental studies are available for representatives of both genera, making it difficult to establish the putative homology of these structures; however, other

Table 8. Armature of swimming legs (L1 to L4) of *Gamispatulus schizodontis* Boeger and Thatcher, 1988 – adult female

Swimming leg	Coxa	Basis	Endopod	Exopod
L1	0-0	0-1	0-1; II-5	II-0; 0-1; II-5
L2	II-0	0-1	0-1; 0-2; I-4	I-0; 0-1; 0-6
L3	II-0	0-1	0-1; 0-2; I-4	I-0; 0-1; 0-6
L4	V-0	0-1	0-1; 0-4	II-4

(Roman numeral = spines; Arabic numerals = setae).

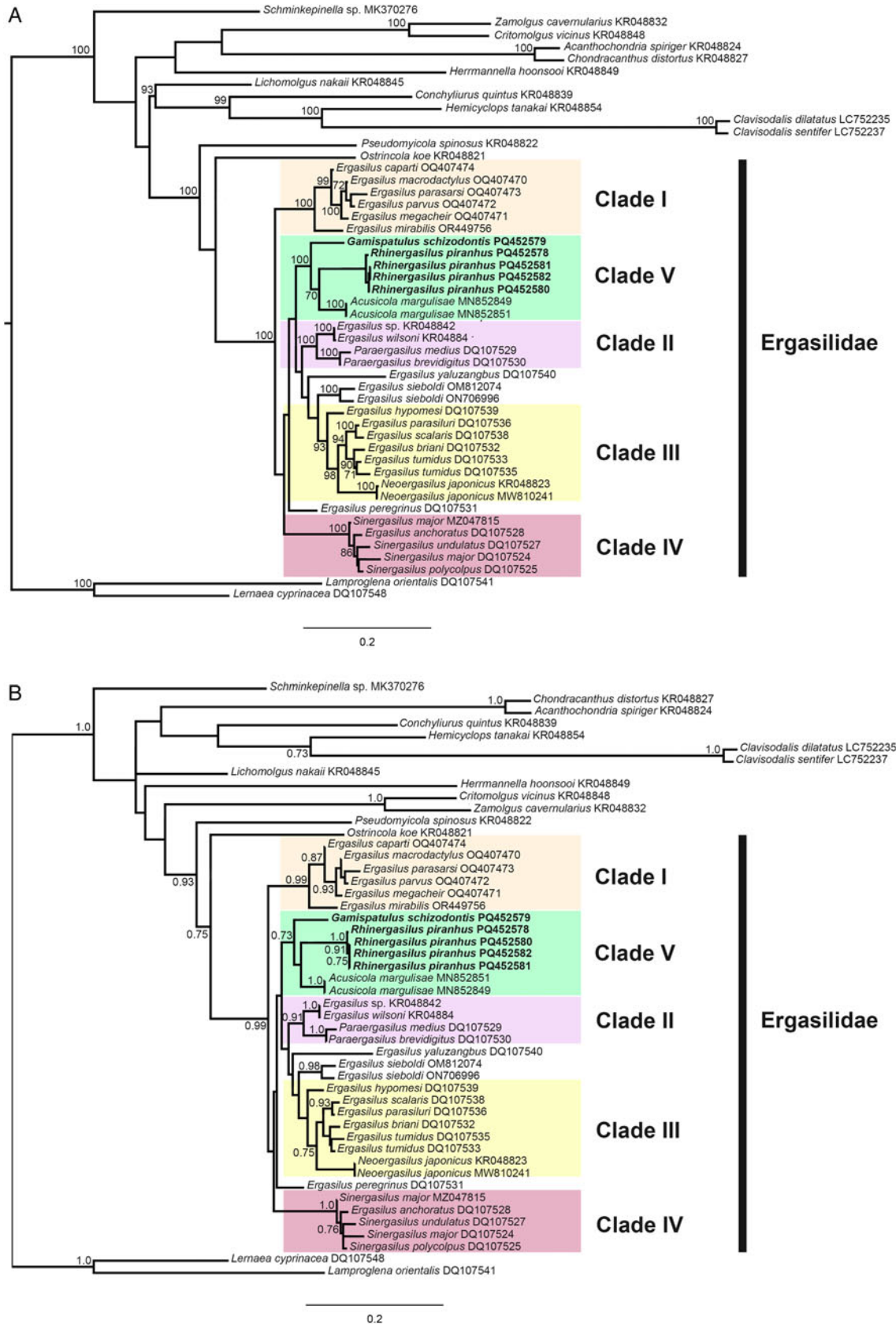


Figure 9. Phylogenetic tree of Ergasilidae copepods based on partial 28S rRNA gene alignments. Newly generated sequences for *Rhinergasilus piranhus* Boeger and Thatcher, 1988 and *Gamispatulus schizodontis* Thatcher and Boeger, 1984 are provided in bold. Nodal support presented above or below branches for (A) Bayesian inference (>0.7) and (B) maximum likelihood (>70%) analyses (BI/ML). Sequences of *Lemaea cyprinaea* Linnaeus, 1758 and *Lamproglena orientalis* Markevich, 1936 were both used as outgroup.

