



Nutrition Society Live 2020 was held virtually on 14–15 July 2020

Symposium three: Physiological determinants for protein requirements

Impact of protein on the composition and metabolism of the human gut microbiota and health

Sylvia H. Duncan* , Ajay Iyer and Wendy R. Russell

Gut Health Group, Rowett Institute, University of Aberdeen, Foresterhill, Aberdeen AB25 2ZD, Scotland, UK

The composition and metabolic activity of the bacteria that inhabit the large intestine can have a major impact on health. Despite considerable inter-individual variation across bacterial species, the dominant phyla are generally highly conserved. There are several exogenous and gut environmental factors that play a role in modulating the composition and activities of colonic bacteria including diet with intakes of different macronutrients, including protein, accounting for approximately 20% of the microbial variation. Certain bacterial species tend to be considered as generalists and can metabolise a broad range of substrates, including both carbohydrate- and protein-derived substrates, whilst other species are specialists with a rather limited metabolic capacity. Metabolism of peptides and amino acids by gut bacteria can result in the formation of a wide range of metabolites several of which are considered deleterious to health including nitrosamines, heterocyclic amines and hydrogen sulphide as some of these products are genotoxic and have been linked to colonic disease. Beneficial metabolites however include SCFA and certain species can use amino acids to form butyrate which is the major energy source for colonocytes. The impact on health may however depend on the source of these products. In this review, we consider the impact of diet, particularly protein diets, on modulating the composition of the gut microbiota and likely health consequences and the potential impact of climate change and food security.

Macronutrients: Protein: Carbohydrates: Gut microbiota: SCFA: Amino acids

The human colon is an anoxic and dynamic environment which constantly interacts with the host's immune system. The colon harbours a dense collection of bacteria that inhabit the large intestine that mainly belongs to five different phyla. These phyla comprise many hundreds of different species and most of these bacteria are anaerobes. Given the advances in the molecular methods available to profile the gut microbiota⁽¹⁾, there is currently a good understanding of composition. Moreover, affordable complete genome sequencing of gut bacteria has meant that it is feasible to mine the genomes of many human gut bacterial species for traits of interest. For example, certain bacterial species, often with large

genomes, such as some *Bacteroides* species, are considered generalists as these have a remarkable repertoire of enzymes⁽²⁾ which allows these bacterial cells to metabolise a broad range of substrates as carbon, nitrogen and energy sources.

Numerous factors influence gut microbial composition such as host genetics, general health status, exposure to microbes during early life and consumption of antibiotics. Diet however is a major contributor to microbial structure and the main dietary macronutrients: carbohydrate, proteins and fat will influence gut microbial activities and metabolic outputs. Also, the types of foods consumed, cooking processes used and the balance of

Abbreviation: HMO, human milk oligosaccharides.

***Corresponding author:** Sylvia H. Duncan, email Sylvia.duncan@abdn.ac.uk

macronutrients and micronutrients are likely to be important drivers of health. The major products of fermentation include the SCFA and butyrate, in particular, has a special role for the host and it is the major energy source for colonocytes⁽³⁾. Other bacterial metabolites formed mainly from the metabolism of proteins can result in the formation of less beneficial products including N-nitrosamines and heterocyclic amines that can be deleterious to health⁽⁴⁾. The overall balance of benefit and detriment for the host will therefore depend on the status of the microbial community in terms of its distribution, diversity, species composition and metabolic outputs⁽⁵⁾.

Increasingly, there are considerable concerns about the types of foods we eat and the global impact of both climate change and the SARS-Cov-2 pandemic has highlighted the urgency to maintain food security. Moreover, this pandemic has highlighted the need for many countries, including the UK, to consider more sustainable food chains and reducing transportation of imported foods. The UK currently imports about half the food consumed. Consideration therefore needs to be given to agricultural practices and food production systems with an additional aim of reducing greenhouse gas emissions⁽⁶⁾. The food we eat impacts on the overall metabolic activities of colonic microbes and dietary changes will have an impact on health and disease. In this review, we consider the complex relationship between diet, particularly protein content, and the gut microbiota and its metabolism and how this may impact on health whilst outlining the impact of different protein diets on the environment.

The human gut microbial ecosystem

The human large intestine performs several key functions including degradation of dietary substrates, nutrient absorption, excretion of waste and is the major site of salt and water absorption. Other than diet, the composition of the gut microbiota may be influenced by many external factors including host genetics which has been estimated to explain approximately 9% of the variation⁽⁷⁾. Other factors that impact on microbial variation may include age, geographical location and antibiotic usage⁽⁸⁾.

Moreover, some host factors may also impact on gut microbial composition including gut transit which can vary. The mean gut transit time across the complete length of the intestinal tract in healthy adults has been estimated to be between 26 and 35 h^(9,10), but can be up to several days thereby allowing for the establishment of an abundance of microorganisms⁽¹¹⁾. There are many discrete physiological environments within the human gastrointestinal tract, which includes the highly acidic conditions in the stomach⁽¹⁰⁾, to the more alkaline pH in the small intestine with changes in pH along the large intestine.

Importantly, diet is thought to explain approximately 20% of the variation in gut microbial composition⁽¹²⁾. Undigested dietary material that escapes digestion by

host enzymes enters the colon and is rapidly fermented by the resident microbiota. This results in rapid microbial growth and production of SCFA and other metabolites, which in turn lowers the pH⁽¹³⁾. As the digesta moves towards the distal colon, carbohydrate sources become depleted; therefore, microbial growth and carbohydrate fermentation decrease whilst peptide fermentation increase, depending on dietary intake, resulting in the formation of a range of nitrogenous products including ammonia which is one of the products that drives an increase in pH towards neutrality⁽¹³⁾.

Colonic microbial composition

The human large intestine harbours viruses many of which are bacteriophages^(14,15), fungi including the dimorphic *Candida* species⁽¹⁶⁾ and bacteria. The latter include thousands of different bacterial species that reach their highest density in the large intestine. The composition and metabolic activities of these microbes are likely to be strongly influenced by diet which will impact the health and disease^(5,12).

The adult microbial community usually contains about 10¹¹ bacterial cells/g of faeces⁽¹⁷⁾. The gut microbiome also contains many more genes (approximately 150-times more) than the human genome, which is currently estimated to possess around 24 000 genes, providing the host with greatly expanded functionality, particularly with regard to complex carbohydrate metabolism⁽¹⁸⁾, and although gut bacterial composition can be decidedly variable, functionally it is somewhat more highly conserved⁽¹⁹⁾.

Despite inter-individual variability at the genus and species level, in the composition of the gut microbiota, there are core species that are found in most healthy individuals. At the phylum level, Firmicutes and Bacteroidetes are the most dominant. The less abundant phyla are the Proteobacteria, Actinobacteria and Verrucomicrobia⁽²⁰⁾. A key species of the latter is the mucin-degrading bacterium *Akkermansia muciniphila*⁽²¹⁾ which is considered to be health protective.

Bacteroidetes usually comprise about 30% of the total microbiota, although this can vary, and it is becoming increasingly apparent that there is a divergence with some individuals tending to be either *Bacteroides* or *Prevotella* dominant⁽⁸⁾. *Bacteroides* and *Prevotella* species can utilise carbohydrate- or protein-derived substrates⁽²²⁾. Commonly occurring species include *B. vulgatus*, *B. fragilis*, *B. distasonis*, *B. uniformis*, *B. thetaiotaomicron* and *B. eggerthii*⁽²³⁾.

The Firmicutes are members of the clostridia class and the predominant human colonic species mainly belong to two phylogenetic groups. One group is the Lachnospiraceae that includes genera such as *Eubacterium*, *Roseburia*, *Butyrivibrio*, *Coproccoccus* and *Lachnospira* and the second is the Ruminococcaceae that encompasses *Faecalibacterium* and *Ruminococcus* species. Other commonly reported genera found in lower abundance include *Bifidobacterium* and *Veillonella* species^(20,24,25).

Facultative anaerobes are usually much less dominant in the healthy colon but their abundance may be elevated in certain diseases⁽²⁶⁾ and includes Enterobacteriaceae species. A number of other, specialised groups may exist at lower levels including the sulphate-reducing bacterial species *Desulfovibrio*⁽²⁷⁾. Proteobacteria are usually in low abundance in the healthy gut but are often more prevalent in frail elderly⁽²⁸⁾. Archaeal methanogens may also be present in approximately 50% of adults with *Methanobrevibacter smithii* as the predominant species⁽²⁹⁾ and methane, which is a major end product, may slow gut transit.

Role of gut environmental factors including anaerobiosis, pH and bile on microbiota composition

There are many factors that are likely to impact on the composition of the gut microbiota. This includes host factors as well as gut environmental factors such as anaerobicity, pH and bile salt levels.

