Effects of 1α -hydroxylated metabolites of cholecalciferol on intestinal radiocalcium absorption in goats

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1. Intestinal absorption of 47 Ca was measured by a double-isotope technique in goats treated with 1, 5 or 25 μ g of 1,25-dihydroxycholecalciferol (1,25(OH)₂D₃). The effects of giving 1,25(OH)₂D₃ by intravenous (iv) infusion for 30–36 h were compared at each dose level with the effects obtained by oral administration of 1,25(OH)₂D₃ either in ethanol or protected against rumen degradation in fatty acid pellets.

2. Dose-dependent increments in absorption followed the treatments, with a doubling of absorption at the 1 μ g dose and three- to fivefold increases with the 5 and 25 μ g doses. ⁴Ca absorption was equally stimulated 2 and 6 d after treatment but had returned to pretreatment levels 12–14 d after treatment.

3. Intravenous and protected oral administration of $1,25(OH)_2D_3$ stimulated ⁴⁷Ca absorption to the same extent, in spite of two- to fivefold higher plasma concentrations of $1,25(OH)_2D_3$ after iv treatment. Somewhat lower increments in ⁴⁷Ca absorption were seen using ethanol as the vehicle for oral administration.

4. The naturally occurring metabolites 1,24(R),25-trihydroxycholecalciferol and 1,25(S),26-trihydroxycholecalciferol had only one-tenth to one-fifteenth the potency of $1,25(OH)_2D_3$ in stimulating ⁴⁷Ca absorption, while synthetic 1α -hydroxycholecalciferol appeared to be twice as effective as $1,25(OH)_2D_3$ when tested at a high $(10 \ \mu g)$ dose.

Adaptation of intestinal calcium absorption has long since been recognized as an integral part of Ca homeostasis (Nicolaysen *et al.* 1953). Such adaptations are vitamin D dependent and mediated by changes in the renal production of 1,25-dihydroxycholecalciferol $(1,25(OH)_2D_3)$. $1,25(OH)_2D_3$ is thought to be the most active metabolite of cholecalciferol and acts as a regulating endocrine signal for intestinal Ca transport (Holick *et al.* 1971; Norman *et al.* 1971; Omdahl *et al.* 1971).

In most mammals studied, changes in Ca absorption can be detected shortly after exogenous administration of $1,25(OH)_2D_3$. Information from ruminants on the effects of this steroid hormone on Ca absorption seems to be lacking. Abdel-Hafeez *et al.* (1982) reported increased absorption of Ca in a sheep 24–48 h after perfusion of an isolated segment of intestine with 1α -hydroxycholecalciferol ($1(OH)D_3$). Hove (1984) found the double-isotope technique for measurement of intestinal radiocalcium absorption useful for studies of rapid changes in Ca absorption in goats. In the present study, this technique was used to compare the effects of various doses and methods of administration of $1,25(OH)_2D_3$. The activities of $1(OH)D_3$ and two naturally occurring trihydroxylated metabolites of cholecalciferol in promoting Ca absorption were also measured.

Cholecalciferol appears to be sensitive to rumen degradation when incubated with rumen liquid in an unprotected form (Sommerfeldt *et al.* 1980). The extent to which this also applies to metabolites of cholecalciferol is unknown. Consequently, the benefits obtained by protecting $1,25(OH)_2D_3$ against rumen degradation during experiments on oral milk fever prophylaxis (Hove & Kristiansen, 1982) could be questioned. A comparison of the effects of oral administration of $1,25(OH)_2D_3$ incorporated in fatty acid pellets (protected) or given in ethanol (unprotected) was therefore included in the present study.

EXPERIMENTAL

Animals and diet

Six female goats aged 2-5 years were used for the studies, which lasted for approximately 1 year. Abomasal cannulas were inserted by standard surgical procedures 1 month before measurements were started. The goats were kept in metabolism cages except for a grazing period of about 3 months. A basal low-Ca diet consisting of (g/d): barley 300, hay 200, extracted soya-bean meal 25, Ca 0.7 and phosphorus 1.6 was given and supplied sufficient protein and energy for maintenance. Sodium chloride and a micromineral-vitamin mixture, providing approximately 1 μ g cholecalciferol, were given daily. In preparation for measurements of radiocalcium absorption, animals were given extra Ca (4 g/d) as calcium acetate in the drinking-water or as an abomasal infusion. The supplementation usually lasted for 10 d before Ca absorption studies were carried out in order to obtain a low initial rate of Ca absorption before administration of the metabolites of cholecalciferol. Abomasal infusions were given for the last 48 h of this 10 d period and discontinued 4 h before the oral tracer was given. Ca supplements were not given until 2 weeks after treatment which resulted in highly active absorption of Ca, in order to reduce the risk of serious hypercalcaemia. Details of the procedures for the measurements of radiocalcium absorption and calculation of the results were given earlier (Hove, 1984).