diagnostic features allow for the easy differentiation of both groups, such as (1) antenna armed with 1 or 2 claws in vaigamids vs 3 claws in *Paraergasilus*; (2) rostrum armed with rostral spine

as present in *Gamispatulus*, *Pseudovaigamus* and *Vaigamus* vs lacking such structure in *Paraergasilus*; (3) L1 enp 2-segmented in all vaigamids vs 3-segmented in *Paraergasilus*; and (4) L5

represented by 1- or 2-setae in vaigamids vs 1-segmented as in *Paraergasilus*. The geographical distribution of these 2 groups is also different: *Paraergasilus* has a wide distribution, with species reported from African, European, Asian and Australian countries (El-Rashidy and Boxshall, 2001a) while all vaigamids are currently restricted to the Brazilian region (Amado *et al.*, 1995). The phylogenetic relationships between these 2 groups are still unclear as the available phylogenetic hypotheses (based on morphological data) are contradictory: (i) Amado *et al.* (1995) and Tang and Kalman (2008) recovered both groups in separated clades – *Paraergasilus* was recovered at the base of Ergasilidae acting as the most ‘ancestral-like’ ergasilid due to the retention of the tree-clawed antennae (in conjunction to other plesiomorphic characters), while vaigamids occupied a more internal position, closer to the other Neotropical genera; whereas in (ii) El-Rashidy (1999) both groups were recovered in a single clade at the base of the family, except for *Pseudovaigamus spinicephalus* who clustered inside the clade ‘Avii’ which consisted exclusively of Neotropical species. Our phylogenetic analyses (discussed in detail in the next section) recovered both groups in 2 separate clades (clade II and V, see Figs 9 and 10); however, the low ML and BI support between the terminal clades did not allow us to completely resolve their positions. Yet, the closer relationship observed between *G. schizodontis* with other Neotropical ergasilids (i.e. *A. margulisiae* and *R. piranhus*) in clade V indicates that the proposal originally made by Amado *et al.* (1995), which includes vaigamids alongside the other Neotropical ergasilids, is probably more accurate.

Regarding the morphology of *G. schizodontis*, the present copepods were consistent with the redescription proposed by Narciso and Silva (2020) for this species. Novelty and/or discrepancies were not observed for this species.

Molecular phylogenetics

In the last 20 years, the number of molecular sequences available for ergasilids has increased (Santacruz *et al.*, 2020; Kvach *et al.*, 2021; Oliveira *et al.*, 2021; Abdel-Radi *et al.*, 2022; Fikiye *et al.*, 2023; Mič *et al.*, 2023, 2024). Especially after the publication of the phylogenies based on ribosomal genes (18S and 28S rDNA) done by Song *et al.* (2008), in which the relationship of Chinese ergasilids (including species of *Ergasilus*, *Neoergasilus*, *Paraergasilus*, *Pseudergasilus* and *Sinergasilus*) was explored for the first time. Among their main results, the polyphyletic status of the type-genus *Ergasilus* was achieved corroborating previous findings made by El-Rashidy (1999) when exploring the phylogeny of this taxon based on morphological features. In our analyses, the polyphyletic status of *Ergasilus* was also recovered – i.e. sequences of the type-species, *Ergasilus sieboldi* Nordmann, 1832 tended to appear as an isolate terminal clade while other *Ergasilus* spp. are scattered across different clades (Figs 9 and 10). For the MI of the 18S gene, *Ergasilus yaluzangbus* Kuang and Qian, 1985 appeared as a sister taxon to *E. sieboldi* but with low support. Since the molecular phylogeny proposed by Song *et al.* (2008), *E. yaluzangbus* tends to ‘jump’ between clades in the available phylogenies, with no defined phylogenetic relationships. The same applies to *E. sieboldi* which, depending on the gene used, has different relationships within Ergasilidae [see phylogenies present in Fikiye *et al.* (2023)]. A major revision of this genus is strongly required but due to several impediments (e.g. a huge number of known species, wide distribution, difficulty in obtaining the type-material, low availability of taxonomists specialized in ergasilids, etc.) such revision is difficult to carry out. Furthermore, the uncertain relationship between *E. sieboldi* (type-species) and its congeners makes it even more difficult to define which are the ‘true’ members of this speciose genus. Despite

this, we believe that minor revisions can be made by exploring highly supported terminal clades that include *Ergasilus* spp.

Connections between terminal clades had low support in our analyses making it impossible to establish reliable phylogenetic relationships between those clades. However, most of the terminal clades showed high support values, resulting in 5 main clades (clade I to V) in our analyses (Figs 9 and 10). Clade I (or Sub-Saharan clade) comprised 6 *Ergasilus* species [*Ergasilus caparti* Mič *et al.*, 2023; *Ergasilus macrodactylus* (Sars G.O., 1909); *Ergasilus megacheir* (Sars G.O., 1909); *E. mirabilis* Oldewage and van As, 1987; *Ergasilus parasarsi* Mič *et al.*, 2023; and *Ergasilus parvus* Mič *et al.*, 2023], all of them described from African countries (mostly in Lake Tanganyika). Recently, Fikiye *et al.* (2023) redescribed *E. mirabilis* Oldewage and van As, 1987 sampled in South Africa and Zambia based on an integrative approach (i.e. including morphological description based on light and SEM microscopy in conjunction with molecular and phylogenetic analyses) and the same well-supported clade was recovered in their analyses. This African group of *Ergasilus* has an antenna with a very particular shape that is quite different from the ‘standard’ shape exhibited by other species of *Ergasilus* like *E. sieboldi*. A more detailed review of this group is necessary, but we believe that this group consists of a putative new genus for this family.

The clade II is constituted by a monophyletic group comprising *Paraergasilus* spp. with *E. wilsoni* Markevich, 1933 (together with *Ergasilus* sp. – KR048842 in 28S analyses) as its sister taxon. *Paraergasilus* Markevich, 1937 is considered a monophyletic genus that retains the (putative) ancestral state of the antenna, which in this taxon is armed with 3 terminal claws [i.e. loss of the lateral claws was considered a synapomorphy in Ergasilidae by Amado *et al.* (1995), El-Rashidy (1999) and Tang and Kalman (2008)]. To date, this genus comprises 16 valid species but only 3 of them have sequences available (for the 18S and/or 28S ribosomal genes): *P. brevidigitus* Yin, 1954; *P. longidigitus* Yin, 1954; and *P. medius* Yin, 1956. Latter species together with *P. rylovi* Markevich, 1937 (type-species) and *P. mimus* Yin, 1962 (the latter 2 without available sequences) can be distinguished from their congeners by having a pair of lateral projections on the cephalothorax (El-Rashidy and Boxshall, 2001a). This feature resembles the ‘retrostylets’ present in Neotropical vaigamids such as *G. schizodontis* but due to the unclear relation between clades and the lack of sequences for the other *Paraergasilus* that lack such projections, it was not possible to trace the evolutionary history of this character either within the genus or family. *Ergasilus wilsoni* was originally described from the Black Sea, which is the marginal Mediterranean Sea located between Europe and Asia (Markevich, 1933). This copepod has a similar morphology to other *Ergasilus* which includes an antenna equipped with a single claw. Nevertheless, phylogenies based on ribosomal genes, including those of the present study, have continuously recovered *E. wilsoni* as a sister taxon to *Paraergasilus*, indicating that this species may also be a good target for future revisions.