Anaerobic ecosystem

The two dominant phyla that inhabit the large intestine are the Firmicutes and Bacteroidetes and their niche in the colon may partly be driven by redox potential (Eh) and gas phase. Gut microorganisms persist in an environment with low partial oxygen pressures and this anaerobic ecosystem has an Eh value of about -250 mV. Anaerobes generally lack electron transport chains found in facultative anaerobic bacteria to regenerate the reduced cofactors and therefore do not gain further energy by electron transport-level phosphorylation. Instead, metabolic intermediates are reduced mainly to acidic fermentation products and gases. Some gut bacteria including Proteobacteria perform anaerobic respiration involving electron transport chains by using electron acceptors such as sulphate or carbon dioxide⁽²⁰⁾.

Bacteroides species have been described as anaerobes as many species can survive for several hours in the presence of oxygen but require anoxic conditions to grow. By comparison, many gut bacterial species belonging to the Firmicutes are considered as strict anaerobes and are unable to survive for even a few minutes upon exposure to air⁽⁵⁾. Interestingly, one of the most abundant Firmicutes species in the large intestine, *Faecalibacterium prausnitzii*, has adapted to using an electron shuttle of thiols and flavins to transfer electrons to oxygen⁽³⁰⁾.

Gastrointestinal pH

As much of the metabolism of gut anaerobes is given over to fermenting dietary macronutrients, particularly non-digestible carbohydrates to SCFA, the pH of the proximal colon which is the most active site of fermentation in healthy subjects is mildly acidic (about pH 5.5–6.0). There are usually less carbohydrates available in the distal colon; bacteria that are resident in this section are also reliant on the metabolism of peptides and amino acids as sources of carbon and nitrogen. This is likely to

result in the formation of higher levels of nitrogenous products including ammonia and will contribute to driving pH values closer to neutrality. Many of the Gram-positive Firmicutes species are more tolerant of the mildly acidic conditions in the proximal colon which is likely to provide a competitive advantage for these bacterial species, whereas the growth of Bacteroidetes species are likely to be restricted here but are likely to be more active in the distal colon where the pH is closer to neutrality⁽²⁷⁾. This has been supported by studies that revealed the major changes in species composition and metabolic products when comparing the impact of pH 5.5 and 6.5 in model colonic *in vitro* fermentor systems⁽³¹⁾ whereby mildly acidic conditions (about pH 5.5) is favoured by, for example, butyrate producing *Roseburia* species. When the pH is closer to neutrality (pH 6.5), *Bacteroides* spp. that have a role in peptide metabolism tend to be favoured⁽³²⁾.

Bile salts

Enzymes in the liver convert cholesterol to bile acids which are secreted into the intestine from the gall bladder. More cholesterol is formed when diets are high in saturated fats and consequently the secretion of bile increases when consuming these diets. The bile acids made in the liver are known as primary bile acids, and in human subjects, there are two major types such as cholic acid and deoxycholic acid. Within the liver, these are usually conjugated into two amino acids, either glycine or taurine. The latter is biosynthesised from methionine and cysteine whilst glycine is from serine.

Interestingly, it is the type of dietary macronutrients that largely dictates whether the bile acids are conjugated with glycine which is likely to be largely plant based or alternatively to taurine which is most likely to occur when diets are high in animal protein and fat. When these conjugated bile acids reach the large intestine, certain gut bacterial species that possess bile salt hydrolases can cleave the linkages that bond the bile acids to the amino acids. In the case of taurocholic acid, the deconjugation process results in the release of taurine and choline into the intestine. Certain bacterial species such as *Bilophila wadsworthii* can metabolise taurine to form ammonia, acetate and hydrogen sulphide and the latter product is genotoxic. *Clostridium scindens* can remove the hydroxyl group from choline to form deoxy choline which is a tumour-promoting agent⁽³³⁾. The activities of these bacterial species in the colon are therefore considered likely to contribute to the promotion of colorectal cancer⁽³⁴⁾.

Moreover, habitual intakes of a typical western-style diet which is usually considered to be low in fibre, but high in refined sugar and fat, have been associated with increased levels of endotoxin-producing bacteria such as *Escherichia coli*. This may be due to high fat resulting in increased bile formation and that these species may be more tolerant of bile than other bacteria that are dominant in the large intestine.

Impact of diet on modulating the gut microbiota

Recent major advances in molecular profiling technologies have progressed our understanding of the composition of the gut microbiota and how it changes through life stages from birth⁽³⁵⁾ to the elderly⁽³⁶⁾. In adults, short-term dietary interventions have demonstrated that these shifts occur rapidly⁽³⁷⁾; however, these changes may be transient.

Dietary influences on the gut microbiota of infants, adults and elderly

Diet is a factor that shapes the composition of the gut microbiota across all the different life stages. The gastrointestinal tract of a fetus is usually considered sterile; then following birth, the gut microbiota of babies is likely to depend on the mode of delivery⁽³⁵⁾, bile acids⁽³⁸⁾ and feeding regimen, including whether babies are breast- or formula-fed⁽³⁹⁾. The intestinal tract of breast-fed babies is largely dominated by members of the *Bifidobacterium* genus, which appear to be exquisitely adapted to utilise human milk oligosaccharides (HMO)⁽⁴⁰⁾. Breast milk is a rich source of peptides, from casein and whey, in addition to non-digestible sugars, usually referred to as HMO is likely to drive bifidobacterial population establishment in the colon. HMO are amongst the most abundant components of human milk after water and lactose. These HMO have a degree of polymerisation from three to thirty-two with about fifty of these different carbohydrates present in mother's milk. Major HMO include lacto-N-tetraose, lacto-N-neotetraose and lacto-N-hexaose along with fucosylated molecules. Formula-fed babies, in contrast, usually possess a more complex gut microbiota that is more adult-like in composition⁽⁴¹⁾. The introduction of solid foods at weaning results in completely altered substrate availability in the colon and triggers the expansion of obligate anaerobic bacterial groups such as the Bacteroidetes and Firmicutes, which are able to breakdown and metabolise more complex polysaccharide sources⁽⁴²⁾.

Following weaning and up to about 3 years of age, the microbiota of infants tends to become more diverse with a high rate of microbial instability and therefore this is a crucial period for the development of the gut microbiota which may impact on long-term health. Beyond 3 years of age, the gut microbiota tends to stabilise, although dietary intakes will influence the microbiota composition. Changes in the composition of the gut microbiota may however undergo more prolonged development than previously suspected with the microbial diversity of children having perhaps greater diversity than that of healthy adults.

Despite a tendency for microbial stability in adulthood with habitual diet providing a constant source of nutrients, the gut microbiota is in a constant state of flux. A diverse diet that includes a number of different types of plant foods has been associated with greater bacterial diversity. A positive relationship has been observed between dietary diversity and microbial stability with *F. prausnitzii* being increased in individuals

that consumed more than thirty plant types per week compared to those that consumed less than ten per week⁽⁴³⁾.

In the elderly, previous studies have compared the differences in the microbiota in the elderly within community dwellers to those who are staying in care homes as the latter tend to have more health problems^(44,45). The study found that care home dwellers had a higher proportion of Bacteroidetes than that found in community dwellers. *Roseburia* species that are known butyrate producers were also present in lower abundance in care home residents and *F. prausnitzii* was lower in abundance in frail elderly. Moreover, the elderly population (>65 years old) has been found to have a gut microbiota that is less diverse than in healthy young adults which may be due to the reduction in diet variation and also deterioration in dentition, salivary function and gut transit⁽³⁶⁾.

Ageing may also affect the ileal microbiota, as has been suggested by examinations of the ileal contents of sudden death elderly patients, which revealed that their microbial community contained high proportional abundances of Proteobacteria, *Bacillus*, *Streptococcus* and *Lactobacillus* species when compared to that of adult ileostomy patients which were observed to have a lower proportional abundance of Proteobacteria and higher abundance of species belonging to Firmicutes⁽⁴⁶⁾.

Dietary macronutrients

Diet has a key role in modulating the composition of the human intestinal microbiota which impacts on health. The balance, type and amount of dietary macronutrients, including carbohydrates and protein, can have a major impact on the composition of the intestinal microbiota whilst micronutrient status including vitamin availability, some of which is derived from the certain species of the microbial community, is also important for health. Many of the dominant bacteria that reside in the human colon may be auxotrophic and therefore are unable to synthesise all vitamins required for growth. These species are therefore dependent on the host or other bacteria for certain vitamins to facilitate growth⁽⁴⁷⁾.