Treatments with metabolites of cholecalciferol

Synthetic 1,25(OH)₂D₃ (lot no. 7975-119), 1,24(R),25-trihydroxycholecalciferol (1,24-(R),25(OH)₃D₃, lot no. 7978-102) and 1,25(S),26-trihydroxycholecalciferol (1,25- $(S),26(OH)_{3}D_{3}$, lot no. 8625-128) were obtained from Hoffman la Roche Inc, Nutley New Jersey, and 1α -hydroxycholecalciferol was obtained from Leo Pharmaceuticals, Copenhagen, Denmark. Three methods of administration of 1,25(OH)₂D₃ were compared. Intravenous (iv) administration was given as a 30-36 h continuous infusion (1.2 ml/h) of a solution of the metabolite in ethanol (500 ml/l). The metabolite for oral administration was either given unprotected against rumen degradation (dissolved in 10 ml ethanol) or protected (incorporated in fatty acid pellets designed for rumen bypass; Rumen Kjemi, A/S, Oslo). The fatty acid pellets contained $10 \,\mu g \, 1.25(OH)_2 D_3/g$ and the average pellet size was 40 mg. The matrix consisted of (g/kg): 540 C_{16} - C_{20} fatty acids, 370 glucose, 60 calcium carbonate, 30 adjuvants (emulgator, antioxidant). The actual extent of protection against rumen degradation obtained for $1,25(OH)_2D_3$ was not tested, but fatty acid pellets give a 70–80% bypass of amino acids and probably an even higher extent of bypass of fat-soluble vitamins. The effects of 1,25(OH)₂D₃ were studied at dose levels of 1, 5 and 25 μ g. At least four experiments were performed with each dose level and each method of administration, except for the 1 μ g dose where the protected metabolite was not given.

Measurements of radiocalcium absorption were done immediately before administration of metabolite and 2, 6 and, in several experiments, 12-14 d later. Additional studies of the effects of daily repeated doses of 5 µg protected $1,25(OH)_2D_3$ were carried out in three animals. Measurements were undertaken after a daily dose of 5 µg for 2 d (10 µg total) and after a daily dose of 5 µg for 5 d (25 µg total). The effects of 10 µg 1(OH)D₃ were studied 2 d after iv and oral (unprotected) administration (three goats per treatment). The effects of 50 µg of each of the two trihydroxylated metabolites were studied 2 d after iv administration (two goats per treatment).

Measurements of $1,25(OH)_2D_3$ in plasma

 $1,25(OH)_2D_3$ was extracted from plasma by diethyl ether. The extracts were purified by open column silica chromatography and by high-pressure liquid chromatography (Aksnes, 1980)

on a silica column (Chrompack Partisil, 5μ particles; Chrompack, Middelburg, The Netherlands). The metabolite was quantified by a radioimmunoassay (Clemens *et al.* 1979) using standard $1,25(OH)_2D_3$, sheep anti- $1,25(OH)_2D_3$ antibody (Sheep 02282) and $[^3H]1,25(OH)_2D_3$ (about 90 Ci/mmol).

RESULTS

Treatments with $1,25(OH)_2D_3$

The mean radiocalcium absorption for all goats measured just before $1,25(OH)_2D_3$ treatment (day 0) was 15 (sD 7)% of the administered dose. Absolute values for day 0, and for days 2, 6 and 12–14 after treatment are given in Figs. 1 and 2. All treatments resulted in increased radiocalcium absorption on day 2 and about the same level of absorption was found 4 d later irrespective of dose or type of administration (Figs. 1 and 2). The stimulation of radiocalcium absorption by $1,25(OH)_2D_3$ resulted in doubling of the initial values with the 1 μ g dose and up to two- to fourfold the initial values at the highest dose given (Figs. 1 and 2).

Radiocalcium absorptions 12–14 d after treatment were close to the values obtained at day 0 for both the 1 and 5 μ g doses (Fig. 1).