Within clade III, 2 subclades were recovered: (i) subclade III.1 formed by 4 species of *Ergasilus* (i.e. 5 for 28S analyses, since *Ergasilus scalaris* Markevich, 1940 was also recovered as part of this subclade); and (ii) subclade III.2 formed by sequences of *N. japonicus* (Haradam, 1930). The position of *Ergasilus hypomesi* Yamaguti, 1936 is uncertain as it varies according to the gene. For 18S, this species was recovered within clade III closer to *N. japonicus* while for 28S it was recovered as the sister taxon of the 2 subclades (Fig. 9). Within subclade III.1, those species share some common features like the fifth leg reduced to a papilla carrying a single seta, and a tendency for somites to fuse (i.e. a feature present in all *Ergasilus* species once classified as *Pseudergasilus* Yamaguti, 1936). Kim and Nagasawa (2006),

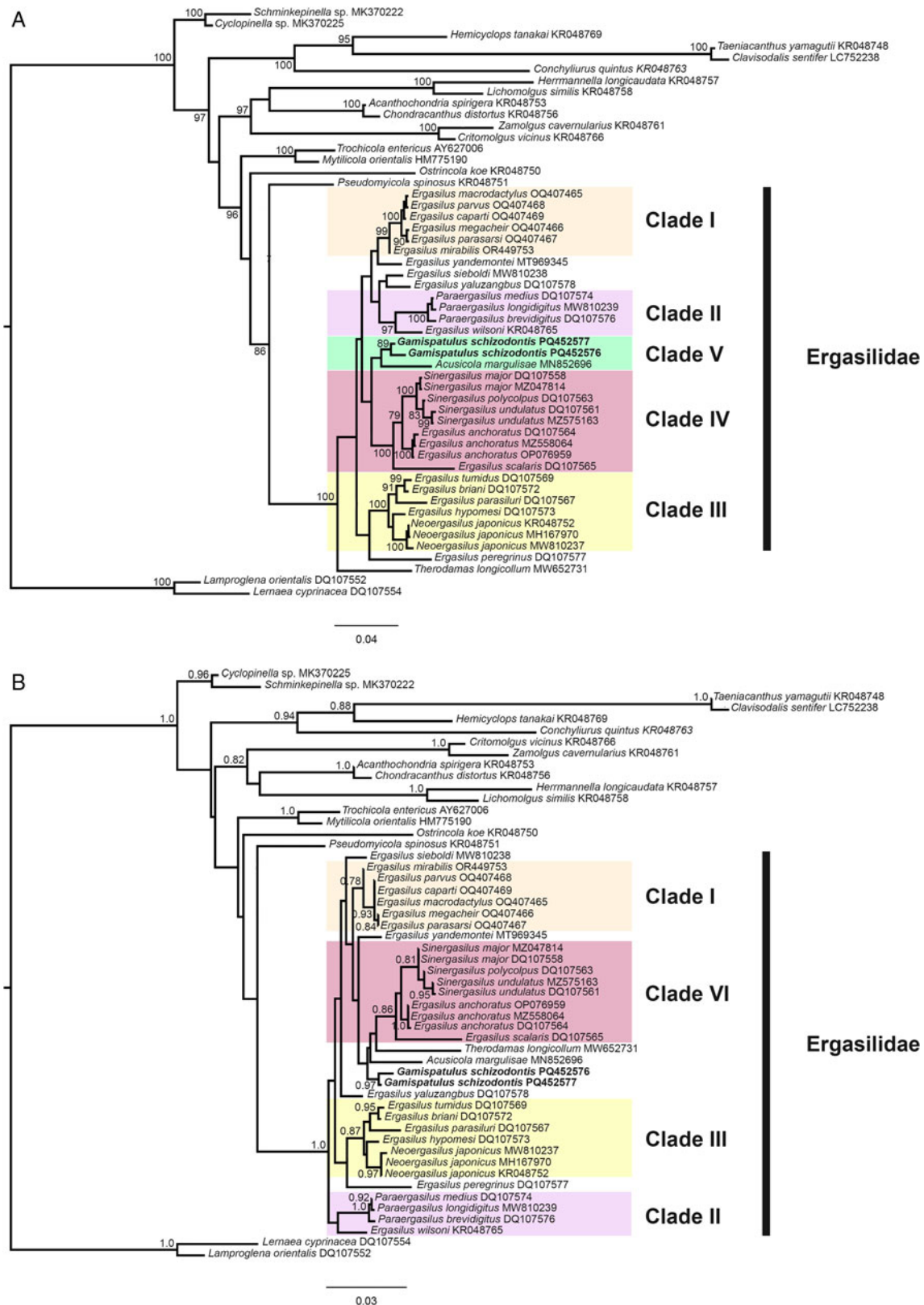


Figure 10. Phylogenetic tree of Ergasilidae copepods based on partial 18S rRNA gene alignments. Newly generated sequences for *Rhinergasilus piranhus* Boeger and Thatcher, 1988 and *Gamispatulus schizodontis* Thatcher and Boeger, 1984 are provided in bold. Nodal support presented above or below branches for (A) Bayesian inference (>0.7) and (B) maximum likelihood (>70%) analyses (BI/ML). Sequences of *Lernaea cyprinacea* Linnaeus, 1758 and *Lamproglena orientalis* Markevich, 1936 were both used as outgroup.

when redescribing *Pseudergasilus zacconis* Yamaguti, 1936, pointed out that these 2 aforementioned features are not exclusive to *Pseudergasilus* in contrast to Yamaguti (1936) proposal, which reinforced the invalidation of this genus. However, given the

polyphyletic status of *Ergasilus* and the high support recovered for the subclade III.1, it indicates that the invalidation of *Pseudovaigamus* needs to be checked and this subclade can be another good target for revision in Ergasilidae.