Dietary carbohydrates. Complex dietary fibre is the most commonly accepted nutrient to exert a beneficial effect on microbial composition⁽⁵⁾. The main carbohydrate consumed by adults and available for utilisation by intestinal microbes includes resistant starches followed by NSP and oligosaccharides⁽¹³⁾. The amounts of these macronutrients consumed daily can be highly variable with intakes of resistant starch ranging from <10 to >40 g daily. Resistant starch is defined as dietary starch that escapes digestion by host enzymes in the upper gastrointestinal tract because of protection provided by other polymers (RS1), particle structure (RS2), retrogradation (RS3) or chemical cross-linking (RS4). The starch-degrading enzyme systems from human gut symbionts that have been studied in detail for *Bacteroides thetaiotaomicron*, *Eubacterium rectale* and more recently *Ruminococcus bromii*, which is a keystone species for the breakdown

of resistant starch in the human large intestine⁽⁴⁸⁾. Unlike starch, pectins found in plant cell walls are structurally highly complex and are classified into homo-polygalacturonan, rhamnogalacturonan I and rhamnogalacturonan II⁽⁴⁹⁾. Pectin degradation requires glycoside hydrolases, polysaccharide lyases and carbohydrate esterases bacterial activities⁽⁴²⁾. Pectin may be degraded by Gram-negative *Bacteroides* species⁽²³⁾ and a few Gram-positive bacterial species have also been reported to ferment pectin or pectin breakdown products including *Eubacterium eligens*^(26,50) which possesses anti-inflammatory activity by promoting the production of IL-10 by epithelial cells⁽⁵¹⁾.

Edible plants can contain several hundreds of phenolic compounds derived by the phenylpropanoid pathway, and based on their structures, these are classified predominantly into phenolic acids, flavonoids, stilbenes, lignans and tannins. On average about 1 g plant phenolics may be consumed daily depending on dietary intakes⁽⁵²⁾. Phenolic compounds can exert dual effects on the gut microbiota as they can inhibit the growth of specific taxa whilst enhancing the growth of others whereby they can be metabolised into bioavailable substrates for the host. Food rich in phenolics, such as fruit, vegetables, cereals, tea and coffee, is associated with a range of health-promoting activities with a reduced risk of chronic disease⁽⁵³⁾. Aromatic amino acids such as tyrosine and phenylalanine are fermented to further phenolic compounds including cresol and phenol derivatives whilst tryptophan is fermented to indoles⁽⁵⁴⁾. Further studies revealed that two abundant phenylpropanoid-derived compounds found in human faecal samples are phenylacetic acid and 4-hydroxyphenylacetic acid, and although they have the potential to be derived from diets rich in plant-based foods, these compounds can also be derived from the microbial fermentation of aromatic amino acids in the colon and are likely to be a major source of phenylpropanoid-derived metabolites in the colon⁽⁵³⁾.

Dietary protein. Approximately 3–18 g dietary protein enters the human large intestine every day⁽⁵⁴⁾, which is diet-dependent. On a very low protein diet, this can range from 3 to 16 g/d on a vegan diet high in unprocessed cereals and grains. This can increase to 18 g/d on meat-rich diets⁽⁵⁵⁾. High protein and low carbohydrate diets may aid weight loss given their impact on satiety⁽⁵⁶⁾. Undigested protein reaching the large intestine may however lead to an increase of pathogenic microorganisms with an associated higher risk of metabolic diseases. High consumption of red meat, which in addition to being rich in protein, also contains haeme and has been associated with an elevated risk of developing colorectal cancer⁽⁵⁷⁾.

Dietary proteins are hydrolysed into peptides and amino acids by both host- and bacterial-derived proteases and peptidases^(58,59). The released peptides and amino acids can be further utilised by both gut bacteria and the host. Bacterial metabolism of extracellular amino acids is likely however to require specific transporters. Peptide and amino acid-fermenting bacteria include species that belong to the following genera: *Bacteroides*, *Prevotella*, *Clostridium*, *Veillonella*, *Megasphaera*,

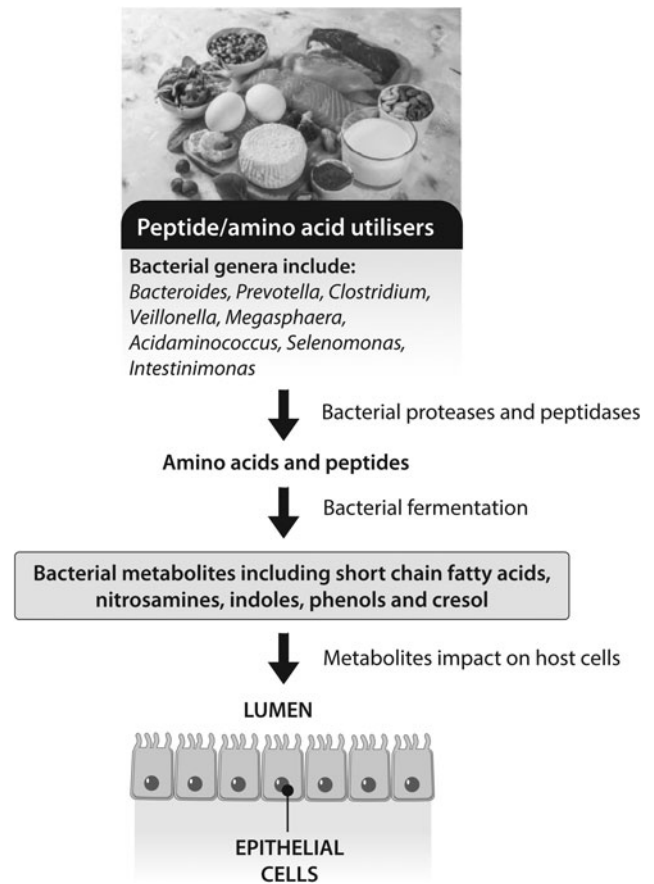


Fig. 1. Protein metabolism by colonic bacteria.

Acidaminococcus and *Selenomonas* (Fig. 1). Certain species possess highly active dipeptidyl peptidase and dipeptidase activities, suggesting that these bacteria might be important for protein digestion and amino absorption in the mammalian digestive tract. Most gut bacteria utilise amino acids and ammonia as their preferred nitrogen source, although for others such as certain *Prevotella* species peptides are the preferred nitrogen source⁽⁶⁰⁾. *Bacteroides* species can secrete proteases with presumed activity near the brush border of absorptive cells and a high abundance of *Bacteroides* species may result in an excess of proteases, which may degrade maltase and sucrase enzymes in the brush borders of enterocytes⁽⁶¹⁾.

The levels of proteins, peptides and amino acids are relatively high in the proximal colon and reduced in the distal colon. Regarding the large intestine, it appears that amino acids are not significantly absorbed by the colonic mucosa, but rather are intensively metabolised by the large intestinal microbiota⁽⁶²⁾. This higher rate of bacterial protein fermentation has been related to high pH and low carbohydrate availability in the large intestine⁽⁶³⁾ resulting in the generation of a complex combination of metabolic end products including SCFA and the major acids in the colon are acetate, propionate and butyrate and the branched-chain fatty acids valerate, isobutyrate and iso-valerate. In addition, microbial metabolism of amino acids will also result in the formation

of ammonia and amines and the latter is produced by decarboxylation of amino acids. The amines mainly produced by the resident microbiota include cadaverine (a decarboxylation product of lysine) and agmatine (a decarboxylation product of arginine)⁽⁶⁴⁾. These amines can have significant physiological effects and agmatine has been shown to influence metabolic functions including elevating tissue cyclic AMP levels, ultimately replicating the effects of energy restriction with respect to metabolic reprogramming and leading to reduced diet-induced weight gain⁽⁶⁵⁾.

Microbial metabolites

Although microbial cells are usually prevented from breaching barriers allowing access to host cells in the large intestine, smaller molecular weight microbial metabolites can cross this barrier by diffusion and active transport. The gut microbiota forms an array of primary and secondary metabolites which can be transported into colonocytes and exert beneficial or deleterious effects on these epithelial cells depending on their concentrations in the lumen. Certain metabolites have been postulated to have a role in a wide range of health conditions, including diabetes, atherosclerosis, kidney disease, inflammatory bowel disease and cancer⁽⁶⁶⁾. The gut anaerobes ferment dietary nutrients to form SCFA which include acetate, propionate, butyrate and gases including carbon dioxide and hydrogen. Some of the weak acidic metabolites including propionate and butyrate are likely to provide health benefits including appetite control, dampen inflammation, maintain gut and systemic health and modulate disease progression. Conversely, lactate which is generally considered as an intermediate fermentation product can result in acidosis unless this product is removed by bacterial cross-feeding⁽⁶⁷⁾. Specialist gut microbial species can release and transform dietary plant phenolics and the spectrum of products formed may provide potent antioxidant and anti-inflammatory activities⁽⁵³⁾. Conversely, consumption of high animal protein and fat diets may lead to the formation of damaging microbial products including elevated levels of nitroso-compounds, hydrogen sulphide and trimethylamine⁽⁴⁾.