Increments in radiocalcium absorption were calculated in order to compare doses and types of administration of $1,25(OH)_2D_3$, since significant differences in the rate of absorption between individual goats were detected at day 0. The increments were linearly



Fig. 1. Absorption of ⁴⁷Ca (% of dose) measured in goats before and 2, 6 and 12 d after treatment with (*a*) 5 μ g 1,25-dihydroxycholecalciferol (1,25(OH)₂D₃) and (*b*) 1 μ g 1,25(OH)₂D₃. Mean values, with their standard errors represented by vertical bars, for three to five animals per treatment. 1,25(OH)₂D₃ given intravenously (\triangle), or orally, dissolved in ethanol (\bigcirc) or in pellets of fatty acids (\spadesuit).

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Fig. 2. Absorption of ⁴⁷Ca (% of dose) measured in goats before and 2 and 6 d after treatment with 25 μ g 1,25-dihydroxycholecalciferol (1,25(OH)₂D₃). Mean values, with their standard errors represented by vertical bars, for four animals per treatment. 1,25(OH)₂D₃ given intravenously (Δ), or orally, dissolved in ethanol (\bigcirc) or in pellets of fatty acids (\oplus).

related to the logarithm of the administered dose (Fig. 3). Linear regression equations were calculated for increments in radiocalcium absorption (y) v. log_e 1,25(OH)₂D₃ dose (x):

iv administration, y = 0.10 + 0.11 x, protected oral administration, y = 0.05 + 0.11 x, unprotected oral administration, y = 0.16 + 0.05 x.

The corresponding regression coefficients were all different from zero (P < 0.001, P < 0.01and P < 0.02 respectively, t test). Significant differences between coefficients were obtained for the iv and oral unprotected treatments with $1,25(OH)_2D_3$ (P < 0.02, t test), while the difference between protected and unprotected oral treatments approached significance (0.1 > P > 0.05). Differences between the three methods of administration were not significant when the 1 and 5 µg dose levels were judged separately (Fig. 3). The difference in increments between iv and unprotected oral treatments was, however, significant at the 25 µg dose level (P < 0.01, t test).

A daily dose of $5 \mu g$ protected $1,25(OH)_2D_3$ for 5 d resulted in an increase in Ca absorption from 15.6 to 69.2% (average of three goats). This increment of 53.6% units corresponded to an increment of 42.0% units when the full dose $(25 \mu g)$ was given at once (Fig. 3). An increment of 38.6% units was seen when $5 \mu g$ was given for two consecutive days. This increment corresponded to a dose of about $20 \mu g$ protected $1,25(OH)_2D_3$ (Fig. 3). Changes in plasma $1,25(OH)_2D_3$ concentrations after treatments with 1, 5 and $25 \mu g 1,25(OH)_2D_3$ were clearly related to the dose given both for the iv and oral treatments (Table 1). Plasma $1,25(OH)_2D_3$ concentrations were generally two to six times



Fig. 3. Increments in ⁴⁷Ca absorption (mean values with their standard errors) in goats after treatments with 1, 5 or 25 μ g of 1,25-dihydroxycholecalciferol (1,25(OH)₂D₃). Administration of the cholecalciferol metabolite by intravenous infusion for 30–36 h (\triangle), or orally, dissolved in ethanol (\bigcirc) or in pellets of fatty acids (\bigcirc).

higher after iv administration than after oral administration. No clearcut differences between concentrations after treatments with oral protected and unprotected metabolites could be detected (Table 1).

	Method of administration	Plasma 1,25-dihydroxyvitamin D (pg/ml)			
Dere		Before treatment		2 d after treatment	
(µg)		Mean	SE	Mean	SE
 25	iv	53	7	554	25
	op	58	7	184	13
	oup	54	14	214	37
5	iv	40	8	247	8
	ор	38	7	124	40
	oup	64	15	89	7
1	iv	49	5	89	4
	oup	44	15	76	14

(Mean values with their standard errors for three to four goats per treatment)

of 1, 5 and 25 µg 1,25-dihydroxycholecalciferol

Table 1. Plasma concentrations of 1,25-dihydroxyvitamin D (pg/ml) before and 2 d after the start of treatment with intravenous (iv), oral protected (op) and oral, unprotected (oup) doses

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Table 2. Effects of $1(OH)D_3$, $1,24(R),25(OH)_3D_3$ and $1,25(S),26(OH)_3D_3$ on radiocalcium absorption in goats. The metabolites of vitamin D_3 were either given by an intravenous (iv) infusion of 30 h duration or orally dissolved in 10 ml ethanol (500 ml/l). Radiocalcium absorption (% of dose absorbed in 180 min; mean values and ranges) was measured just before and 2 d after the start of treatment