Since the publication of Song *et al.* (2008), the 3 valid species of *Sinergasilus* Yin, 1949 and *E. anchoratus* Markevich, 1946 constitute a highly supported group within Ergasilidae (see phylogenies in Santacruz *et al.*, 2020; Kvach *et al.*, 2021; Oliveira *et al.*, 2021; Abdel-Radi *et al.*, 2022; Fikiye *et al.*, 2023; Mič *et al.*, 2023; Mič *et al.*, 2024). The same clade is also recovered in the present study. The placement of *E. anchoratus* in clade IV is strongly supported; however, its relationships within the clade are still uncertain as it can act as a sister taxon for *Sinergasilus* species or even as an insider into the latter genus (i.e. causing *Sinergasilus* to be paraphyletic) (Figs 9 and 10). Song *et al.* (2008) when evaluating this clade pointed out that *Sinergasilus* might have evolved from an ancestral *Ergasilus* species possibly due to host switching. Recently, the same gene arrangement in the mitochondrial genome was observed by Hua *et al.* (2024) for *E. anchoratus* and *Sinergasilus undulatus* (Markevich, 1940) providing additional evidence that supports the close relationship between those copepods in available phylogenies.

Finally, the novel sequences generated for *R. piranhus* and *G. schizodontis* formed a clade (clade V or Neotropical clade) in conjunction with *A. margulisae*. The latter species was described by Santacruz *et al.* (2020) based on copepods found parasitizing 3 cichlids [*Amphilophus citrinellus* (Günther, 1864); *Parachromis managuensis* (Günther, 1867); and *Oreochromis* sp.] and 1 poeciliid (*Poecilia mexicana* Steindachner, 1863) from a crater lake in Nicaragua. With the exception of *Acusicola tenax* (Roberts, 1965) which was originally described in Texas, USA (Roberts, 1965), the remaining *Acusicola* species occur exclusively in the Neotropical region (Santacruz *et al.*, 2020; Couto *et al.*, 2023). Based on the historical biogeography of freshwater Cyclopinae copepods (Suárez-Morales *et al.*, 2004), *Acusicola* probably also originated in South America and then spread northwards as a consequence of the faunal exchange in conjunction with the closure of the Panama gap during the Pliocene explaining the occurrence of this single species in the Nearctic region. *Rhinergasilus* and *Gamispatulus* are restricted to Brazil (see the checklists in Narciso and Silva, 2020; and Narciso *et al.*, 2020). The Neotropical clade was strongly supported in 28S analyses, while for 18S a similar clade was recovered but with low support. More sequences of Neotropical species are required to test the validity of clade V.

The genus *Therodamas* Krøyer, 1863 is also restricted to the Neotropical region. Like the other mesoparasitic genera (i.e. species whose adult females live embedded into the host tissues), *Therodamas* species are also easily recognized by their long 'necks' (El-Rashidy and Boxshall, 2001b; Tang and Kalman, 2008). Despite their morphological similarities, the origin and/or relationships between mesoparasitic taxa remain unclear in Ergasilidae – the mesoparasitism probably appeared independently at least twice in Ergasilidae given the non-homology of the necks (i.e. pre-oral neck in *Mugilicola* Tripathi, 1960 and *Paeonodes* Wilson C.B., 1944 vs post-oral neck in *Majalincola* Tang and Kalman, 2008 and *Therodamas*). *Therodamas longicollum* Oliveira *et al.*, 2021 is the only representative of the mesoparasitic ergasilids whose ribosomal sequences are available (unique sequence for 18S rDNA). In its original description, the authors recovered *T. longicollum* as the sister taxon of all other ergasilids with no close relationship with *A. margulisae* (i.e. the only Neotropical ergasilid with sequences available at that time). Our BI analyses also recovered this same basal position of *T. longicollum*. Interestingly, concerning the morphology of the swimming legs, all *Therodamas* species have the L1 with a 2-segmented enp, which is a common (and unique) diagnostic feature for all Neotropical species. On the other hand, the other 3 mesoparasitic genera, including *Majalincola* who shares the post-oral neck with *Therodamas*, remain with the L1 enp-3 with 3 segments. More

data are necessary for a better understanding of the relationships between *Therodamas* and the other Neotropical ergasilids (or even among the 4 mesoparasitic genera). Current findings may indicate that (at least) 2 distinct lineages have colonized South America independently.

Overall, the clades found in our studies are consistent with those presented in previous phylogenetic analyses. The 5 well-supported clades observed herein can act as interesting targets for future revisions within this family, which would help to minimize big taxonomic problems like the 'elephant in the room' caused by the paraphyletic status of *Ergasilus*. However, more studies combining morphological and molecular analyses are still needed in order to achieve a better understanding of the evolutionary relationships within Ergasilidae – especially in the Neotropical region, which despite having the greatest known diversity of ergasilids in the world still has the least amount of available molecular sequences.