SCFA

Microbial fermentation in the large intestine results in the formation of a range of SCFA and the main acids detected in the large intestine are acetate, propionate and butyrate that make up about 90% of acids in the colon and are usually detected in molar proportions of about 3:1:1 but this is dependent on diet and the composition of each individual's microbiota. Some minor SCFA including iso-butyrate and iso-valerate are formed by bacterial fermentation of branched-chain amino acids. The total level of SCFA is usually in the region of 60–180 mM depending on factors such as diet and gut transit^(37,68). The majority of intestinal bacteria use the glycolytic pathway and the pentose phosphate pathway to harvest energy from carbohydrates, both pathways

lead to the formation of pyruvate which is a key intermediate in SCFA formation⁽¹¹⁾. Although acetate reaches the highest concentration of any of the SCFA in faeces, it is known that many human faecal bacteria are net consumers of acetate in pure culture⁽⁶⁹⁾ including the dominant butyrate producers *F. prausnitzii*, *Roseburia* species and *E. rectale*⁽⁷⁰⁾. Butyrate is generally believed to be synthesised via two main routes namely butyrate kinase or butyryl CoA:acetate CoA transferase routes⁽⁷¹⁾. Butyrate is generally considered to provide a number of health benefits and is the preferred energy source for the colonocytes^(3,72,73). Increased levels of butyrate have been associated with increased intestinal transit⁽⁷⁴⁾.

Propionate can stimulate the gut hormones, peptide YY and glucagon-like peptide-1, which increase satiety and thereby reducing energy intake and body weight gain in adults^(75,76). A large group of the gut bacteria can generate propionate including the abundant Bacteroidetes phylum⁽⁷⁷⁾. Propionate can be formed via three different metabolic routes and these are the acrylate, succinate and the propanediol pathways⁽⁷⁸⁾.

Amino acids utilised by gut anaerobes that can be metabolised to acetate include glycine, threonine, glutamate, lysine, ornithine and aspartate⁽⁶¹⁾. Threonine can give rise to all three major SCFA and with propionate mainly being produced from threonine⁽⁶¹⁾. Butyrate can be generated from the metabolism of threonine, glutamate and lysine. The latter can be used by species of *Intestinimonas* to form butyrate⁽⁷⁹⁾. The branched-chain amino acids namely valine, leucine and isoleucine give rise to the formation of the branched-chain fatty acids, iso-leucine, iso-valine and valine as has been reported for *Anaerostignum* species⁽⁸⁰⁾.

Hydrogen sulphide

Hydrogen can be formed by fermentative bacteria in the large intestine and in turn can be consumed by methanogens, acetogens and sulphate-reducing bacteria. These bacteria are likely to compete for hydrogen. The end product of sulphate reduction, hydrogen sulphide can be formed by bacterial species such as *Desulfovibrio piger*⁽²⁷⁾ and can inhibit butyrate metabolism and is therefore highly toxic to the colonic mucosa⁽⁸¹⁾. This bacterial metabolic product can also inhibit colonic smooth muscle contractility⁽⁵⁵⁾. Hydrogen sulphide is produced by fermentation of sulphur-containing amino acids, such as methionine and cysteine which is also derived from the reduction of inorganic sulphate and sulphite additives, and the catabolism of intestinal sulphomucins.

Ammonia and other nitrogenous bacterial metabolites

Peptides and amino acids are metabolised by gut bacteria following deamination and decarboxylation to several metabolites including ammonia, polyamines, phenols and indoles. Ammonia is generally found at millimolar concentrations in the large intestine and concentrations increase from the ascending to the descending colon, which is consistent with a higher rate of protein metabolism in the distal compared to the proximal colon. The

ammonia concentration in the large intestine is mainly a microbial metabolite associated with amino acid deamination and urea hydrolysis. Intestinal microbiota can use ammonia, and ammonia can also be absorbed by the epithelial cells. Urea hydrolysis in the intestinal lumen is performed via bacteria urease activities which are better understood in ruminants than in human subjects⁽⁸²⁾. A reduction in urease activity will result in a reduction in blood ammonia levels which is beneficial to health as high levels of ammonia have been linked to encephalopathy⁽⁸³⁾. Nitrosamines are known carcinogens and can be detected in human faeces. Gastric formation of nitrosamines has been well described in human subjects and the involvement of the microbiota has been demonstrated by comparing germ-free and conventional rats⁽⁸⁴⁾. Several bacterial species are capable of nitrosamine production including species belonging to Proteobacteria^(85,86).

The bacterial deamination of aromatic amino acids leads to the production of phenolic compounds and tyrosine deamination mainly yields phenol and *p*-cresol. The main food sources of tyrosine are egg, cod, seaweed and cheese and an increase of the nutritional protein load in healthy individuals principally results in greater urinary excretion of *p*-cresol. An example of possible health impacts is that the tryptophan metabolite indole-3-propionic acid, which has been shown to be a potent anti-non-alcoholic steatohepatitis microbial metabolite in preclinical models⁽⁸⁷⁾.

Polyamines are biogenic amines involved in host cell growth and differentiation and are produced by bacterial metabolism of species belonging to several genera including *Bacteroides*, *Lactobacillus*, *Veillonella*, *Bifidobacterium* and *Clostridium*. These bacteria can produce polyamines including putrescine, cadaverine, tyramine and histamine following the metabolism of amino acids including arginine, ornithine, lysine, tyrosine, histidine and methionine⁽⁸⁸⁾. Preclinical studies have shown that putrescine and spermidine in the colon are dependent on colonic microbiota⁽⁸⁹⁾ and that pectin fermentation by *B. thetaiotaomicron* and *Fusobacterium varium* stimulated polyamine production. Microbial synthesis of polyamines is considered as a therapeutic target⁽⁹⁰⁾, but there is limited information from human intervention studies on the impact of diet.

Regular consumption of cooked or processed meat can increase the risk of colon cancer and heterocyclic amines, such as 2-amino-1-methyl-6-phenylimidazo [4,5-b] pyridine, are considered to be a contributing factor. There is some evidence that the gut microbiota and, in particular, the most abundant carcinogenic heterocyclic amine, 2-amino-1-methyl-6-phenylimidazo [4,5-b] pyridine, can be transformed by representatives of the phyla Firmicutes, Bacteroidetes and Proteobacteria⁽⁹¹⁾. Similarly, the genotoxicity of mutagen 2-amino-3-methylimidazo[4,5-f]quinoline was impacted on by the gut microbiota⁽⁹²⁾.

Human dietary protein studies

High levels of proteins and peptides in the large intestine could lead to increased production of deleterious

metabolites. Magee *et al.*⁽⁹³⁾ reported that when subjects were fed a high-protein diet, the levels of sulphide were elevated due to the bacterial fermentation of sulphur-containing amino acids. Butyrate concentrations and numbers of butyrate-producing bacteria are decreased in the large intestine as well in the faeces⁽²⁰⁾. It is widely regarded that butyrate is the main energy source for colonic epithelial cell; thus, a decrease in butyrate concentration and an increase in concentrations of ammonia and sulphide may explain the detrimental effect of high protein diet on the large intestine (e.g. increased incidence of colon cancer).

Consumption of red meat is often considered to have negative health outcomes; however, it is perhaps important to take into consideration the intakes, levels of processing and other dietary factors. The consumption of high-quality red meat is poorly associated with diabetes risk and CHD for a serving of 100 g red meat daily. In contrast, higher risks were observed for processed meat consumption with an increased incidence of colorectal cancer (by 22%), heart disease (by 42%) and type 2 diabetes (by 19%)^(94,95). Moreover, there were no associations with stroke for any of the meat-type products. Processed meats tend to contain higher sodium levels which may worsen cardiovascular conditions over habitual intake. The links between red meat and poorer health outcomes may therefore be confounded by the effects of processing. Higher plant protein intake and a lower intake of some animal-based protein sources may contribute to the lower risk of disease associated with vegetarian diets. It maybe however that the benefits of high plant protein intakes are linked to other nutrients.

The protein intake of children in western countries is very high and the average protein intake in children between 4 and 6 years old is about 55 g/d. In infants where energy, protein and amino acid requirements are high, protein requirements are primarily met by intakes of human milk and infant formula. It is not clear if the protein requirements of older adults are higher than that of younger adults or are only higher in the frail elderly who are at risk of malnutrition because of acute or chronic illness.

Vegetarians exclude meat and fish from their diets and therefore there is a gradient of protein intake from meat eaters to vegans in western countries. In general, the adult population in western countries have a protein intake of about 1.3/kg/d which is about twice the estimated average requirement of 0.66 g/kg/d although a proportion of lacto-ovo-vegetarians may have protein intakes that do not meet their individual requirements.