	Dose (µg)	Method of administration	No. of animals	Radiocalcium absorption			
				Before treatment		2 d after treatment	
Vitamin D ₃ metabolite				Mean	Range	Mean	Range
1(OH)D,	10	iv	3	29	(26-35)	75	(70-80)
1(OH)D,	10	oral	3	16	(13–18)	52	(45-58)
1,24(R),25(OH),D ₃	50	iv	2	14	(12 - 15)	37	(36-39)
1,25(S),26(OH) ₃ D ₃	50	iv	2	20	(16-23)	43	(41-46)

 $1(OH)D_3 = 1\alpha$ -hydroxycholecalciferol; 1,24(R), 25(OH)₃D₃ = 1,24(R),25-trihydroxycholecalciferol; 1,25(S),26 (OH)₃D₃ = 1,25(S),26-trihydroxycholecalciferol.

Treatments with $1(OH)D_3$, $1,24(R),25(OH)_3D_3$ and $1,25(S),26(OH)_3D_3$

 $1(OH)D_3$ and the two trihydroxylated metabolites increased radiocalcium absorption, with $1(OH)D_3$ the most potent metabolite (Table 2). Absorption increased on average 46% units after iv administration of a 10 µg dose and 36% units after oral administration of unprotected $1(OH)D_3$. These increments were of the same order as increments obtained with $20-25 \mu g 1,25(OH)_2D_3$ (Fig. 3). Both $1,24(R),25(OH)_3D_3$ and $1,25(S),26(OH)_3D_3$ increased radiocalcium absorption by an average of 24% units when given intravenously in a dose of 50 µg (Table 2). The potency of these two metabolites thus seemed equivalent to approximately $3-4 \mu g$ of $1,25(OH)_2D_3(Fig. 3)$.

DISCUSSION

Dose-dependent increases in radiocalcium absorption were detected after treatment with $1,25(OH)_2D_3$. Giving a high-Ca diet to adult non-lactating goats with low Ca requirements prior to the measurements resulted in a low initial rate of Ca absorption. This allowed observations of increments in absorption also at the lowest dose of $1,25(OH)_2D_3$ used in the present study (1 μ g equivalent to 20–25 ng/kg body-weight). The fact that an increased rate of radiocalcium absorption was observed in the present study both 2 and 6 d after a single treatment with the 1 μ g dose of $1,25(OH)_2D_3$ showed that this low dose was well above the threshold for stimulation of Ca absorption. Most trials in humans with exogenous $1,25(OH)_2D_3$ have utilized doses of the metabolite comparable to the 1 μ g dose level in the present study. This dose is close to the estimated rate of $1,25(OH)_2D_3$ production in normal man (140–680 ng/d; Mawer *et al.* 1976; Norman, 1979) and, on a body-weight basis, considered to be near physiological levels in the rat (Rizzoli *et al.* 1977; Bonjour *et al.* 1978; Lee *et al.* 1981). The sensitivity of intestinal Ca absorption in the goat to exogenous $1,25(OH)_2D_3$ thus seems to be similar to the sensitivity found in both rat and man.

Increasing the dose of $1,25(OH)_2D_3$ above the $1 \mu g$ level resulted in gradually lower increments in Ca absorption, as emphasized by the logarithmic relationship between the dose of $1,25(OH)_2D_3$ and the increments in Ca absorption (Fig. 3). An efficiency in Ca absorption of 40-50% is commonly thought to represent the maximum which can be

achieved by dietary means in mature ruminants. Absorptions in this range were seen 2 d after administration of $5 \mu g 1,25(OH)_2D_3$, while the 25 μg dose resulted in even higher values. Individual absorption values well above 70% were observed. Single doses of $1,25(OH)_2D_3$ in the range of 5–25 $\mu g (100-500 \text{ ng/kg body-weight})$ thus appeared to induce a near maximal stimulation of the intestinal Ca-transport system. The time during which the intestinal mucosal cells were exposed to increased concentrations of $1,25(OH)_2D_3$ clearly influenced the adaptation in Ca absorption, as evidenced by the cumulative effect of repeated daily doses of 5 $\mu g 1,25(OH)_2D_3$.