New records and host switching

Rhinergasilus piranhus was originally proposed based on copepods found inside the nostrils of the 'red piranha' *P. nattereri* Kner, 1858 (= *Serrasalmus nattereri*) from the Amazon region, Brazil. Since its description, this species has been continuously reported from the nostrils of different fishes across the country [see Table 4 in Narciso *et al.* (2020)]. A similar story also applies to *G. schizodontis* which was also found in the nostrils of an Amazonian fish, the anostomid *S. fasciatus* Spix and Agassiz, 1829 [see Table 5 in Narciso and Silva (2020)]. Recently, Narciso (2020), when evaluating the ergasilid fauna of fishes from 2 rivers in São Paulo State (Veados and Paranapanema rivers), reported *Rhinergasilus* sp.1 parasitizing the nostrils of *Psalidodon fasciatus* (Cuvier, 1815). Morphological analysis of the latter specimens together with comparisons with our copepods allowed us to identify these specimens as *R. piranhus*. Our study reaffirms the presence of *R. piranhus* and *G. schizodontis* in São Paulo (i.e. corroborating Narciso, 2020; and Narciso and Silva, 2020) and expands their distribution to a new river. Furthermore, it also broadens the list of fish hosts for both species – i.e. 5 new hosts for *R. piranhus* (i.e. including *P. fasciatus* reported by Narciso, 2020) and 1 for *G. schizodontis*.

The occurrence of *R. piranhus* in the nostrils of *Cy. modestus* and *Co. rendalli* expands the number of taxa (beyond species level) that act as hosts for this copepod: (i) 2 new families – Curimatidae Gill, 1858 and Cichlidae Bonaparte, 1835; and (ii) 1 new order – Cichliformes. The low parasitological indices observed for both hosts may characterize this occurrence as an erratic parasitism (i.e. when the parasite is found outside its usual host) (see Table 5). Separating each fish into individual plastic bags prevented those occurrences happened as a result of cross-infection. In contrast to other fish ectoparasites (mainly monogeneans), ergasilids tend to have a lower specificity regarding their hosts (Boxshall and Halsey, 2004). Even though it may be erratic, the occurrence of this copepod (if maintained) can give rise to new lineages that may undergo speciation in the future (Araujo *et al.*, 2015). The occurrence of *R. piranhus* in *Co. rendalli*, which is an exotic fish, may be evidence of host switching since this parasite has never been reported outside the country.

This latter mechanism has already been observed in other ergasilids [e.g. evidence of *Sinergasilus major* (Markevich, 1940) switching hosts in Hungary – see Dos Santos *et al.*, 2021] and the acquisition of native parasites by exotic species is a well-documented phenomenon in Ichthyoparasitology (Paterson *et al.*, 2012, 2013). Like the introduction of alien parasites, the acquisition of native parasites by introduced hosts can also be a problem for the local ecosystem by altering the native

host–parasite dynamics, resulting in spillback or even dilution of the native infection (Kelly *et al.*, 2009). However, more studies are needed to confirm this parasite–host relationship and even the possible impacts of this association on the local fauna of Pardo River.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S003118202400129X>.

Data availability statement. Sequence data are available on the NCBI GenBank database. All other necessary data are included in the article and its supplementary materials.

Acknowledgements. We thank the following agencies for the financial support to this study: Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) and the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq). We thank Eduardo Meneguzzi Brambilla for his help in identifying the fish. Finally, we also thank Linda van der Spuy, North-West University for her help with the scanning electron microscopy analyses performed in this study.

Author contributions. All authors contributed to the study's conception and design. Material preparation, data collection and analysis were performed by R. B. N. The first version of the manuscript was written by R. B. N. and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Financial support. We thank the following agencies for the financial support to this study: Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP – #2020/05412-9; and #2019/26831-2), and the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq – 311635-2021-0). Molecular work at the North-West University Water Research Group was partially funded by the National Research Foundation (NRF) of South Africa (grant number 138573). Opinions, findings, conclusions and recommendations expressed are those of the authors, and the NRF accepts no liability whatsoever in this regard.

Competing interests. None.

Ethical standards. All applicable international, national and institutional guidelines for the care and use of animals were followed. Fish sampling was authorized by the Instituto Chico Mendes de Conservação da Biodiversidade – ICMBio and Sistema de Autorização e Informação em Biodiversidade – SISBIO # 60640-1 and all procedures followed the recommendations of the Ethical Commission for Animal Experimentation from the São Paulo State University (Unesp), Institute of Biosciences, Botucatu, Brazil (CEUA no 9415260520).