It is often considered that amino acids may be inadequate in vegetarian diets although almost all plant-based foods contain all twenty amino acids, including the nine indispensable amino acids. The distribution profile of the amino acids however is less optimal in plant foods than in animal foods with lysine often being present in much lower than optimal proportions for human needs in grains. Also, the sulphur-containing amino acids, methionine and cysteine, are proportionally slightly lower in legumes than would be optimal for human needs. Mixing complementary protein sources

within the same meal however may simply be a practical way to secure long-term adequacy when total protein intake is low.

Meat consumers were found to have the lowest fibre intake of <10 g daily and lower PUFA consumption. Inadequacies for folate intake were also reported. However, the corresponding intake of micronutrients was found to be the highest for this group for zinc, phosphorus and vitamin B₁₂. Iron intake was found to be inadequate in the case of women. Vegetarians were found to comply with most dietary requirements and among the three groups and have a fibre intake of approximately 33% greater than meat-eaters. Vegans however were found to have a fibre intake which was 75% greater than the meat-eating group and had mineral and micronutrient intake values similar to those observed in vegetarians. For people consuming plant-based diets, further scientific evidence is required to determine if the protein intakes of vegetarian and vegan diets are adequate⁽⁹⁶⁾. The amino acid requirements are therefore considered to be adequately met for vegetarian diets although some inadequacies were observed in the Adventist study population data which may have likely risen from over dependence on a few protein sources⁽⁹⁷⁾.

Ten different diets were compared and based on average consumption of various plant and animal products consumed, in a Swiss study. Diets high in animal products were found to be detrimental to health and the environment, although this model failed to consider micronutrient intake which may affect long-term population health.

Across the Nutri-Santé and Oxford EPIC studies^(98,99), protein intake followed the expected trend of meat eaters > pescatarians > vegetarians > vegans. Furthermore, the drop in protein intake across each group was approximately 0.1 g/kg body weight (translating to about 1.2% protein with meat eaters having an intake of about 1.2 g/kg body weight). It is common in high-income countries to have diets with a protein contribution >15% in the daily energy requirement. Vegans who form the baseline for an animal-free diet were found to consume 0.99 g protein/kg body weight in previously published studies. Given the minimum protein requirement to initiate anabolism is 30 g daily for a 70 kg person (translating to about 10% of energy intake or 0.8 g/kg body weight), protein intake appears to be adequate across all diets. The average contribution to energy from protein across various food groups is shown in Fig. 2.

Food security

There is considerable interest in the impact of plant-based diets on the environment^(100–102). Current insecurities around food production stem from the inefficiencies of food distribution, poor intensification strategies, water use and waste management^(103–105). The allocation of resources towards protein production is currently centred towards animal husbandry as the relative price of animal produce is about ten times higher than most plant products. The Scottish land mass is predominantly marginal

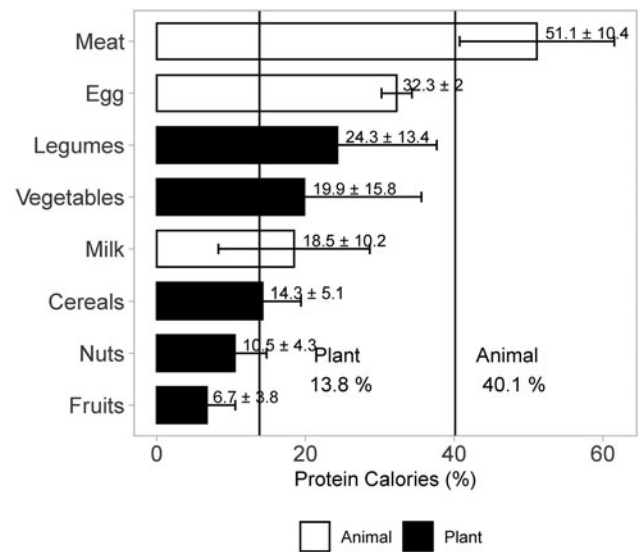


Fig. 2. Relative contribution of protein to energy for a range of food group from FAOSTAT Database (2020)⁽¹¹⁸⁾. Lines refer to the mean energy contribution of protein in plant- and animal-derived foods.

owing to its hilly terrain. Consequently, the nature of agriculture favours animal rearing rather than high-intensity cropping (Fig. 3). For sustaining a given population, about 0.2 ha of land is required per individual⁽¹⁰⁶⁾, which implies a land mass of 1.1×10^6 ha capable of high-intensity cropping. Existing capable land in Scotland is about 6×10^5 ha which is about 45% lower than required. Animal husbandry is therefore important to ensure sustained food supply. Moreover, climatic conditions make it difficult to cultivate a variety of vegetables which are often imported from countries with favourable conditions.

The emissions associated with agriculture in Scotland are shown in Fig. 4. Greenhouse gas emissions of food production as a share of anthropogenic emissions are

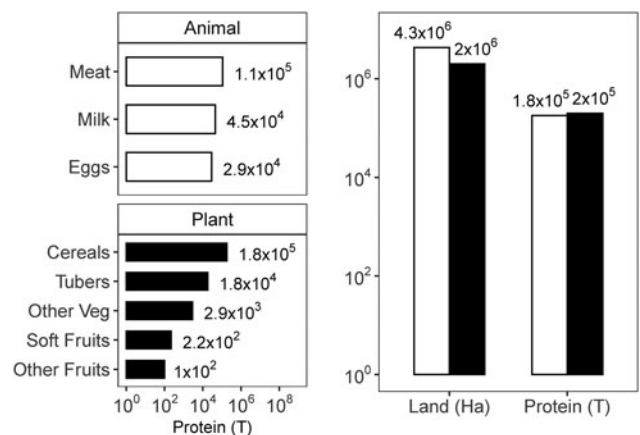


Fig. 3. Protein production from Scottish agriculture^(119,120). Left panel accounts for protein from major animal- and plant-based produce. The right panel compares the land allocation for the animal- and plant-based agriculture and the corresponding production of protein from these sources, respectively.

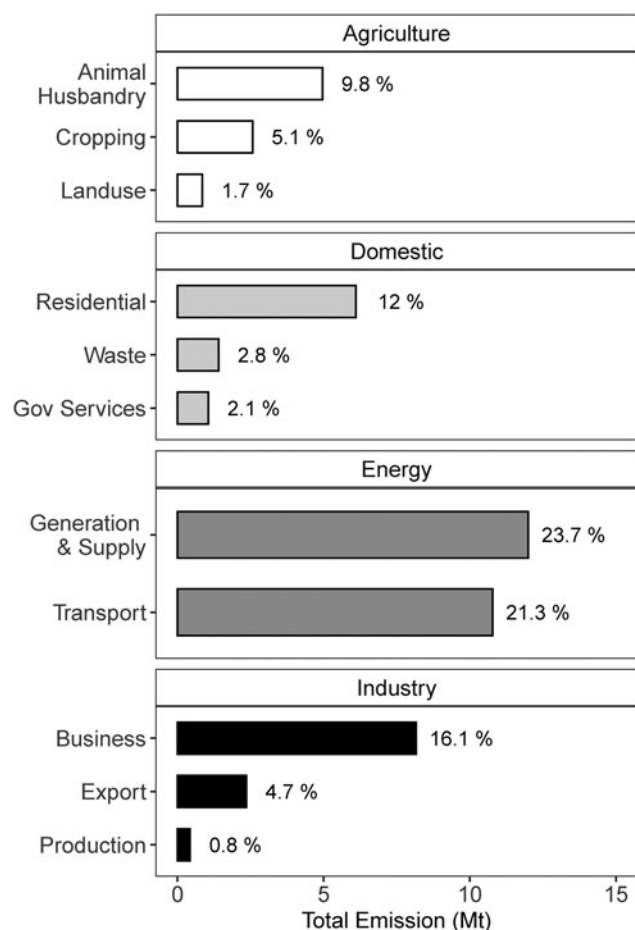


Fig. 4. Scottish anthropogenic emissions⁽¹²¹⁾.

comparable to global averages. However, satisfying the indigenous nutritional requirement is mostly dependent on meat. In terms of carbon efficiency in Scotland, 1 kg protein produced from animal sources results in an emission of 102.4 kg CO₂ eq. while plant protein results in 13.5 kg CO₂ eq. The average food prices in Scotland are relatively higher than global averages⁽¹⁰⁷⁾, diversity in diet is low, dependency on imports is high, and consequently, food security and environmental impact of food production is significant.