Plasma concentrations of $1,25(OH)_2D_3$ varied according to the dose and the method of administration. Greatly elevated levels were seen after the 25 μ g dose, in accordance with the activated Ca absorption. Attention should be drawn to the fact that the concentrations of $1,25(OH)_2D_3$ (200–500 pg/ml plasma) resulting from doses which gave rise to nearly maximal stimulation of Ca absorption, were in the same range as reported in cows with parturient hypocalcaemia (Horst et al. 1977). Plasma 1,25(OH)₂D₃ concentrations were only measured once in the present study. From experiments with cows (Hove et al. 1983) it may be inferred that plasma $1,25(OH)_{a}D_{a}$ concentrations in the present experiments would have returned to pretreatment levels 3-4 d after treatment. The persistence of stimulated Ca absorption beyond the 6th day after treatment may indicate that the biological action of $1,25(OH)_2D_3$ is related to the life span of the epithelial cells of the intestinal mucosa. In agreement with recent results in the cow (Hove et al. 1983), plasma concentrations were two to five times higher after iv than after oral treatment (Table 1). Oral administration was, nevertheless, as effective in promoting Ca absorption as iv administration, except when $25 \,\mu g$ was given unprotected. A logical explanation of these observations would be that the metabolite did expose the intestinal absorptive cells to high local concentrations at the time of absorption. This could in turn give rise to much higher rates of Ca absorption than those which would be expected from the ensuing plasma $1,25(OH)_2D_3$ concentration. Some loss of $1,25(OH)_2D_3$ was indicated especially at the 25 μ g dose level when the steroid was given unprotected orally, since the mean increment in Ca absorption was lower than after protected oral administration. This was, however, mainly due to the effects of different starting points, since mean values for Ca absorption 2 d after treatment were nearly identical (Fig. 2). It may be concluded that protection of cholecalciferol metabolites against rumen degradation is of limited value when Ca absorption is to be enhanced by oral treatment in ruminants.

A 10 μ g dose of 1(OH)D₃ had a pronounced effect on radiocalcium absorption in the goats irrespective of the method of administration (Table 2). The fact that a 10 μ g dose of 1(OH)D₃ gave increments in absorption equivalent to about 20 μ g of 1,25(OH)₂D₃ seems surprising since 1(OH)D₃ is generally thought to be about half as active as 1,25(OH)₂D₃. The explanation may, however, be that the elimination of a clearly supraphysiological dose of 1,25(OH)₂D₃ occurs much more rapidly than elimination of the 1,25(OH)₂D₃ formed gradually from injected 1(OH)D₃ as shown in the cow (Hove *et al.* 1983). Braithwaite (1978, 1980) observed maximal stimulation of Ca absorption in sheep already at a daily dose of 20 ng/kg, which would be equivalent to the 1 μ g dose in the present study. The apparent discrepancy in sensitivity to 1(OH)D₃ in the sheep and the goat can probably be explained by the pronounced cumulative effect of repeated doses of 1 α -hydroxylated cholecalciferol metabolities, since the sheep were treated for 10 d before measurements were completed.

The trihydroxylated metabolites used in the present study occur in very low concentrations relative to $1,25(OH)_2D_3$ (Holick *et al.* 1973; Kleiner-Bossaler & DeLuca, 1974; Reinhardt *et al.* 1981). The potency of $1,24(R),25(OH)_3D_3$ and $1,25(S),26(OH)_3D_3$ was only about one-tenth to one-fifteenth of the potency of $1,25(OH)_2D_3$ in stimulating Ca absorption in goats. In accordance with this finding, Hove *et al.* (1983) showed $1,25(OH)_2D_3$ to be

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approximately ten times as effective as the two trihydroxylated metabolites in promoting hypercalcaemia in cows. Binding of $1,25(OH)_2D_3$ to a specific receptor is recognized as a necessary step in the mediation of the metabolic effects of this hormone. Interestingly, the affinity of the two trihydroxylated compounds for intestinal cytosol receptors was about one-tenth the affinity of $1,25(OH)_2D_3$ itself (Kream *et al.* 1977; Reinhardt *et al.* 1981).

It seems likely, therefore, that the stimulation of Ca absorption induced in the goats by the trihydroxylated metabolites was effected through the intestinal $1,25(OH)_2D_3$ receptor. It cannot be ruled out, however, that changes in $1,25(OH)_2D_3$ metabolism and plasma concentrations resulting from exogenous $1,24(R),25(OH)_3D_3$ and $1,25(S),26(OH)_3D_3$ might influence calcium absorption (Horst *et al.* 1983). With the low activity of the two trihydroxylated metabolites any role in regulation of Ca absorption in the normal state can be ruled out.

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