References

- Abdel-Radi S, Rashad MM, Ali GE, Eissa AE, Abdelsalam M and Abou-Okada M (2022) Molecular characterization and phylogenetic analysis of parasitic copepoda; *Ergasilus sieboldi* isolated from cultured gilthead sea bream (*Sparus aurata*) in Egypt, associated with analysis of oxidative stress biomarkers. *Journal of Parasitic Diseases* **46**, 1080–1089.
- Abou-Okada M, Rashad MM, Ali GE, Abdel-Radi S and Hassan A (2023) Oxidative stress, gene expression and histopathology of cultured gilthead sea bream (*Sparus aurata*) naturally co-infected with *Ergasilus sieboldi* and *Vibrio alginolyticus*. *BMC Veterinary Research* **19**, 277.
- Alston S, Boxshall GA and Lewis JW (1993) A redescription of adult females of *Ergasilus briani* Markewitsch, 1933 (Copepoda: Poecilostomatoida). *Systematic Parasitology* **24**, 217–227.
- Altschul SF, Gish W, Miller W, Myers EW and Lipman DJ (1990) Basic local alignment search tool. *Journal of Molecular Biology* **215**, 403–410.
- Amado MAPM, Ho J and Rocha CEF (1995) Phylogeny and biogeography of the Ergasilidae (Copepoda, Poecilostomatoida), with reconsideration of the taxonomic status of the Vaigamidae. *Contributions to Zoology* **65**, 233–243.
- Araujo SB, Braga MP, Brooks DR, Agosta SJ, Hoberg EP, Von Hartenthal FW and Boeger WA (2015) Understanding host-switching by ecological fitting. *PLoS ONE* **10**, e0139225.
- Bernot JP, Boxshall GA and Crandall KA (2021) A synthesis tree of the Copepoda: integrating phylogenetic and taxonomic data reveals multiple origins of parasitism. *PeerJ* **9**, e12034.
- Boeger WA and Thatcher VE (1988) *Rhinergasilus piranhus* gen. et sp. n. (Copepoda, Poecilostomatoida, Ergasilidae) from the nasal cavities of piranha caju, *Sermsalmus nattereri*, in the Central Amazon. *Proceedings of the Helminthological Society of Washington* **55**, 87–90.
- Boxshall GA (2016) A new species of *Ergasilus* von Nordmann, 1832 (Copepoda: Cyclopoida) from the gills of a dasyatid ray, *Himantura oxyrhyncha* (Sauvage, 1878) from West Kalimantan, Indonesia. *Zootaxa* **4174**, 93–103.
- Boxshall GA and Defaye D (2008) Global diversity of copepods (Crustacea: Copepoda) in freshwater. *Hydrobiologia* **595**, 195–207.
- Boxshall GA and Halsey SH (2004) *An Introduction to Copepod Diversity*, 2nd Edn. Andover, UK: Ray Society.
- Boxshall G and Hayes P (2019) Biodiversity and taxonomy of the parasitic Crustacea. In Smit N, Bruce N and Hadfield K (eds), *Parasitic Crustacea*. Zoological Monographs, Vol. 3. Cham, Switzerland: Springer, pp. 73–134.
- Bush AO, Lafferty KD, Lotz JM and Shostak AW (1997) Parasitology meets ecology on its own terms: Margolis, et al. Revisited. *The Journal of Parasitology* **83**, 575–583.
- Castro-Romero R, Montes MM, Martorelli SR, Sepulveda D, Tapia S and Martínez-Aquino A (2016) Integrative taxonomy of *Peniculus*, *Metapeniculus*, and *Trifur* (Siphonostomatoida: Pennellidae), copepod parasites of marine fishes from Chile: species delimitation analyses using DNA barcoding and morphological evidence. *Systematics and Biodiversity* **14**, 466–483.
- Chernysheva NB and Purasjoki KJ (1991) A redescription of *Paraergasilus rylovi* Markevich, 1937 (Copepoda, Ergasilidae). *Systematic Parasitology* **20**, 165–171.
- Couto JV, Nazaré Pereira A, Luque JL, Paschoal F and Pereira FB (2023) Two new species of *Acusicola* Cressey, 1970 (Copepoda: Cyclopoida: Ergasilidae) parasitic on the gills of two estuarine actinopterygians off Brazil. *Systematic Parasitology* **100**, 133–148.
- Damborenea C, Rogers DC and Thorp JH (2020) *Thorp and Covich's Freshwater Invertebrates: Volume 5: Keys to Neotropical and Antarctic Fauna*. Amsterdam, Netherlands: Academic Press.
- Darriba D, Taboada GL, Doallo R and Posada D (2012) Jmodeltest 2: more models, new heuristics and high-performance computing. *Nature Methods* **9**, 772.
- Dos Santos QM, Avenant-Oldewage A, Piasecki W, Molnár K, Sellyei B and Székely C (2021) An alien parasite affects local fauna – confirmation of *Sinergasilus major* (Copepoda: Ergasilidae) switching hosts and infecting native *Silurus glanis* (Actinopterygii: Siluridae) in Hungary. *International Journal for Parasitology: Parasites and Wildlife* **15**, 127–131.
- El-Rashidy HH (1999) *Ergasilid Copepods and Grey Mullet* (PhD thesis). Queen Mary University of London, London, UK.
- El-Rashidy H and Boxshall GA (2001a) Biogeography and phylogeny of *Paraergasilus* Markevich, 1937 (Copepoda: Ergasilidae) with descriptions of two new species from the gills of grey mullet. *Journal of Natural History* **35**, 1807–1819.
- El-Rashidy H and Boxshall GA (2001b) The mesoparasitic genera of the Ergasilidae (Copepoda): with descriptions of new species of *Paenodes* Wilson and *Therodamas* Krøyer. *Systematic Parasitology* **50**, 199–217.
- Elías-Gutiérrez M, Valdez-Moreno M, Topan J, Young MR and Cohuo-Colli JA (2018) Improved protocols to accelerate the assembly of DNA barcode reference libraries for freshwater zooplankton. *Ecology and Evolution* **8**, 3002–3018.
- Elsner NO, Jacobsen S, Thielges DW and Reise K (2011) Alien parasitic copepods in mussels and oysters of the Wadden Sea. *Helgoland Marine Research* **65**, 299–307.
- Fikiye PP, Smit NJ, Van As LL, Truter M and Hadfield KA (2023) Integrative morphological and genetic characterisation of the fish parasitic copepod *Ergasilus mirabilis* Oldewage & van As, 1987: insights into host specificity and distribution in southern Africa. *Diversity* **15**, 965.
- Folmer O, Black M, Hoeh W, Lutz R and Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* **3**, 294–299.
- Fricke R, Eschmeyer WN and Van der Laan R (2024) Eschmeyer's catalog of fishes: genera, species, references. Retrieved from California Academy of Sciences website: <https://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp> target="_blank"> <https://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp> (Accessed 2 September 2024).
- Hua CJ, Huang Y, Li HQ, Feng JM, Qiao JL, Hu S, Liu W and Shu XJ (2024) Characterization of the complete mitochondrial genome of *Ergasilus anchoratus* Markevich, 1946 (Ergasilidae) and phylogeny of Copepoda. *Systematic Parasitology* **101**, 33.