The WHO healthy plate guideline aims to recommend foods which meet nutritional requirements as well as ensuring low greenhouse gas emissions. These recommendations do not account for real-world wastage of food in common households, which was estimated to contribute about 9% of total household dietary emissions⁽¹⁰⁸⁾. Furthermore, the single largest contributor to dietary emissions comes from animal products and in particular red meat. In India, for example, the contribution from red meat consumption among non-vegetarians is negligible, and consequently leading to a dietary per capita emission of 757 kg CO₂ eq. per annum compared to the WHO healthy diet (1288 kg CO₂ eq.)⁽¹⁰⁹⁾. This can be explained by the unique reliance on pulses, fish and poultry to obtain nutrition which saved about 20% of emissions. Current research is aimed at meat

replacements^(110,111) and reducing enteric emissions by altering diet and gut composition of ruminants^(112,113), but sustainable long-term solutions are mostly directed towards locally sourced, low red-meat high plant diets⁽¹¹⁴⁾. Climate change affects food availability either by directly disrupting crop growth through unfavourable conditions or through altering crop quality due to increased atmospheric CO₂ levels⁽¹¹⁵⁾. Smith and Myers⁽¹¹⁶⁾ established a close linear relation between protein and mineral content in plant-based diets in low-income countries which is not observed in animal-based products where mineral composition is relatively independent of the protein content. Paradoxically, WHO-recommended diet to mitigate nutrient deficiency relies on supplementation using animal products which of course adds to emissions⁽¹¹⁷⁾.

Conclusions

Dietary protein is metabolised by proteases and peptidases in the human small intestine, and the released amino acids from dietary protein can be used for protein synthesis by gut microbes. This contributes to the nitrogen cycling and utilisation between the microbiota and host. Moreover, the undigested protein and amino acids are mainly fermented into various bacterial metabolites, such as SCFA, hydrogen sulphide, ammonia and other nitrogenous and aromatic metabolites. Some of these bacterial metabolites can be transported inside colonocytes and exert beneficial or deleterious effects. These effects might be attributed to the modulation of the intestinal barrier function and immune defence by the altered gut microbiota. Further studies will be necessary to elucidate the relationship between dietary protein and gut microbiota as well as the interaction of microbial function and host health. This becomes increasingly important with a changing agricultural landscape addressing sustainability in our food system and transition to a more circular economy.

Acknowledgements

We would like to thank Pat Bain for help in preparing Fig. 1.

Financial Support

The Rowett Institute is funded by the Scottish Government Rural and Environmental Sciences and Analytical Services (SG-RESAS).

Conflict of Interest

None.

Authorship

The authors had joint responsibility for all aspects of preparation of the paper.

References

- Walker AW, Duncan SH, Louis P *et al.* (2014) Phylogeny, culturing, and metagenomics of the human gut microbiota. *Trends Microbiol* **22**, 267–274.
- Lapébie P, Lombard V, Drula E *et al.* (2019) Bacteroidetes use thousands of enzyme combinations to break down glycans. *Nat Commun* **10**, 2043.
- Louis P & Flint HJ (2017) Formation of propionate and butyrate by the human colonic microbiota. *Environ Microbiol* **19**, 29–41.
- Scott KP, Gratz SW, Sheridan PO *et al.* (2013) The influence of diet on the gut microbiota. *Pharma Res* **69**, 52–60.
- Flint HJ, Scott KP, Louis P *et al.* (2012) The role of the gut microbiota in nutrition and health. *Nat Rev Gastroenterol Hepatol* **9**, 577–589.
- Smith P & Gregory PJ (2013) Climate change and sustainable food production. *Proc Nutr Soc* **72**, 21–28.
- Goodrich JK, Davenport ER, Beaumont M *et al.* (2016) Genetic determinants of the gut microbiome in UK twins. *Cell Host Microbe* **19**, 731–743.
- Wu GD, Chen J, Hoffmann C *et al.* (2011) Linking long-term dietary patterns with gut microbial enterotypes. *Science* **334**, 105–108.
- Wang YT, Mohammed SD, Farmer AD *et al.* (2015) Regional gastrointestinal transit and pH studied in 215 healthy volunteers using the wireless motility capsule: influence of age, gender, study country and testing protocol. *Aliment Pharmacol Ther* **42**, 761–772.
- Abuhelwa AY, Foster DJR & Upton RN (2016) A quantitative review and meta-models of the variability and factors affecting oral drug absorption – part I: gastrointestinal pH. *AAPS J* **18**, 1309–1321.
- Macfarlane S & Macfarlane GT (2003) Regulation of short-chain fatty acid production. *Proc Nutr Soc* **62**, 67–72.
- Johnson AJ, Vangay P, Al-Ghalith GA *et al.* (2019) Daily sampling reveals personalized diet-microbiome associations in humans. *Cell Host Microbe* **25**, 789–802.e5.
- Cummings JH & Macfarlane GT (1991) The control and consequences of bacterial fermentation in the human colon. *J Appl Bacteriol* **70**, 443–459.
- Columpsi P, Sacchi P, Zuccaro V *et al.* (2016) Beyond the gut bacterial microbiota: the gut virome. *J Med Virol* **88**, 1467–1472.
- Hofer U (2013) Variation in the gut virome. *Nat Rev Microbiol* **11**, 596–596.
- Gouba N, Raoult D & Drancourt M (2014) Eukaryote culturomics of the gut reveals new species. *PLoS ONE* **11**, e106994.
- Zoetendal EG, Raes J, van den Bogert B *et al.* (2012) The human small intestinal microbiota is driven by rapid uptake and conversion of simple carbohydrates. *ISME J* **6**, 1415–1426.
- Bäckhed F, Ley RE, Sonnenburg JL *et al.* (2005) Host-bacterial mutualism in the human intestine. *Science* **307**, 1915–1920.
- Qin J, Li R, Raes J *et al.* (2010) A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* **464**, 59–65.
- Duncan SH, Louis P & Flint HJ (2007) Cultivable bacterial diversity from the human colon. *Lett Appl Microbiol* **44**, 343–350.
- Derrien M, Vaughan EE, Plugge CM *et al.* (2004) *Akkermansia muciniphila* gen. nov., sp. nov., a human intestinal mucin-degrading bacterium. *Int J Syst Evol Microbiol* **54**, 1469–1476.
- Macy JM & Probst I (1979) The biology of gastrointestinal Bacteroides. *Annu Rev Microbiol* **33**, 561–594.
- Salyers AA, Vercellotti JR, West SE *et al.* (1977) Fermentation of mucin and plant polysaccharides by strains of Bacteroides from the human colon. *Appl Environ Microbiol* **33**, 319–322.
- Neish AS (2002) The gut microflora and intestinal epithelial cells: a continuing dialogue. *Microbes Infect* **4**, 309–317.
- Isolauri E, Kirjavainen PV & Salminen S (2002) Probiotics: a role in the treatment of intestinal infection and inflammation? *Gut* **50**(Suppl. 3), 54–59.
- Lopez-Siles M, Khan TM, Duncan SH *et al.* (2012) Cultured representatives of two major phylogroups of human colonic *Faecalibacterium prausnitzii* can utilize pectin, uronic acids, and host-derived substrates for growth. *Appl Environ Microbiol* **78**, 420–428.
- Marquet P, Duncan SH, Chassard C *et al.* (2009) Lactate has the potential to promote hydrogen sulphide formation in the human colon. *FEMS Microbiol Lett* **299**, 128–134.
- O'Toole PW & Jeffery IB (2015) Gut microbiota and aging. *Science* **350**, 1214–1215.
- Eckburg PB, Bik EM, Bernstein CN *et al.* (2005) Diversity of the human intestinal microbial flora. *Science* **308**, 1635–1638.
- Khan MT, Duncan SH, Stams AJM *et al.* (2012) The gut anaerobe *Faecalibacterium prausnitzii* uses an extracellular electron shuttle to grow at oxic–anoxic interphases. *ISME J* **6**, 1578–1585.
- Walker AW, Duncan SH, McWilliam Leitch EC *et al.* (2005) Ph and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. *Appl Environ Microbiol* **71**, 3692–3700.
- Flint HJ & Duncan SH (2014) *Bacteroides* and *Prevotella*. In *Encyclopedia of Food Microbiology*, 2nd ed., pp. 203–208 [CA Batt and ML Tortorello, editors]. [Internet]. Oxford: Academic Press. Available at <http://www.science-direct.com/science/article/pii/B9780123847300000318>.
- Flint HJ (2020) *Why Gut Microbes Matter: Understanding Our Microbiome*. Cham: Springer International Publishing (Fascinating Life Sciences). Available at <http://link.springer.com/10.1007/978-3-030-43246-1>.
- Ridlon JM, Harris SC, Bhowmik S *et al.* (2016) Consequences of bile salt biotransformations by intestinal bacteria. *Gut Microbes* **7**, 22–39.
- Dominguez-Bello MG, Costello EK, Contreras M *et al.* (2010) Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci* **107**, 11971–11975.
- Duncan SH & Flint HJ (2013) Probiotics and prebiotics and health in ageing populations. *Maturitas* **75**, 44–50.
- David LA, Maurice CF, Carmody RN *et al.* (2014) Diet rapidly and reproducibly alters the human gut microbiome. *Nature* **505**, 559–563.
- van Best N, Rolle-Kampczyk U, Schaap FG *et al.* (2020) Bile acids drive the newborn's gut microbiota maturation. *Nat Commun* **11**, [Epublication 23 July 2020].
- Timmerman HM, Rutten NBMM, Boekhorst J *et al.* (2017) Intestinal colonisation patterns in breastfed and formula-fed infants during the first 12 weeks of life reveal sequential microbiota signatures. *Sci Rep* **7**, [Epublication 21 August 2020].
- Vandenplas Y, Carnielli VP, Ksiazek J *et al.* (2020) Factors affecting early-life intestinal microbiota development. *Nutrition* **78**, [Epublication 25 March 2020].