- Humes AG and Gooding RU** (1964) A method for studying the external anatomy of copepods. *Crustaceana* **6**, 238–240.
- Huys R, Llewellyn-Hughes J, Olson PD and Nagasawa K** (2006) Small sub-unit rDNA and Bayesian inference reveal *Pectenophilus ornatus* (Copepoda *incertae sedis*) as highly transformed Mytilicolidae, and support assignment of Chondracanthidae and Xarifiidae to Lichomolgoidea (Cyclopoida). *Biological Journal of the Linnean Society* **87**, 403–425.
- Johnson SC, Treasurer JW, Bravo S, Nagasawa K and Kabata Z** (2004) A review of the impact of parasitic copepods on marine aquaculture. *Zoological Studies* **43**, 229–243.
- Katoh K, Misawa K, Kuma KI and Miyata T** (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* **30**, 3059–3066.
- Kelly DW, Paterson RA, Townsend CR, Poulin R and Tompkins DM** (2009) Parasite spillback: a neglected concept in invasion ecology? *Ecology* **90**, 2047–2056.
- Khodami S, Mercado-Salas NF, Tang D and Arbizu PM** (2019) Molecular evidence for the retention of the Thaumatosyllidae in the order Cyclopoida (Copepoda) and establishment of four suborders and two families within the Cyclopoida. *Molecular Phylogenetics and Evolution* **138**, 43–52.
- Kim IH and Nagasawa K** (2006) Redescription of *Ergasilus zacconis* (Copepoda: Poesilostomatoida: Ergasilidae), parasitic on the freshwater fish *Zacco platypus* from Japan. *Korean Journal of Systematic Zoology* **22**, 121–125.
- Kumar S, Stecher G and Tamura K** (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* **33**, 1870–1874.
- Kvach Y, Tkachenko MY, Seifertová M and Ondračková M** (2021) Insights into the diversity, distribution and phylogeny of three ergasilid copepods (Hexanauplia: Ergasilidae) in lentic water bodies of the Morava River basin, Czech Republic. *Limnologia* **91**, 125922.
- Lacerda ACF, Takemoto RM, Lizama MDLAP and Pavanelli GC** (2007) Parasitic copepods in the nasal fossae of five fish species (Characiformes) from the upper Paraná River floodplain, Paraná, Brazil. *Acta Scientiarum. Biological Sciences* **29**, 429–435.
- Luque JL, Pavanelli G, Vieira F, Takemoto R and Eiras J** (2013) Checklist of Crustacea parasitizing fishes from Brazil. *Check List* **9**, 1449–1470.
- Markevich AP** (1933) Descrizione di due species nuove di *Ergasilus* provenienti dalla Russia (URSS). *Memorie della Società Entomologica Italiana* **12**, 129–141.
- Marques TM and Boeger WA** (2018) Proposal of *Tiddergasilus* gen. nov. (Ergasilidae: Cyclopoida) for *T. iheringi* comb. nov. from the gills of *Hoplias malabaricus* (Erythrinidae: Characiformes) from Brazil. *Zoologia (Curitiba)* **35**, e21577.
- Marques TM, Boeger WA and Carvalho Brasil-Sato M** (2015) Two new species of *Ergasilus* Nordmann, 1832 (Copepoda: Ergasilidae) and a redescription of *Ergasilus salmini* Thatcher & Brazil-Sato, 2008 from *Salminus brasiliensis* Cuvier and *S. franciscanus* Lima & Britsky (Teleostei: Characidae) in Brazil. *Systematic Parasitology* **90**, 81–89.
- Míč R, Řehulková E and Seifertová M** (2023) Species of *Ergasilus* von Nordmann, 1832 (Copepoda: Ergasilidae) from cichlid fishes in Lake Tanganyika. *Parasitology* **150**, 579–598.
- Míč R, Řehulková E, Šimková A, Razanabolana JR and Seifertová M** (2024) New species of *Dermoergasilus* Ho & Do, 1982 (Copepoda: Cyclopoida: Ergasilidae) parasitizing endemic cichlid *Paretroplus polyactis* (Bleeker) in Madagascar. *Parasitology* **151**, 1–18.
- Miller MA, Pfeiffer W and Schwartz T** (2010) Creating the CIPRES science gateway for inference of large phylogenetic trees. 2010 Gateway Computing Environments Workshop (GCE), New Orleans, LA, 14 November 2010, pp. 1–8.
- Narciso RB** (2020) *Taxonomia e diversidade de espécies de ergasilídeos parasitas de peixes de água doce dos tributários e lagoas do reservatório de Jurumirim, SP* (MSc thesis). Universidade Estadual Paulista ‘Júlio de Mesquita Filho’ – UNESP, Botucatu, Brazil.
- Narciso RB and Silva RJ** (2020) Two *Gamispatus* Thatcher & Boeger, 1984 (Cyclopoida: Ergasilidae) from *Schizodon intermedius* Garavello & Britski (Actinopterygii: Anostomidae), with description of a new species. *Zootaxa* **4803**, 463–482.
- Narciso RB, Brandão H, Perbiche-Neves G and Silva RJ** (2020) A new species of *Rhinergasilus* Boeger et Thatcher, 1988 (Copepoda: Ergasilidae) from gills of *Astyanax fasciatus* (Cuvier, 1819) (Actinopterygii: Characidae). *Acta Parasitologica* **65**, 327–334.
- Narciso RB, Perbiche-Neves G and Silva RJ** (2021) *Rhinergasilus unguilongus* n. sp. (Copepoda: Ergasilidae): a gill parasite of the freshwater fish *Prochilodus lineatus* (Valenciennes, 1837) (Actinopterygii: Prochilodontidae) from the neotropical region, Brazil. *Acta Parasitologica* **66**, 155–162.
- Narciso RB, Vieira DHMD and Silva RJ** (2022) A new species of *Tiddergasilus* Marques & Boeger, 2018 (Copepoda: Ergasilidae) from the gills of *Astyanax lacustris* (Lütken) (Osteichthyes: Characidae) in Brazil. *Systematic Parasitology* **99**, 671–681.
- Oliveira MS, Corrêa LL, Adriano EA and Tavares-Dias M** (2021) Integrative taxonomy of a new species of *Therodamas* (Ergasilidae) infecting the Amazonian freshwater fish *Leporinus fasciatus* (Anostomidae). *Parasitology Research* **120**, 3137–3147.
- Ondračková M, Fojtů J, Seifertová M, Kvach Y and Jurajda P** (2019) Non-native parasitic copepod *Neoergasilus japonicus* (Harada, 1930) utilizes non-native fish host *Lepomis gibbosus* (L.) in the floodplain of the River Dyje (Danube basin). *Parasitology Research* **118**, 57–62.
- Ota RR, Deprá GDC, Graça WJD and Pavanelli CS** (2018) Peixes da planície de inundação do alto rio Paraná e áreas adjacentes: revised, annotated and updated. *Neotropical Ichthyology* **16**, e170094.
- Paterson RA, Townsend CR, Tompkins DM and Poulin R** (2012) Ecological determinants of parasite acquisition by exotic fish species. *Oikos* **121**, 1889–1895.
- Paterson RA, Rauque CA, Fernandez MV, Townsend CR, Poulin R and Tompkins DM** (2013) Native fish avoid parasite spillback from multiple exotic hosts: consequences of host density and parasite competency. *Biological Invasions* **15**, 2205–2218.
- Piasecki W, Goodwin AE, Eiras JC and Nowak BF** (2004) Importance of Copepoda in freshwater aquaculture. *Zoological Studies* **43**, 193–205.
- Rambaut A** (2012) *FigTree* v1. 4. Retrieved from Molecular Evolution, Phylogenetics and Epidemiology website: `##a href="http://tree.bio.ed.ac.uk/software/figtree/" target="_blank"> http://tree.bio.ed.ac.uk/software/figtree/` (Accessed 2 January 2024).
- Ribeiro DC, Chagas JMA, Thereza MR and Langeani F** (2019) Checklist and key for the identification of fish fauna of the Uberaba River, upper Paraná River system, Brazil. *ZooKeys* **875**, 129–155.
- Rindoria NM, Gichana Z, Morara GN, van Wyk C, Smit WJ, Smit NJ and Luus-Powell WJ** (2023) Scanning electron microscopy and first molecular data of two species of *Lamproglana* (Copepoda: lernaecidae) from *Labeo victorianus* (Cyprinidae) and *Clarias gariepinus* (Clariidae) in Kenya. *Pathogens* **12**, 980.
- Roberts LS** (1965) *Ergasilus tenax* sp. n. (Copepoda: Cyclopoida) from the white crappie, *Pomoxis annularis* Rafinesque. *The Journal of Parasitology* **51**, 987–989.
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA and Huelsenbeck JP** (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* **61**, 539–542.
- Rosim DF, Boxshall GA and Ceccarelli PS** (2013) A novel microhabitat for parasitic copepods: a new genus of Ergasilidae (Copepoda: Cyclopoida) from the urinary bladder of a freshwater fish. *Parasitology International* **62**, 347–354.
- Santacruz A, Morales-Serna FN, Leal-Cardín M, Barluenga M and Pérez-Ponce de León G** (2020) *Acusicola margulisiae* n. sp. (Copepoda: Ergasilidae) from freshwater fishes in a Nicaraguan crater lake based on morphological and molecular evidence. *Systematic Parasitology* **97**, 165–177.
- Song Y, Wang GT, Yao WJ, Gao Q and Nie P** (2008) Phylogeny of freshwater parasitic copepods in the Ergasilidae (Copepoda: Poecilostomatoida) based on 18S and 28S rDNA sequences. *Parasitology Research* **102**, 299–306.
- Stamatakis A** (2014) RAXML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**, 1312–1313.
- Suárez-Morales E, Reid JW, Fiers F and Iliffe TM** (2004) Historical biogeography and distribution of the freshwater cyclopine copepods (Copepoda, Cyclopoida, Cyclopinae) of the Yucatan Peninsula, Mexico. *Journal of Biogeography* **31**, 1051–1063.
- Tang D and Kalman JE** (2008) A new genus and species of mesoparasitic ergasilid (Copepoda: Cyclopoida) from brackish water pufferfishes collected in northern Australian waters. *Systematic Parasitology* **69**, 89–99.
- Thatcher VE** (1998) Copepods and fishes in the Brazilian Amazon. *Journal of Marine Systems* **15**, 97–112.
- Thatcher VE** (2006) *Amazon Fish Parasites*, 2nd Edn, Sófia, Bulgaria: Pensoft Publishers.