41. Praveen P, Jordan F, Priami C *et al.* (2015) The role of breast-feeding in infant immune system: a systems perspective on the intestinal microbiome. *Microbiome* **3**, 41.
42. Flint HJ, Scott KP, Duncan SH *et al.* (2012) Microbial degradation of complex carbohydrates in the gut. *Gut Microbes* **3**, 289–306.
43. Leeming ER, Johnson AJ, Spector TD *et al.* (2019) Effect of diet on the gut microbiota: rethinking intervention duration. *Nutrients* **11**, [Epublication 22 November 2020].
44. Harmsen HJM, Wildeboer-Veloo ACM, Raangs GC *et al.* (2000) Analysis of intestinal flora development in breast-fed and formula-fed infants by using molecular identification and detection methods. *J Pediatr Gastroenterol Nutr* **30**, 61–67.
45. Jeffery IB, Claesson MJ, O'Toole PW *et al.* (2012) Categorization of the gut microbiota: enterotypes or gradients? *Nat Rev Microbiol* **10**, 591–592.
46. Booijink CCGM, El-Aidy S, Rajilić-Stojanović M *et al.* (2010) High temporal and inter-individual variation detected in the human ileal microbiota. *Environ Microbiol* **12**, 3213–3227.
47. Soto-Martin EC, Warnke I, Farquharson FM *et al.* (2020) Vitamin biosynthesis by human gut butyrate-producing bacteria and cross-feeding in synthetic microbial communities. *mBio* **11**, (article no. e00886-20), 1–18.
48. Ze X, Ben David Y, Laverde-Gomez JA *et al.* (2015) Unique organization of extracellular amylases into amylo-somes in the resistant starch-utilizing human colonic Firmicutes bacterium *Ruminococcus bromii*. *mBio* **6**, e01058-15.
49. Caffall KH & Mohnen D (2009) The structure, function, and biosynthesis of plant cell wall pectic polysaccharides. *Pectin Struct Funct* **344**, 1879–1900.
50. Chung WSF, Meijerink M, Zeuner B *et al.* (2017) Prebiotic potential of pectin and pectic oligosaccharides to promote anti-inflammatory commensal bacteria in the human colon. *FEMS Microbiol Ecol* **93**, [Epublication 3 October 2017].
51. Chung WSF, Walker AW, Vermeiren J *et al.* (2018) Impact of carbohydrate substrate complexity on the diversity of the human colonic microbiota. *FEMS Microbiol Ecol* **95**, [Epublication 9 October 2018].
52. Nordström C (1972) Release of enteropeptidase and other brush-border enzymes from the small intestinal wall in the rat. *Biochim Biophys Acta BBA – Enzymol* **289**, 367–377.
53. Russell WR, Hoyles L, Flint HJ *et al.* (2013) Colonic bacterial metabolites and human health. *Curr Opin Microbiol* **16**, 246–254.
54. Smith EA & Macfarlane GT (1997) Formation of phenolic and indolic compounds by anaerobic bacteria in the human large intestine. *Microb Ecol* **33**, 180–188.
55. Yao CK, Muir JG & Gibson PR (2016) Review article: insights into colonic protein fermentation, its modulation and potential health implications. *Aliment Pharmacol Ther* **43**, 181–196.
56. Johnstone AM, Horgan GW, Murison SD *et al.* (2008) Effects of a high-protein ketogenic diet on hunger, appetite, and weight loss in obese men feeding ad libitum. *Am J Clin Nutr* **87**, 44–55.
57. Kim E, Coelho D & Blachier F (2013) Review of the association between meat consumption and risk of colorectal cancer. *Nutr Res* **33**, 983–994.
58. Neis E, Dejong C & Rensen S (2015) The role of microbial amino acid metabolism in host metabolism. *Nutrients* **7**, 2930–2946.
59. Dai Z-L (2011) Amino acid metabolism in intestinal bacteria: links between gut ecology and host health. *Front Biosci* **16**, 1768–1786.
60. McIntosh FM, Shingfield KJ, Devillard E *et al.* (2009) Mechanism of conjugated linoleic acid and vaccenic acid formation in human faecal suspensions and pure cultures of intestinal bacteria. *Microbiology* **155**, 285–294.
61. Smith EA & Macfarlane GT (1997) Dissimilatory amino acid metabolism in human colonic bacteria. *Anaerobe* **3**, 327–337.
62. Davila A-M, Blachier F, Gotteland M *et al.* (2013) Intestinal luminal nitrogen metabolism: role of the gut microbiota and consequences for the host. *Pharmacol Res* **68**, 95–107.
63. Macfarlane GT, Allison C, Gibson SAW *et al.* (1988) Contribution of the microflora to proteolysis in the human large intestine. *J Appl Bacteriol* **64**, 37–46.
64. Sánchez-Jiménez F, Ruiz-Pérez MV, Urdiales JL *et al.* (2013) Pharmacological potential of biogenic amine-polyamine interactions beyond neurotransmission: amine interplay beyond neurotransmission. *Br J Pharmacol* **170**, 4–16.
65. Nissim I, Horyn O, Daikhin Y *et al.* (2014) The molecular and metabolic influence of long term arginine consumption. *J Biol Chem* **289**, 9710–9729.
66. Hughes R, Magee EAM & Bingham S (2000) Protein degradation in the large intestine. *Curr Issues Intest Microbiol* **1**, 51–58.
67. Duncan SH, Louis P & Flint HJ (2004) Lactate-utilizing bacteria, isolated from human feces, that produce butyrate as a major fermentation product. *Appl Environ Microbiol* **70**, 5810–5817.
68. Louis P, Scott KP, Duncan SH *et al.* (2007) Understanding the effects of diet on bacterial metabolism in the large intestine. *J Appl Microbiol* **102**, 1197–1208.
69. Barcenilla A, Pryde SE, Martin JC *et al.* (2000) Phylogenetic relationships of butyrate-producing bacteria from the human gut. *Appl Environ Microbiol* **66**, 1654–1661.
70. Duncan SH, Hold GL, Harmsen HJM *et al.* (2002) Growth requirements and fermentation products of *Fusobacterium prausnitzii*, and a proposal to reclassify it as *Faecalibacterium prausnitzii* gen. nov., comb. nov. *Int J Syst Evol Microbiol* **52**, 2141–2146.
71. Pryde SE, Duncan SH, Hold GL *et al.* (2002) The microbiology of butyrate formation in the human colon. *FEMS Microbiol Lett* **217**, 133–139.
72. Hamer HM, Jonkers D, Venema K *et al.* (2007) Review article: the role of butyrate on colonic function. *Aliment Pharmacol Ther* **27**, 104–119.
73. Scheppach W (1994) Effects of short chain fatty acids on gut morphology and function. *Gut* **35**(1 Suppl.), S35–S38.
74. Lewis SJ & Heaton KW (1997) Increasing butyrate concentration in the distal colon by accelerating intestinal transit. *Gut* **41**, 245–251.
75. Tolhurst G, Heffron H, Lam YS *et al.* (2012) Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. *Diabetes* **61**, 364–371.
76. Chambers ES, Viardot A, Psichas A *et al.* (2015) Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut* **64**, 1744–1754.
77. Yang J, Martínez I, Walter J *et al.* (2013) *In vitro* characterization of the impact of selected dietary fibers on fecal microbiota composition and short chain fatty acid production. *Anaerobe* **23**, 74–81.