- Thatcher VE and Boeger WA** (1984) The parasitic crustaceans of fishes from the Brazilian Amazon, 15., *Gamispatulus schizodontis* gen. et sp. nov. (Copepoda: Poecilostomatoida: Vaigamidae) from the nasal fossae of *Schizodon fasciatus* Agassiz. *Amazoniana* **9**, 119–126.
- Thatcher VE and Robertson BA** (1984) The parasitic crustaceans of fishes from the Brazilian Amazon. 11. Vaigamidae fam. nov. (Copepoda: Poecilostomatoida) with males and females of *Vaigamus retrobarbatus* gen. et sp. nov. and *V. spinicephalus* sp. nov. from plankton. *Canadian Journal of Zoology* **62**, 716–729.
- van der Sleen P and Albert J** (2017) *Field Guide to the Fishes of the Amazon, Orinoco, and Guianas*. Princeton, USA: Princeton University Press.
- Vasquez AA, Bonnici BL, Kashian DR, Trejo-Martinez J, Miller CJ and Ram JL** (2022) The biodiversity of freshwater Crustaceans revealed by taxonomy and mitochondrial DNA barcodes. In Zupo V (ed.), *Crustaceans: Endocrinology, Biology and Aquaculture*. Boca Raton, USA: CRC Press, pp. 166–181.
- Waicheim MA, Marques TM, Rauque CA and Viozzi G** (2021) New species of *Ergasilus* von Nordmann, 1832 (Copepoda: Ergasilidae) from the gills of freshwater fishes in Patagonia, Argentina. *Systematic Parasitology* **98**, 131–139.
- Yamaguti S** (1936) *Parasitic copepods from fishes of Japan, Part 1. Cyclopoida, I*. Kyoto Imperial University. Published by author.