78. Louis P, Duncan SH, McCrae SI *et al.* (2004) Restricted distribution of the butyrate kinase pathway among butyrate-producing bacteria from the human colon. *J Bacteriol* **186**, 2099–2106.
79. Bui TPN, de Vos WM & Plugge CM (2014) *Anaerostipes rhammosivorans* sp. Nov., a human intestinal, butyrate-forming bacterium. *Int J Syst Evol Microbiol* **64**, 787–793.
80. Ueki A, Goto K, Ohtaki Y *et al.* (2017) Description of *Anaerotignum aminivorans* Gen. Nov., sp. Nov., a strictly anaerobic, amino-acid-decomposing bacterium isolated from a methanogenic reactor, and reclassification of *Clostridium propionicum*, *Clostridium neopropionicum* and *Clostridium lactatifermentans* as species of the genus *Anaerotignum*. *Int J Syst Evol Microbiol* **67**, 4146–4153.
81. Attene-Ramos MS, Wagner ED, Gaskins HR *et al.* (2007) Hydrogen sulfide induces direct radical-associated DNA damage. *Mol Cancer Res* **5**, 455–459.
82. Wallace RJ (1996) Ruminal microbial metabolism of peptides and amino acids. *J Nutr* **126**(Suppl. 4), 1326S–1334S.
83. Jin Y, Singh P, Chung H-J *et al.* (2018) Blood ammonia as a possible etiological agent for Alzheimer's disease. *Nutrients* **10**, [Epublication 4 May 2018].
84. Massey RC, Key PE, Mallett AK *et al.* (1988) An investigation of the endogenous formation of apparent total N-nitroso compounds in conventional microflora and germ-free rats. *Food Chem Toxicol* **26**, 595–600.
85. Matsui M, Nagai F, Suzuki E *et al.* (1984) Structure-activity relationships of nitrosamines and nitramines which stimulate UDP-glucuronosyltransferase activities *in vitro*. *Biochem Pharmacol* **33**, 2647–2651.
86. Suzuki K & Mitsuoka T (1984) N-nitrosamine formation by intestinal bacteria. *IARC Sci Publ* **57**, 275–281.
87. Del Bo C, Bernardi S, Marino M *et al.* (2019) Systematic review on polyphenol intake and health outcomes: Is there sufficient evidence to define a health-promoting polyphenol-rich dietary pattern? *Nutrients* **11**, [Epublication 16 June 2017].
88. Zhao Z-H, Xin F-Z, Xue Y *et al.* (2019) Indole-3-propionic acid inhibits gut dysbiosis and endotoxin leakage to attenuate steatohepatitis in rats. *Exp Mol Med* **51**, 1–14.
89. Noack J, Kleessen B, Proll J *et al.* (1998) Dietary guar gum and pectin stimulate intestinal microbial polyamine synthesis in rats. *J Nutr* **128**, 1385–1391.
90. Gerner EW & Meyskens FL (2009) Combination chemoprevention for colon cancer targeting polyamine synthesis and inflammation. *Clin Cancer Res* **15**, 758–761.
91. Zhang J, Lacroix C, Wortmann E *et al.* (2019) Gut microbial beta-glucuronidase and glycerol/diol dehydratase activity contribute to dietary heterocyclic amine biotransformation. *BMC Microbiol* **19**, [Epublication 16 May 2019].
92. Kassie F (2001) Intestinal microflora plays a crucial role in the genotoxicity of the cooked food mutagen 2-amino-3-methylimidazo[4,5-f]quinoline (IQ). *Carcinogenesis* **22**, 1721–1725.
93. Magee EA, Richardson CJ, Hughes R *et al.* (2000) Contribution of dietary protein to sulfide production in the large intestine: an *in vitro* and a controlled feeding study in humans. *Am J Clin Nutr* **72**, 1488–1494.
94. Mozaffarian D (2016) Dietary and policy priorities for cardiovascular disease, diabetes and obesity. *Circulation* **133**, 187–225.
95. Micha R, Wallace SK & Mozaffarian D (2010) Red and processed meat consumption and risk of incident coronary heart disease, stroke, and diabetes mellitus: a systematic review and meta-analysis. *Circulation* **121**, 2271–2283.
96. Agnoli C, Baroni L, Bertini I *et al.* (2017) Position paper on vegetarian diets from the working group of the Italian society of human nutrition. *Nutr Metab Cardiovasc Dis* **27**, 1037–1052.
97. Rizzo NS, Sabate J, Jaceldo-Siegl K *et al.* (2011) Vegetarian dietary patterns are associated with a lower risk of metabolic syndrome: the Adventist health study 2. *Diabetes Care* **34**, 1225–1227.
98. Mariotti F & Gardner CD (2019) Dietary protein and amino acids in vegetarian diets – a review. *Nutrients* **11**, 1–19.
99. Gluba-Brzózka A, Franczyk B & Rysz J (2017) Vegetarian diet in chronic kidney disease – a friend or foe. *Nutrients* **9**, [Epublication 10 April 2017].
100. Sabaté J, Sranacharoenpong K, Harwatt H *et al.* (2015) The environmental cost of protein food choices. *Public Health Nutr* **18**, 2067–2073.
101. Hedenus F, Wirsenius S & Johansson DJA (2014) The importance of reduced meat and dairy consumption for meeting stringent climate change targets. *Clim Change* **124**, 79–91.
102. Rööös E, Bajželj B, Smith P *et al.* (2017) Protein futures for Western Europe: potential land use and climate impacts in 2050. *Reg Environ Change* **17**, 367–377.
103. Fitton N, Alexander P, Arnell N *et al.* (2019) The vulnerabilities of agricultural land and food production to future water scarcity. *Glob Environ Change* **58**, 101944.
104. Brears RC (2015) The circular economy and the water-food nexus. *Future Food: J Food Agric Soc* **4**, 73–74.
105. Mueller ND, Gerber JS, Johnston M *et al.* (2012) Closing yield gaps through nutrient and water management. *Nature* **490**, 54–57.
106. Myers N (2000) The new millennium: an ecology and an economy of hope. *Curr Sci* **78**, 686–693.
107. Barton KL, Wrieden WL, Armstrong J *et al.* (2011) Energy density of the Scottish diet: analysis from the expenditure and food survey. *Proc Nutr Soc* **70**, E26.
108. Xue L, Liu G, Parfitt J *et al.* (2017) Missing food, missing data? A critical review of global food losses and food waste data. *Environ Sci Technol* **51**, 6618–6633.
109. Ritchie H, Reay DS & Higgins P (2018) Beyond calories: a holistic assessment of the global food system. *Front Sustain Food Syst* **2**, [Epublication 14 September 2018].
110. Joshi V & Kumar S (2015) Meat analogues: plant based alternatives to meat products – a review. *Int J Food Ferment Technol* **5**, 107.
111. Multari S, Stewart D & Russell WR (2015) Potential of fava bean as future protein supply to partially replace meat intake in the human diet. *Compr Rev Food Sci Food Saf* **14**, 511–522.
112. Basarab JA, Beauchemin KA, Baron VS *et al.* (2013) Reducing GHG emissions through genetic improvement for feed efficiency: effects on economically important traits and enteric methane production. *Animal: Int J Anim Biosci* **7**(Suppl. 2), 303–315.
113. Matthews C, Crispie F, Lewis E *et al.* (2019) The rumen microbiome: a crucial consideration when optimising milk and meat production and nitrogen utilisation efficiency. *Gut Microbes* **10**, 115–132.
114. Chen C, Chaudhary A & Mathys A (2019) Dietary change scenarios and implications for environmental, nutrition, human health and economic dimensions of food sustainability. *Nutrients* **11**, [Epublication 16 April 2020].



115. Fernando N, Panozzo J, Tausz M *et al.* (2012) Rising atmospheric CO₂ concentration affects mineral nutrient and protein concentration of wheat grain. *Food Chem* **133**, 1307–1311.
116. Smith MR & Myers SS (2018) Impact of anthropogenic CO₂ emissions on global human nutrition. *Nat Clim Change* **8**, 834–839.
117. Ritchie H, Reay DS & Higgins P (2018) The impact of global dietary guidelines on climate change. *Glob Environ Change* **49**, 46–55.
118. Gennari P, Seid Y, Sorrenti S *et al.* (2020) FAOSTAT. Available at <http://www.fao.org/faostat/en/#home>.
119. Food Policy and Food Science Service (2016) Amino acid content of food and biological data on proteins. Food and Nutrition Series – Collection FAO. Available at <http://www.fao.org/3/ac854t/AC854T00.htm>.
120. UK Government (2018) Agriculture in the United Kingdom data sets. Available at <https://www.gov.uk/government/statistical-data-sets/agriculture-in-the-united-kingdom>.
121. Scottish Government (2020) *Greenhouse Gas Emissions 2018: Estimates*. Scottish Government Statistics. Available at <https://www.gov.scot/publications/scottish-greenhouse-gas-emissions-2018/>.